

# Epigenome editing ; application of genome editing tools

2016.06.09  
M1 Yamaji Kyohei

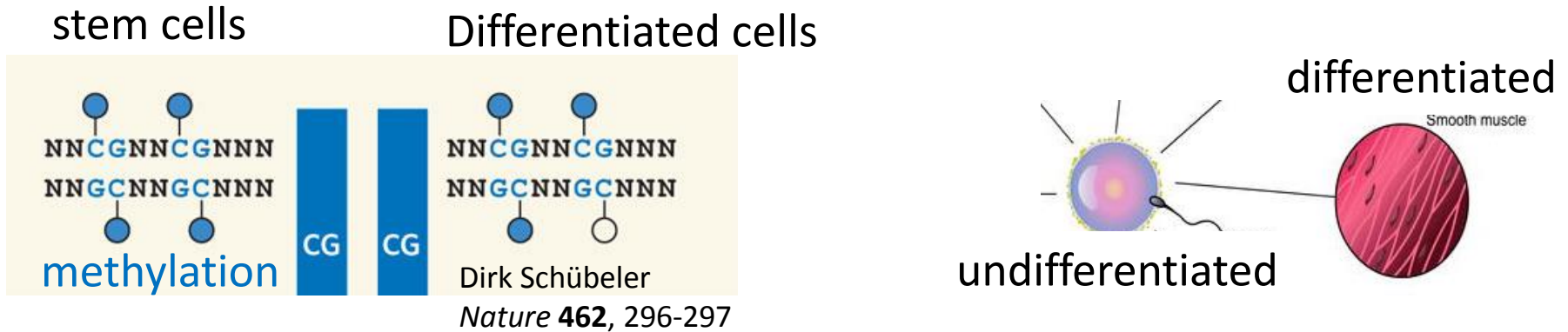
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1. Genome Targeting in Catalysis Medicine
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# Epigenetics ; chemical modifications

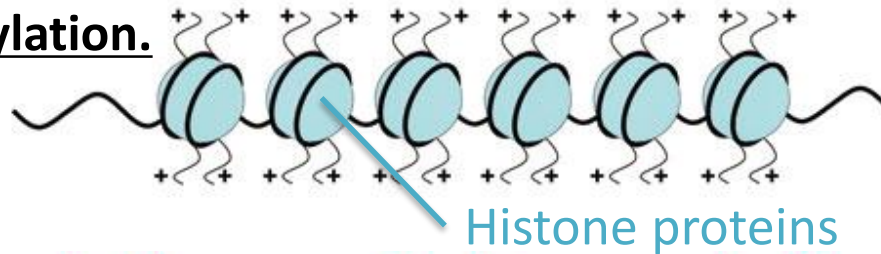
**DNA/Histone modifications can control its gene expression patterns.**

**DNA methylation pattern is different between stem cells and differentiated cells**



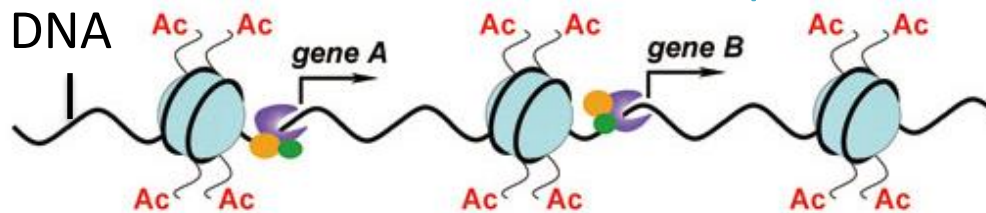
**Expression of some genes is suppressed in differentiated cells with reduced global Histone acetylation.**

Differentiated cells



~~Expression of gene A and B~~

stem cells

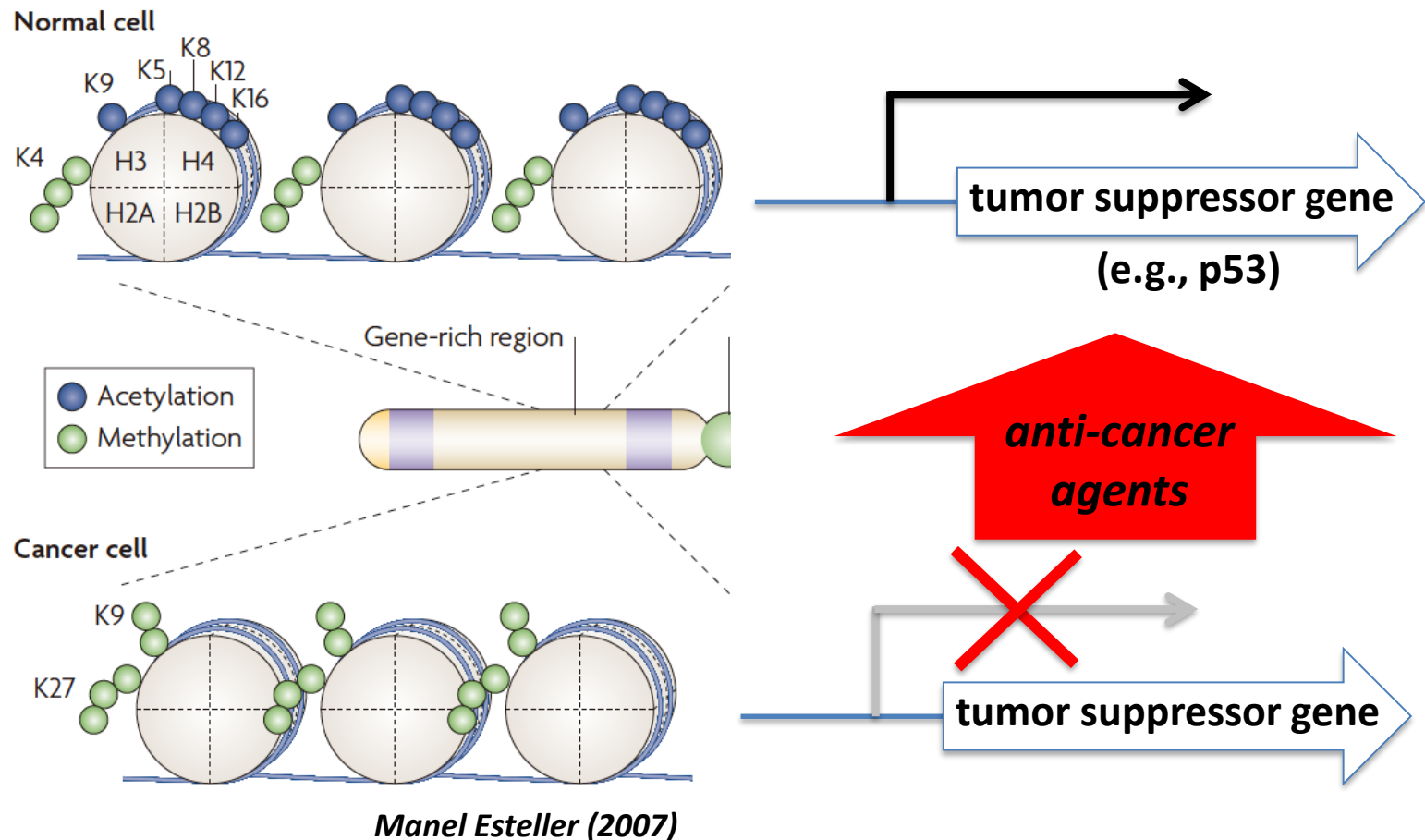


Expression of gene A and B<sub>3</sub>

# Catalysis medicine

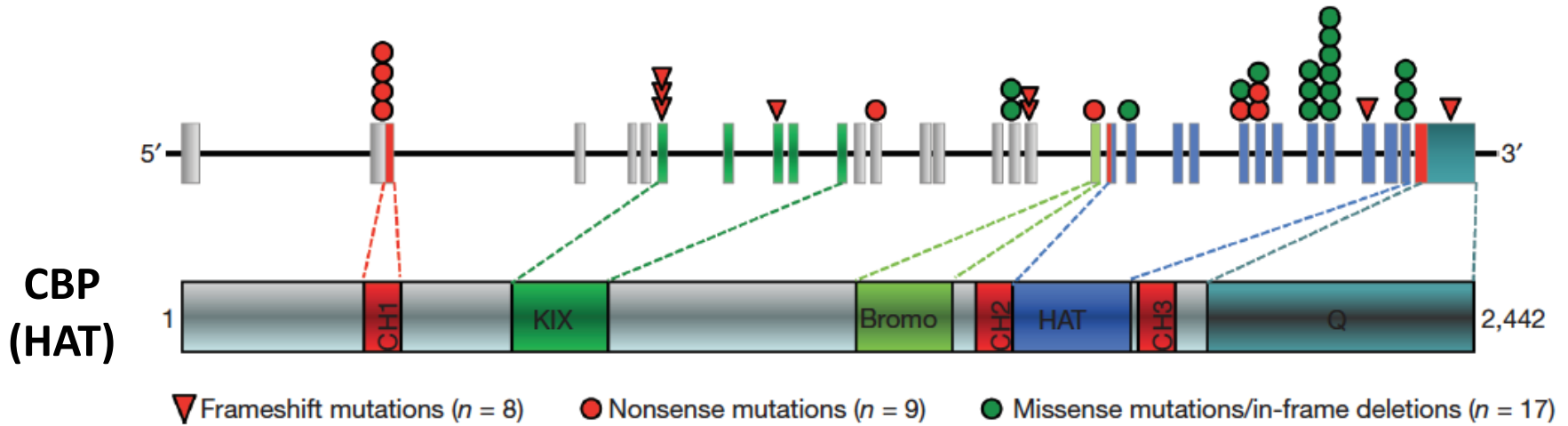
In some cancer cells, tumor suppressor genes are suppressed with decreased Histone acetylation.

Inducing Histone acetylation can be a hopeful anti-cancer strategy.



# Cataysis medicine

Mutations of HAT are frequently found in B-cell lymphoma.



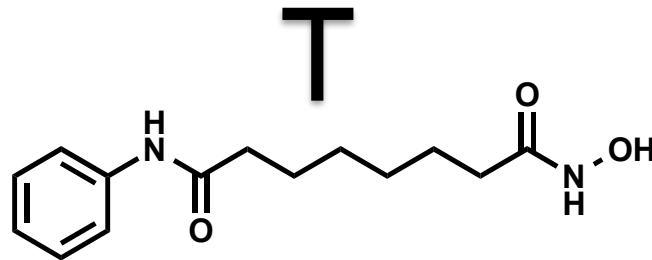
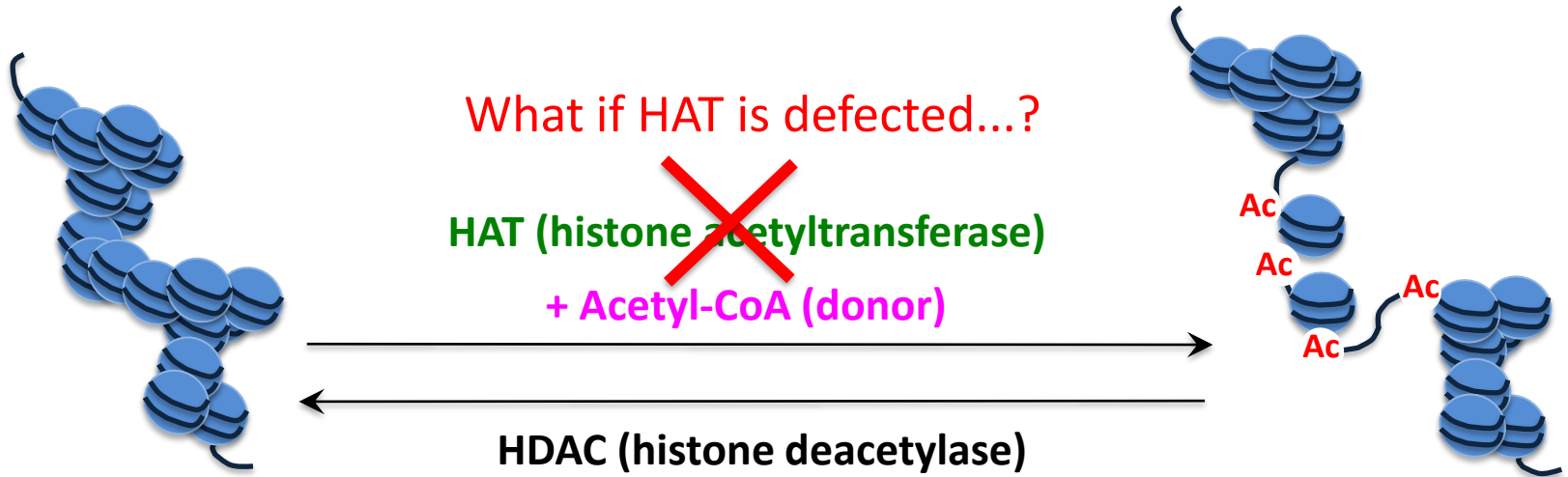
*Inactivating mutations of acetyltransferase genes in B-cell lymphoma*

Pasqualucci, L. *et al. Nature* 2011, 471, 189.

HAT-independent Histone acetylation can be the solution?

# Cataysis medicine

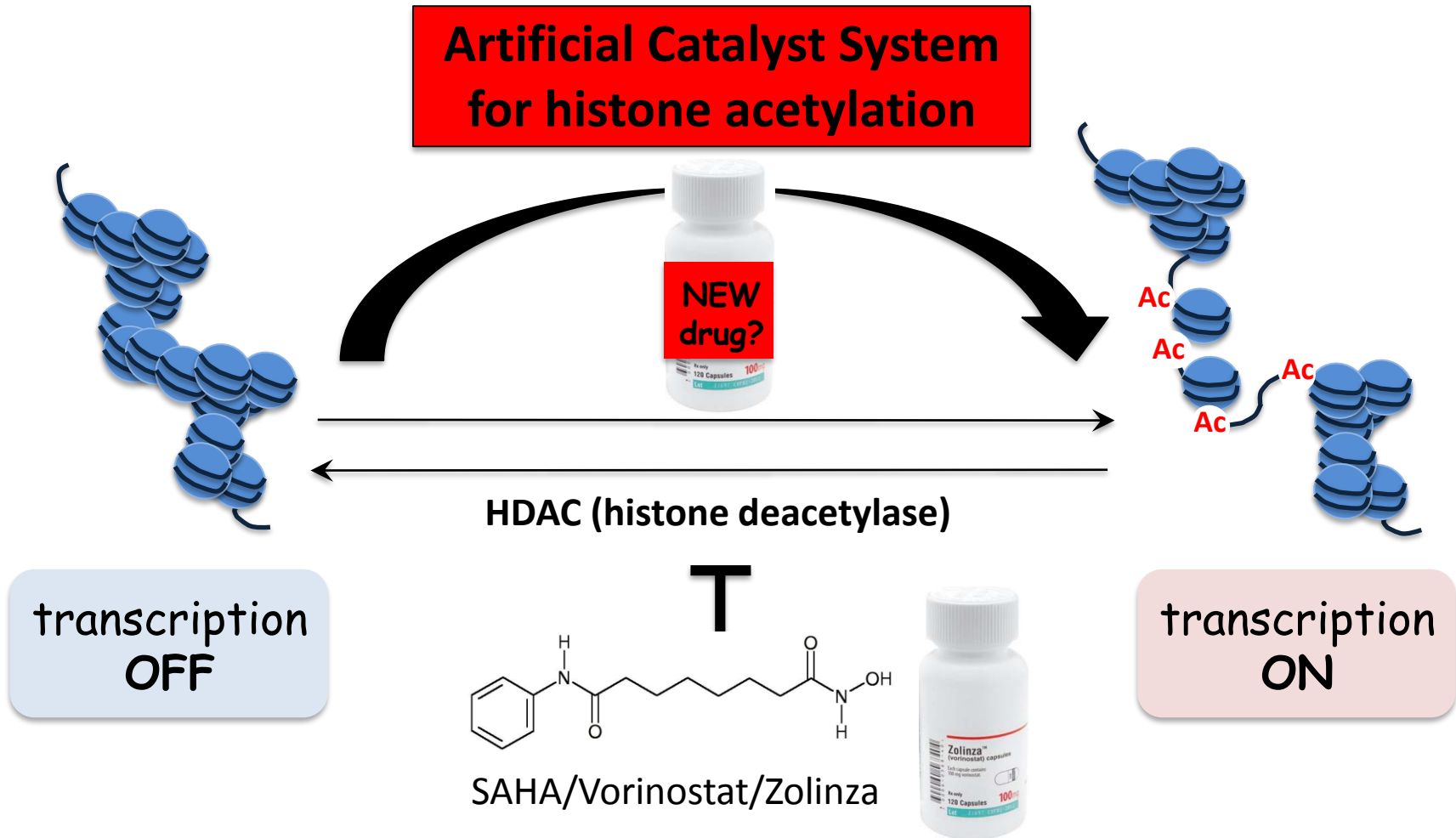
Histone acetylation level is regulated on the balance of two catalysts called HAT and HDAC.



SAHA/Vorinostat/Zolinza

HDAC inhibitor is actually used as an anti-cancer drug.

# HAT-independent histone acetylation by artificial catalyst system

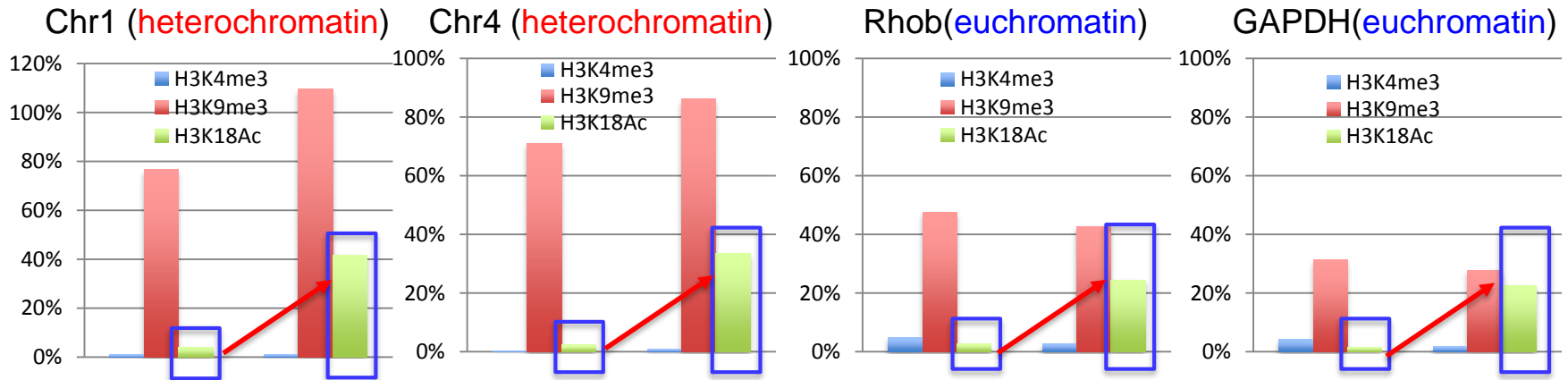


# Current catalysis medicine

## SynCac acetylate genome-wide histones nonspecifically

The result of *In nucleo* ChIP (No compound vs SynCac) by Dr.Ishiguro

•HeLa nuclei were acetylated with 50 $\mu$ M 8DMAP, 10 $\mu$ M 3NMD-8R @25 $^{\circ}$ C for 3hrs in SynCac.  
(8DMAP : batch-C by Dr. Amamoto, 3MMD-8R : KY2457 by Dr.Yamatsugu)



Both hetero/euchromatin are targets of SynCac.  
(heterochromatin acetylation might be stronger)

**Without targeting, genome-wide histone acetylation is observed**



# Gene targeting and catalysis medicine

## The usefulness of Gene targeting of artificial catalyst

- medical application
- applications as biological tools

## Applications of artificial catalyst as biological tools

- roles of **various acylations** on histone
  - roles of modifications on **various residues** of histone
- internal controls are easily obtained in targeting of catalyst

For further applications...

- **genome-wide screening** (acetylation)

And etc...

Gene targeting can expand the possibility of artificial catalyst.

# Contents

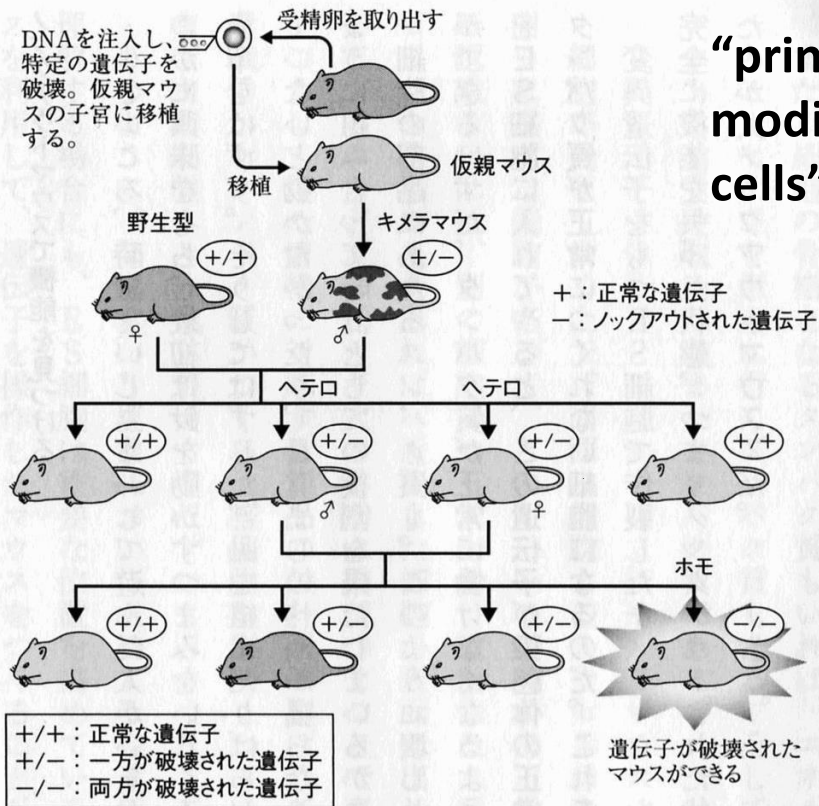
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# The utility of Genome editing

## Genome editing

Genetic engineering in which DNA is inserted, deleted, or replaced in the genome of an organism using engineered nucleases, or "molecular scissors." (wikipedia)

### General procedures for producing knockout/knockin mice



The 2007 Nobel Prize in Physiology or Medicine  
“principles for introducing specific gene modifications by the use of embryonic stem cells” (established : 1989)

“the beginning of new era in genetics”

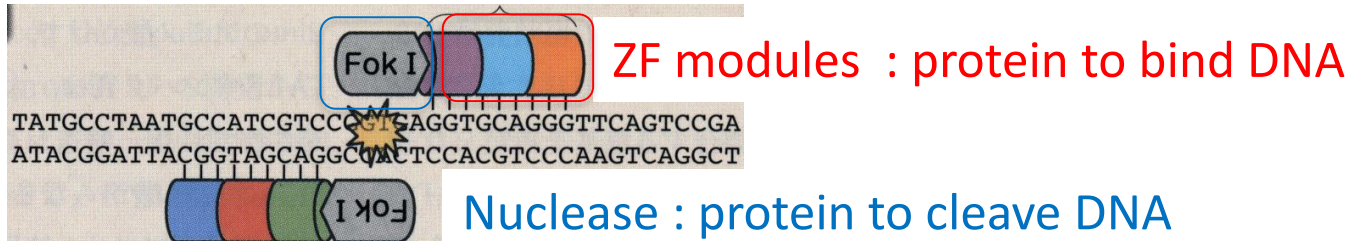
However...

Takes so much time and trouble

Genome editing tools have made this drastically easier

# Genome editing tools

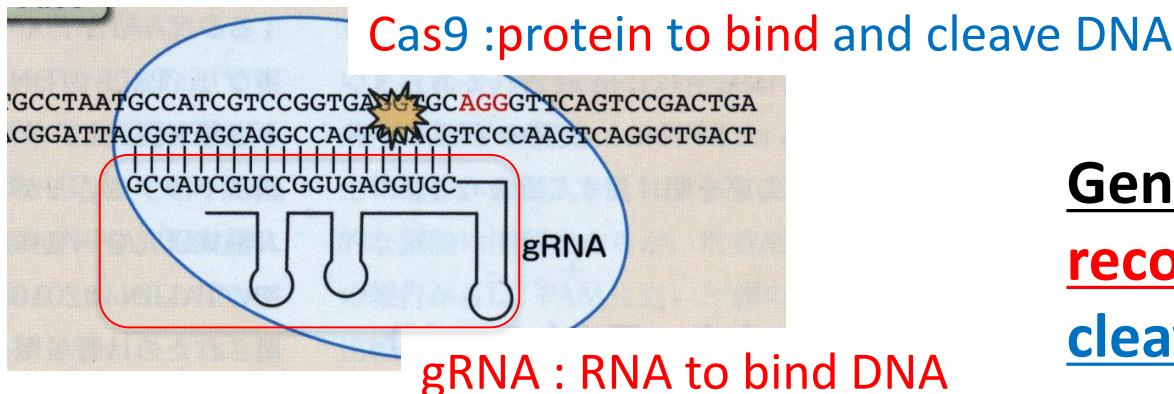
## 1<sup>st</sup> generation : ZFN (zinc finger nuclease)



## 2<sup>nd</sup> generation : TALEN (transcription-activator-like effector nuclease)



## 3<sup>rd</sup> generation : CRISPR/Cas9

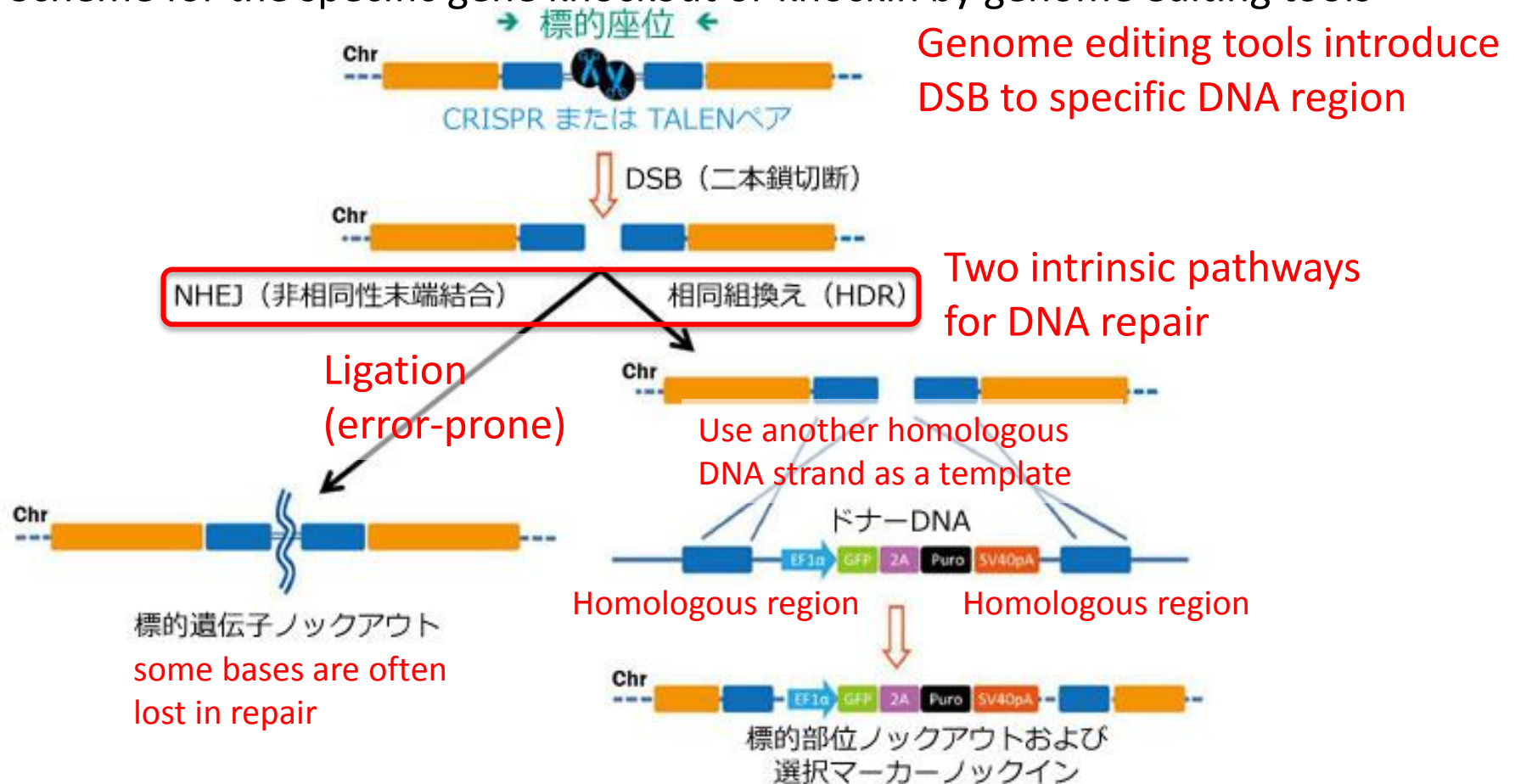


Genome editing tools...  
recognize specific sequences  
cleave DNA strand

# Principle of genome editing

## genome editing tools promote DNA repair dependent mutations to specific DNA regions

Scheme for the specific gene knockout or knockin by genome editing tools



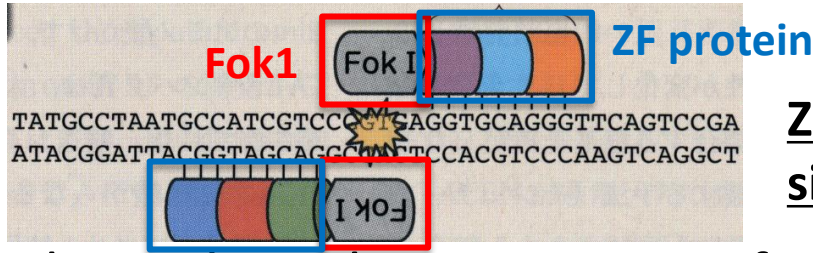
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# Fok1 endonuclease

ZFN consists of **Fok1 endonuclease cleavage domain** and **ZF protein**.

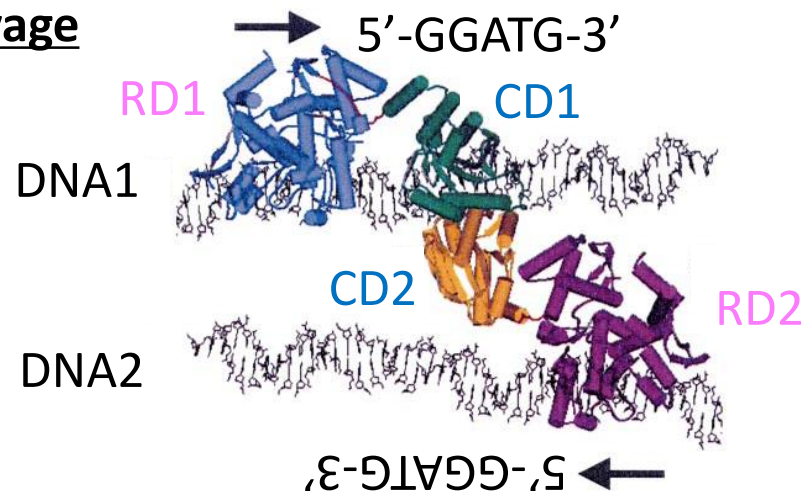
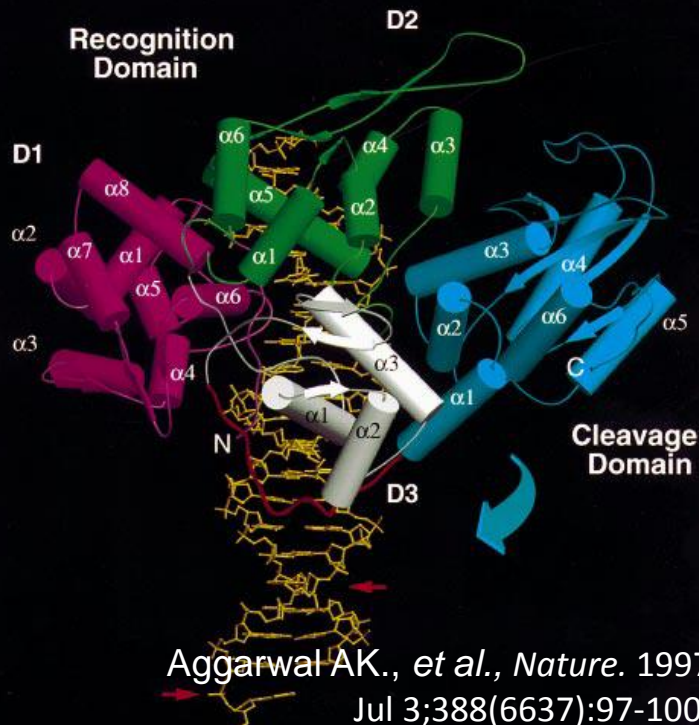


ZFN needs to be designed to put the cleavage site in between

Fok1 endonuclease consists of **Recognition domain (RD)** and **Cleavage domain (CD)**

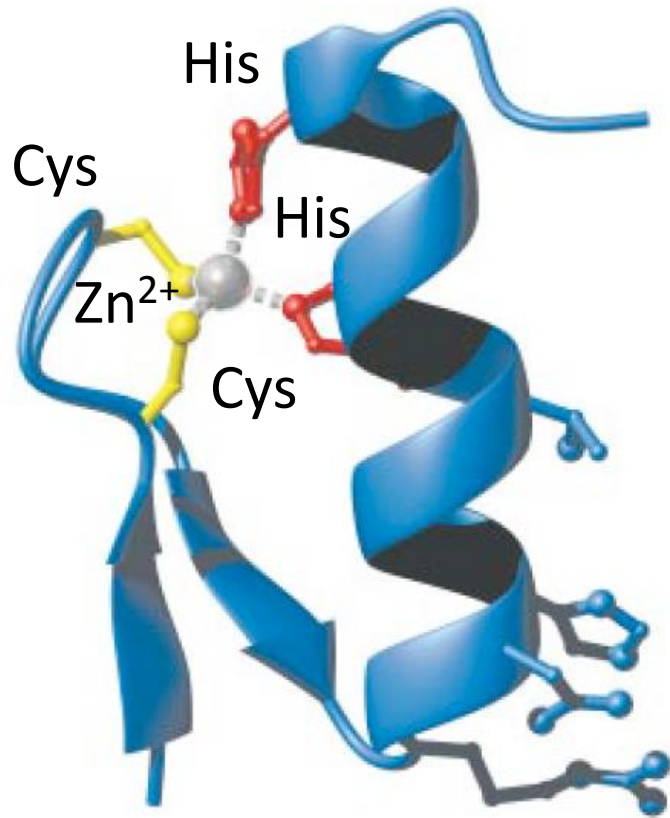
**Recognition domain** binds 5'-GGATG-3'  
 (natural Fok1 is sequence specific)

Cleavage domain dimerization is necessary for the cleavage



# structure of zinc finger protein

**ZF motif contains Zn, which interacts with His2Cys2 and stabilizes  $\beta\beta\alpha$**   
 Structure of finger 2 from Zif286, a kind of ZF motifs



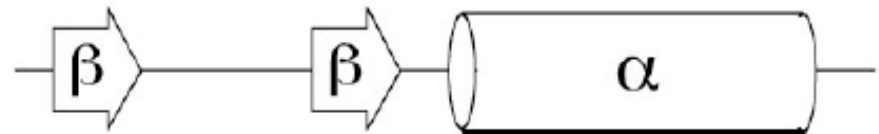
finger

			-1	2	3	6	3-5 amino acids																										
			↓	↓	↓	↓																											
1	M	E	R	P	Y	A	C	P	V	E	S	C	D	R	R	F	S	R	S	D	E	L	T	R	H	I	R	I	H	T	G	Q	K
2	P	F	Q	C	R	I	--	C	M	R	N	F	S	R	S	D	H	L	T	H	I	R	T	H	T	G	E	K					
3	P	F	A	C	D	I	--	C	G	R	K	F	A	R	S	D	E	R	K	R	H	T	K	I	H	L	R	Q	K	D			

2 or 4 amino acids

hydrophobic

6  
3  
2  
-1



**Potential DNA binding residues**

(numbered from the start of  $\alpha$  helix)

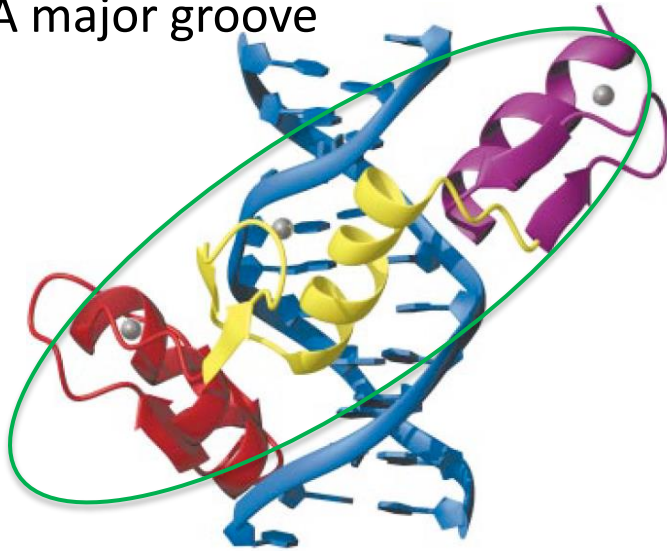
Residues closest to DNA binds DNA  
 (distance depends on the structure of ZF)



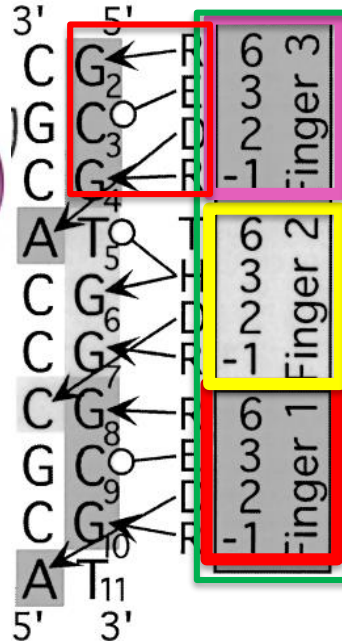
# DNA recognition by zinc finger protein

## How ZF motifs recognize DNA

ZF motifs interact with DNA major groove



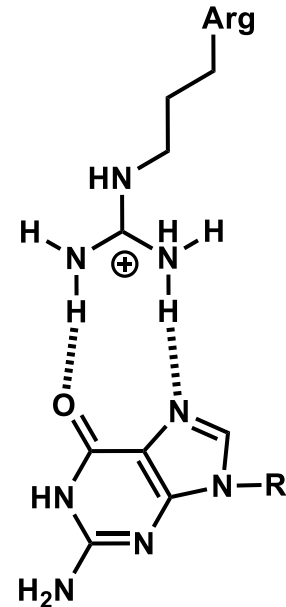
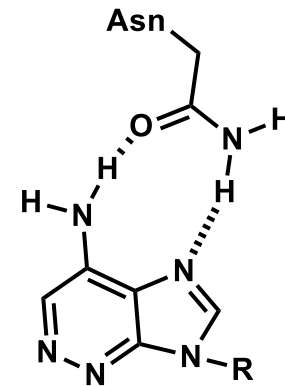
For sufficient sequence specificity, 3-6 motifs are usually combined.



One residue recognizes one nucleobase

Arginine-guanine

Asparagine-adenine



Pabo CO., et al., *Annu. Rev. Biophys. Biomol. Struct.* 1999. 3:183–212

Pabo CO., et al., *Annu. Rev. Biophys. Biomol. Struct.* 2001. 70:313–40

# Stabilizing residues of zinc finger-DNA complex

## Other interactions between ZF motifs and DNA

- His** (N $\delta$ )-phosphate of the DNA backbone (5'- primary strand)
- Lys or Arg**-phosphate of the DNA backbone (the same phosphate as a. or its 3'-strand)
- Lys** in the linker (TGE**K**P)-phosphate of the DNA backbone (5'- primary strand)

## The linker (TGEKP) contributes to the stability of ZF-DNA complex

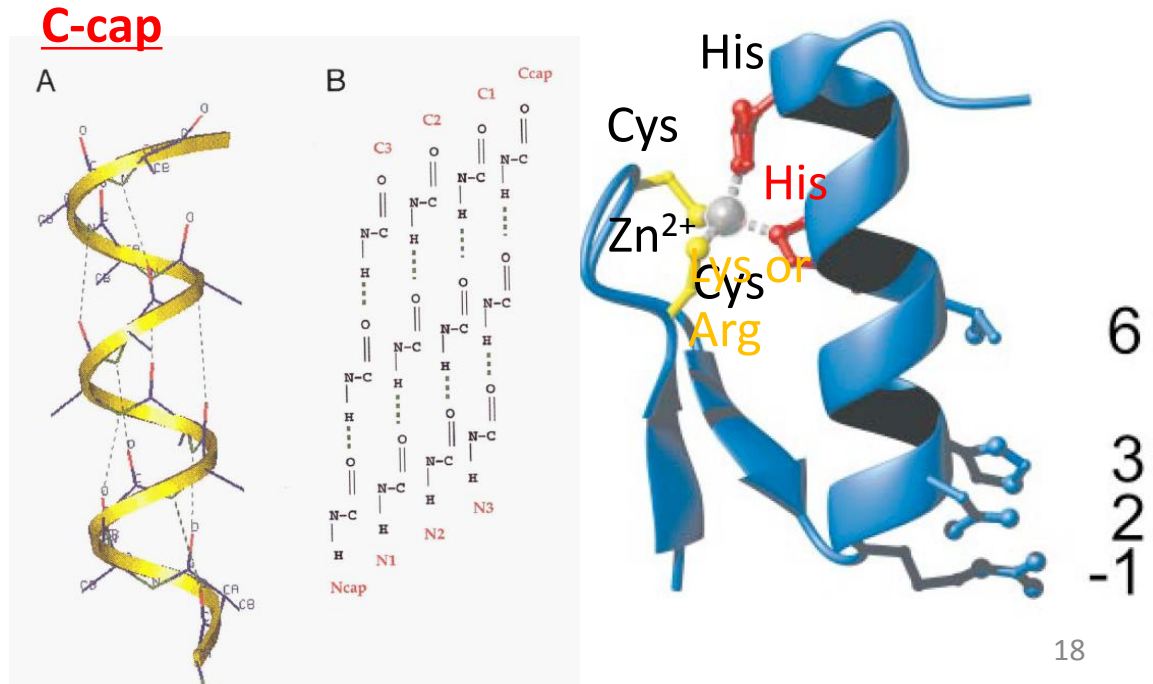
**T** : make hydrogen bond with  
NH in the next Helix (**C-cap**)

**G** : complete the **C-cap**

**E** : **stabilize** finger-finger  
interaction (contact R of  
HXRXH in the next finger)

**K** : contact **DNA backbone**

**P** :  $\pi$ - $\pi$  **stacking** with -1 residue  
in the next finger

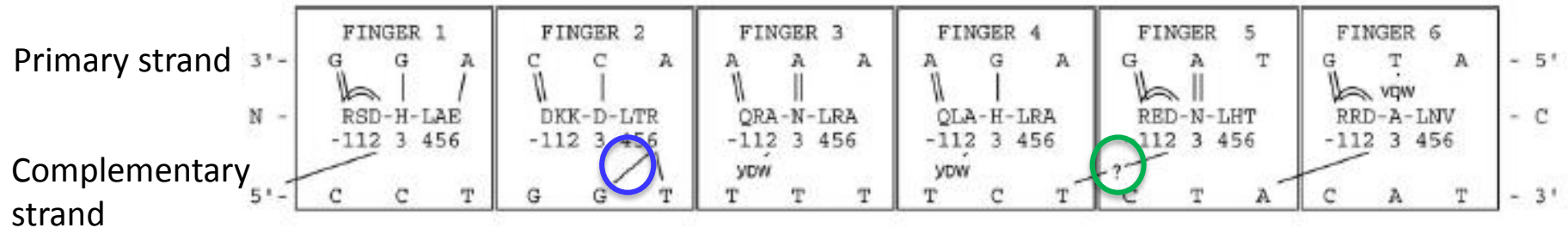


# Drawback of zinc finger protein

## drawback of ZFN

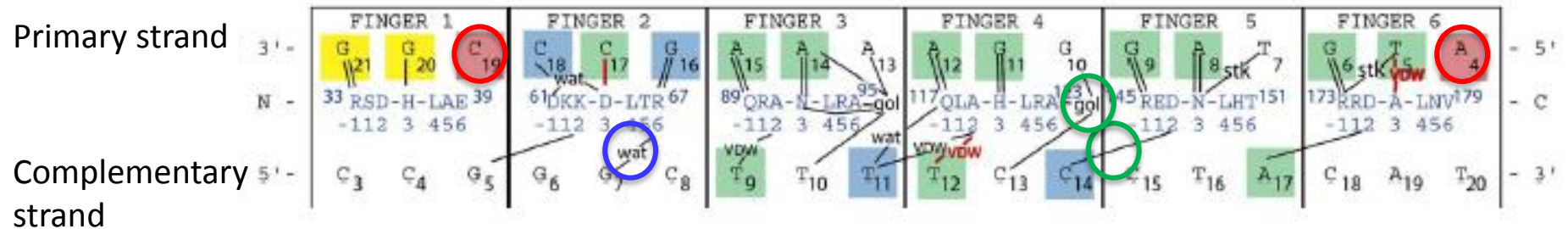
Assembled motifs often fail to recognize predicted sequence

Predicted sequence : GGA-CCA-AAA-AGA-GAT-GTA



actual sequence : GGC-CCG-AAA-AGG-GAT-GTA

Weak specificity

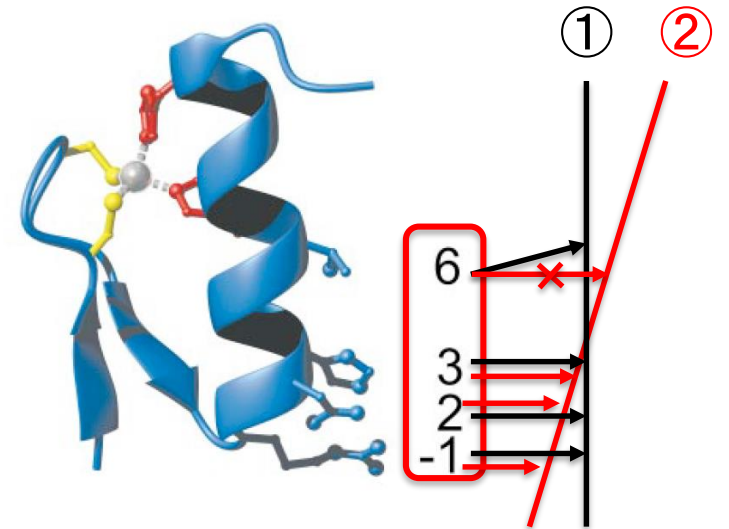
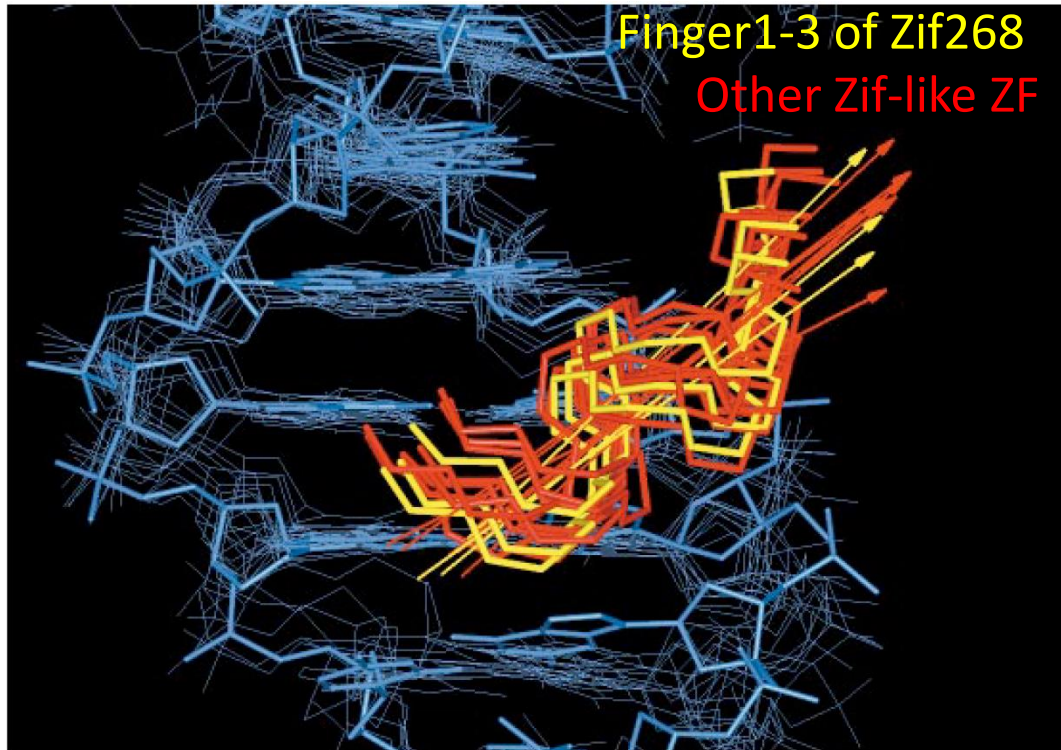


Unexplained specificity, changed interaction by water, changed interaction by glycerol(gol, necessary for crystallization, substitution of water?)

# Structural insight behind the DNA recognition by ZF

## reason for drawback of ZFN

The broad range of variation in the docking arrangement of ZF-DNA complex



The change in docking arrangement sometimes leads to the loss of base recognition by some residues

**Modular assembly may change the docking arrangement**

Pabo CO., et al., *Annu. Rev. Biophys. Biomol. Struct.* 2001. 70:313–40

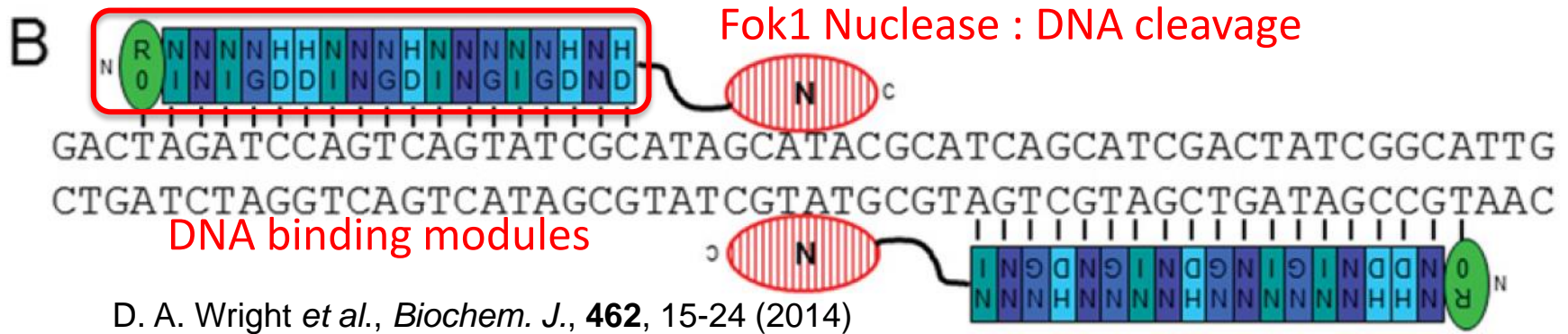
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# Overlook on TALEN

**TALEN consists of Fok1 nuclease domain and TALE (DNA binding protein)**



TALEN needs to be designed to put the cleavage site in between like ZFN

**TALEN has overcome ZFN in terms of its easiness in designing and specificity**

Predicted sequence is (almost) always recognized. (drawback of ZFN is solved)

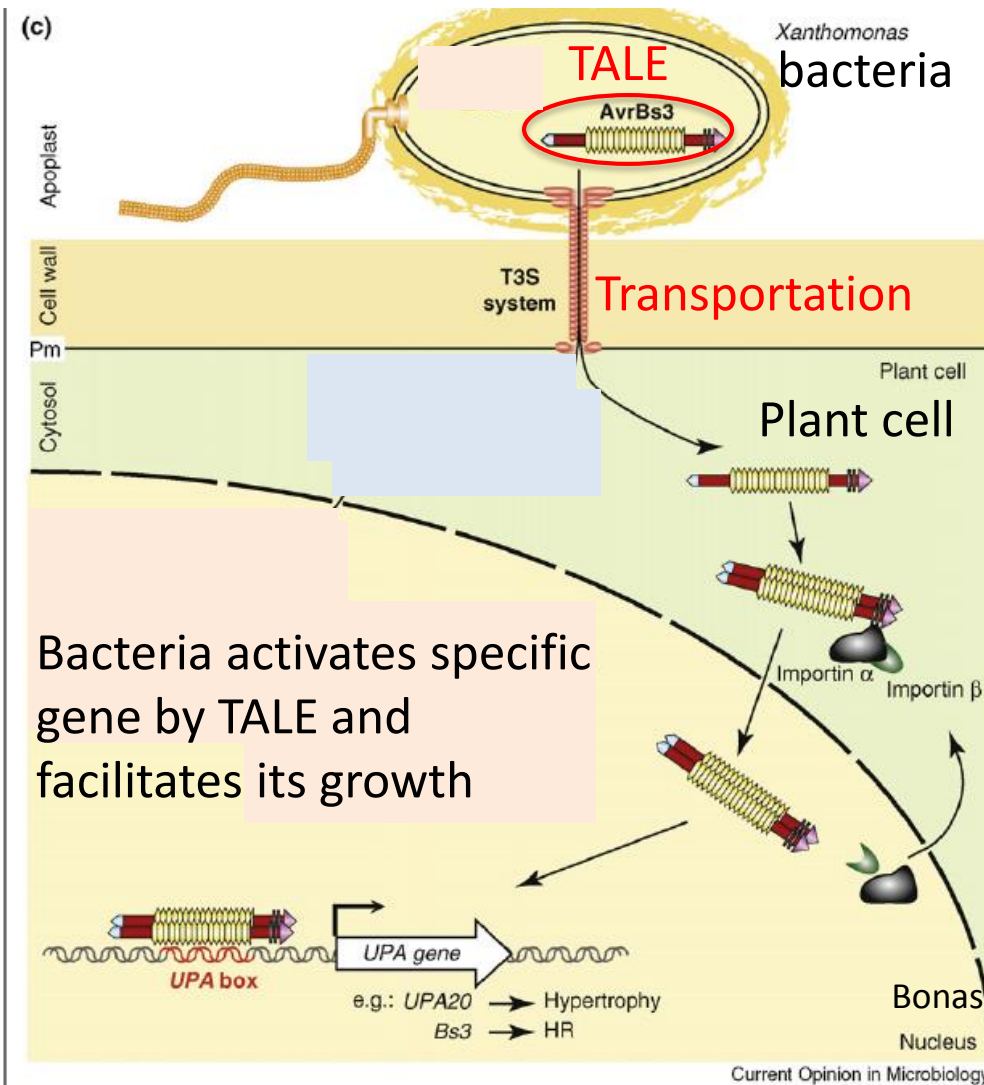
Only restraint is that the target sequence has to start from thymine

High specificity (many more bases can be recognized than the other two tools)

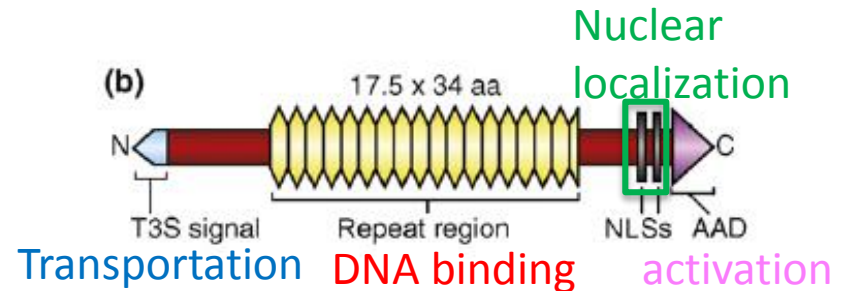
Easy to design, usable to various sequences, and high accuracy of targeting

# TALE in natural world

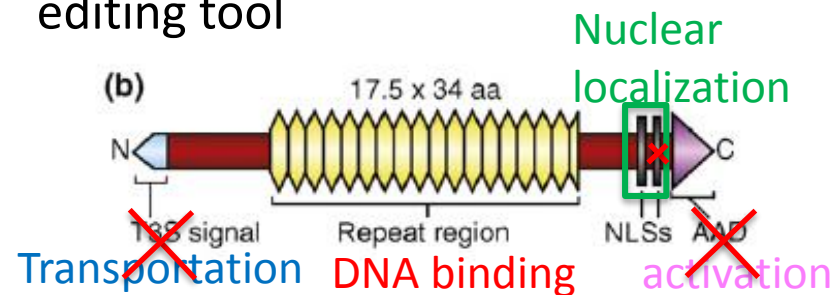
## TALE (Transcription-activator like effector) in plant infectious bacteria



Schematic view of domains in TALE



TALE in application as a genome editing tool

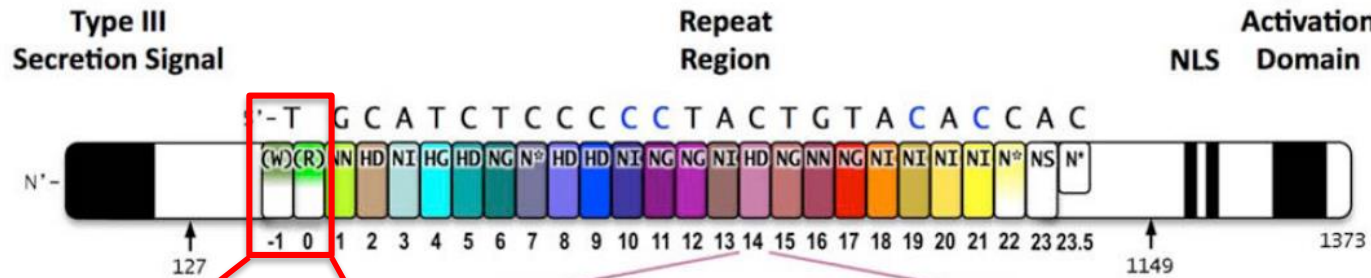
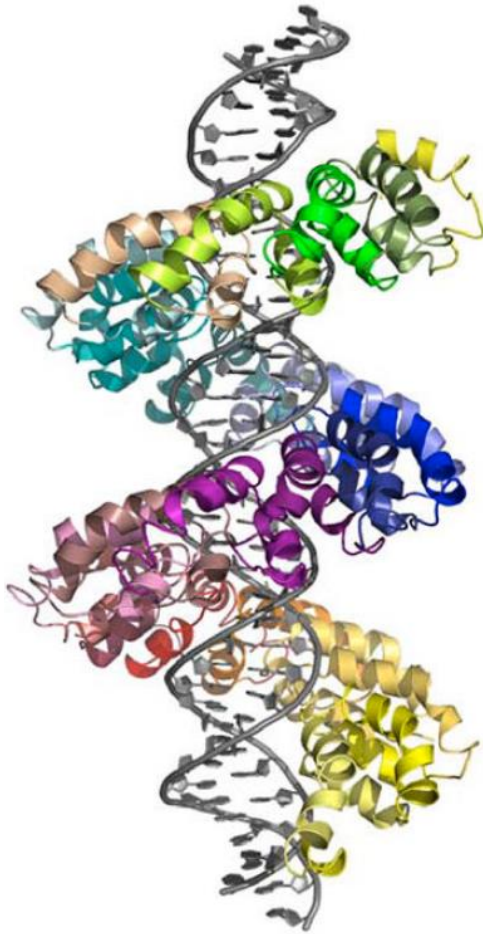


Bonas U. et al., *Curr Opin Microbiol.* 2009 Feb;12(1):37-43.

# Structure of TALEN

## Mechanisms for DNA binding

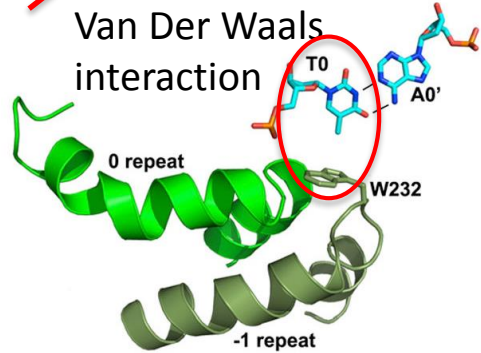
## Schematic view of TALE sequence



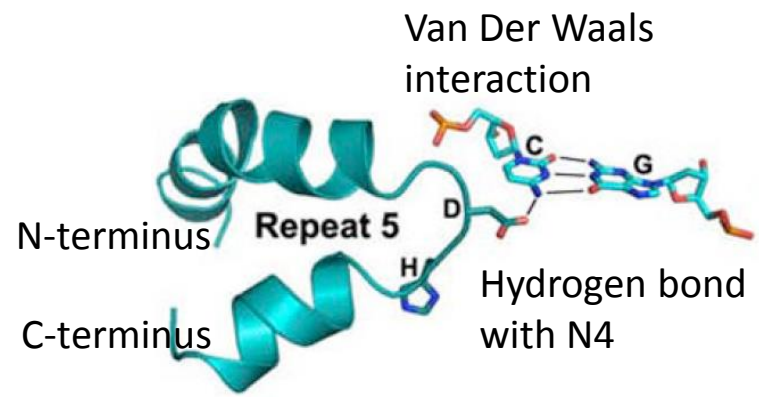
LTPAQVVAIAS HDGGKQALETVQRLLPVLCQAHG

RVD : repeat-variable diresidue

'HD' recognizes Cytosine



The repeat (0, -1) recognizes Thymine

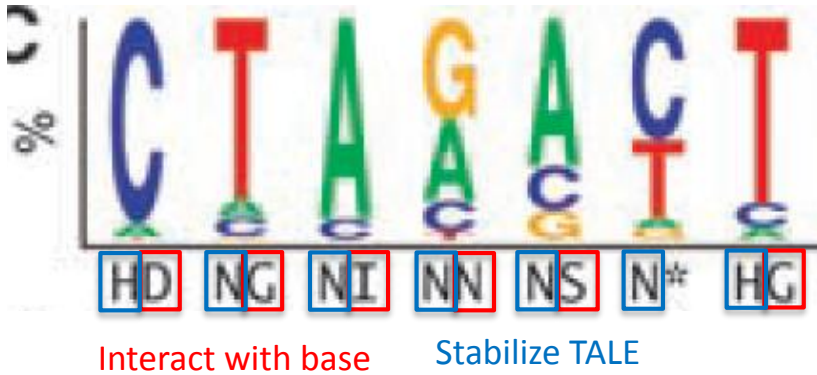


Interactions between TALE and DNA major groove



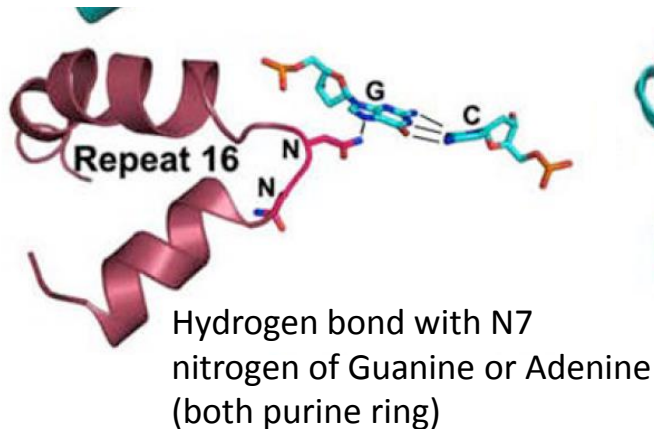
# DNA sequence recognition by TALE

## RVDs and their specificity

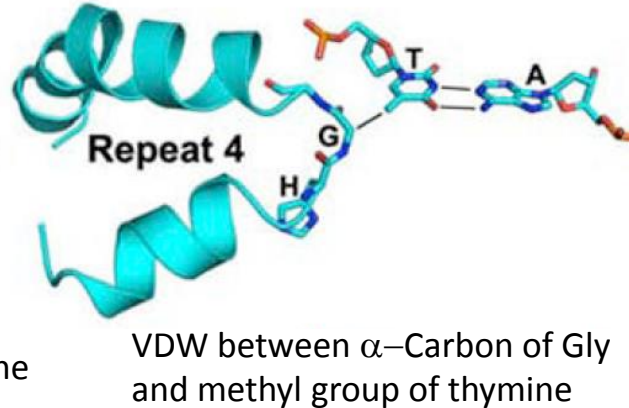


Bogdanove AJ., *et al.*, *Science*. 2009 Dec 11;326(5959):1501.

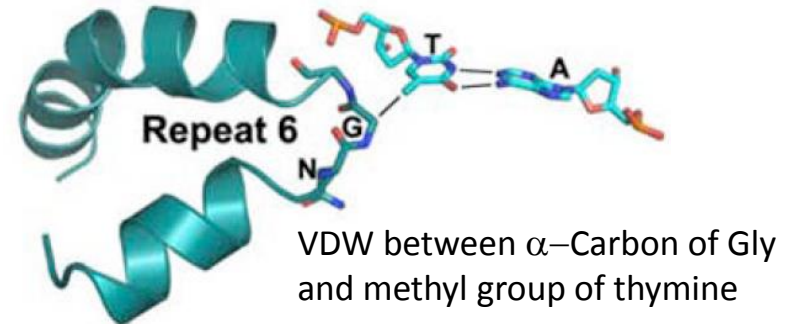
### Interaction between 'NN' and Guanine



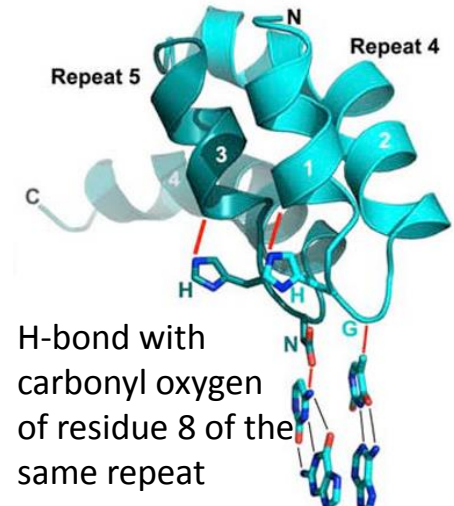
### Interaction between 'NN' and Thymine



### Interaction between 'NG' and Thymine



N or H (the 1<sup>st</sup> of RVD) stabilize the structure



A. N. Mak *et al.*, *Science*, **335**, 716-719 (2012)

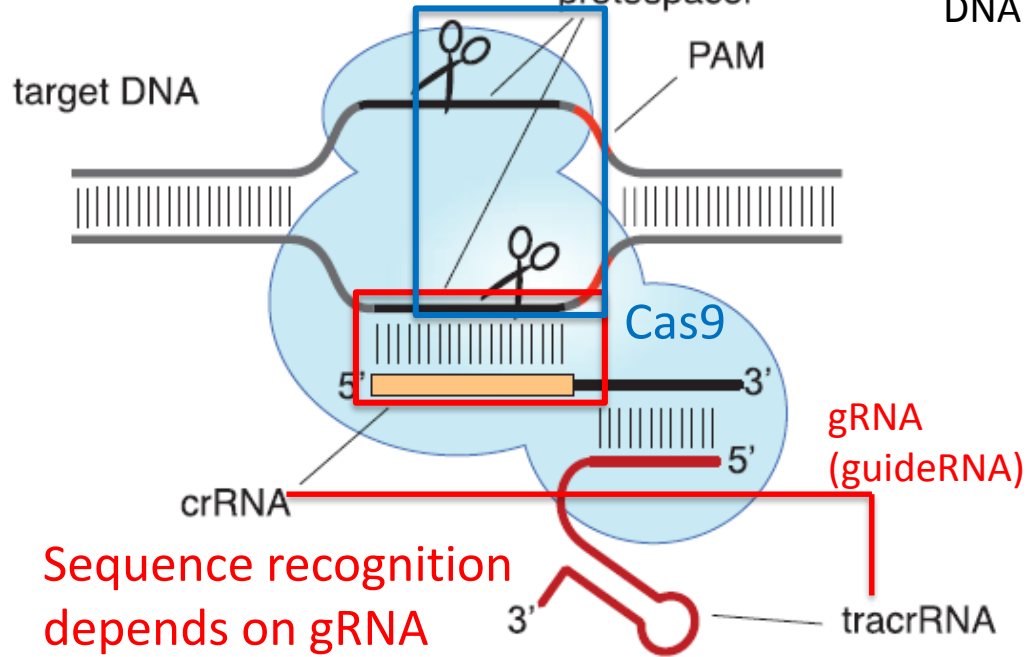
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# Overlook on CRISPR/Cas9

## CRISPR/Cas9 consists of Cas9 protein and gRNA

Cas9 cleaves DNA strand



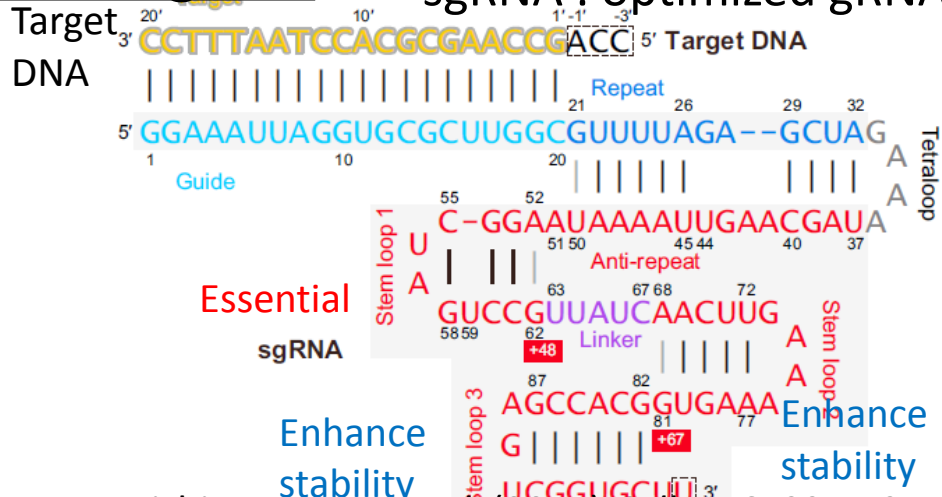
Sequence recognition depends on gRNA

Jinek, M., et al. (2012) *Science*, 337, 816-21

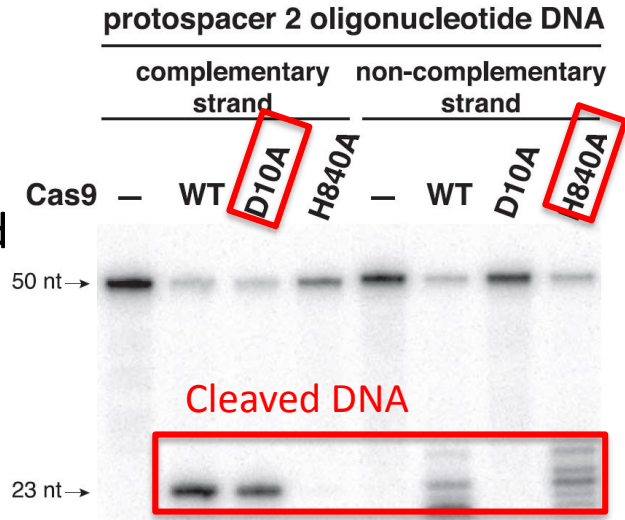
DNA recognition depends on RNA, easily modified  
 restraint of PAM sequence(only 3~6 base)  
 Multiplex editing

Little cost and effort for design

sgRNA : optimized gRNA



Nishimatsu, H., et al. (2014) *Cell*, 156, 935-49  
 Residues responsible for DNA cleavage

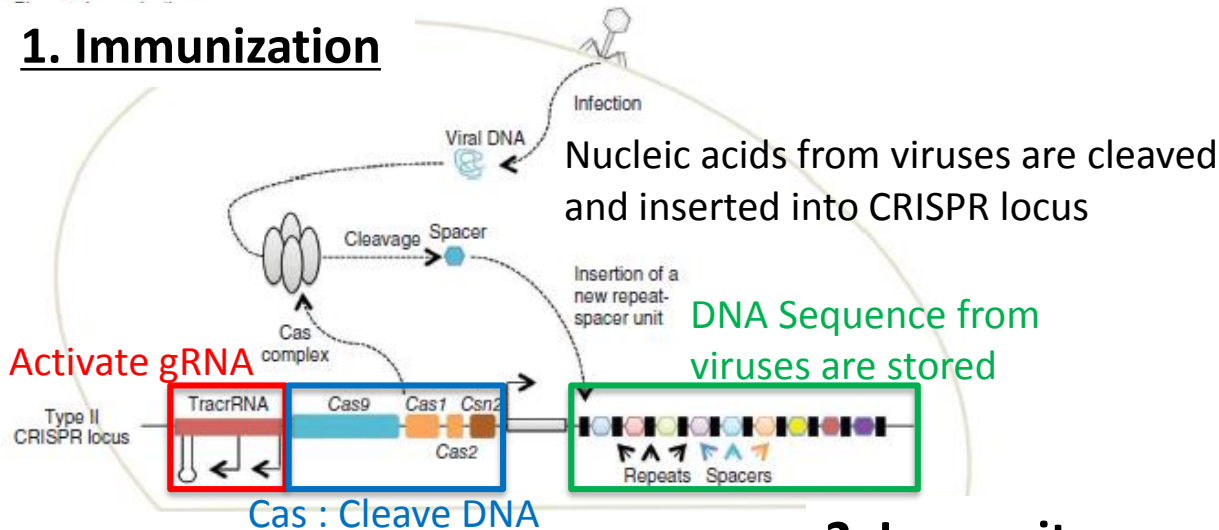


Jinek, M., et al. (2012) *Science*, 337, 816-21

# CRISPR/Cas9 in natural world

## CRISPR/Cas is originally an immune system against nucleic acids from viruses

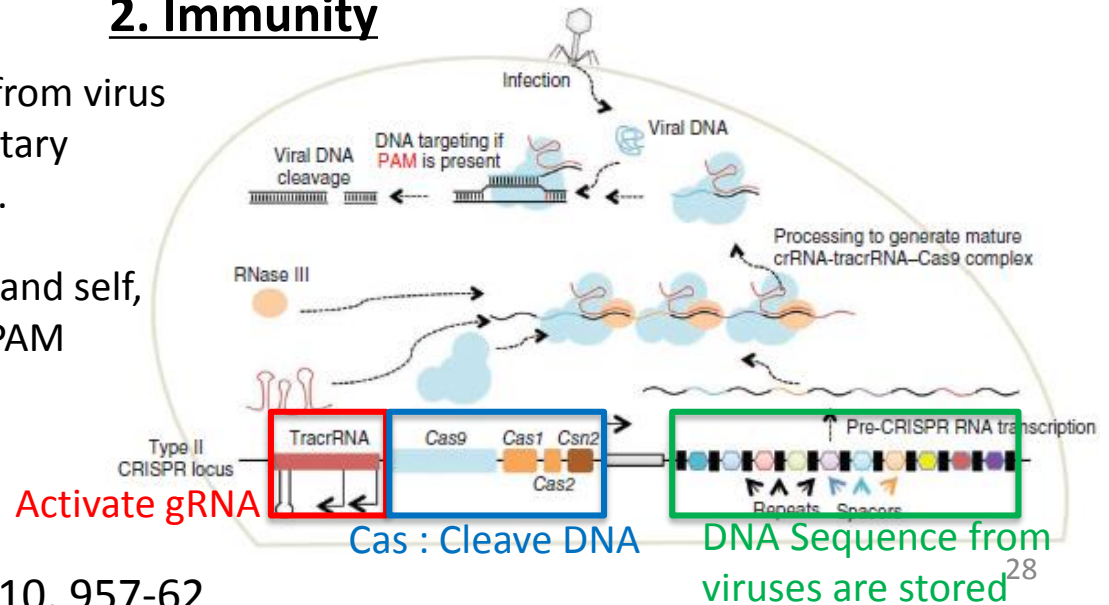
### 1. Immunization



### 2. Immunity

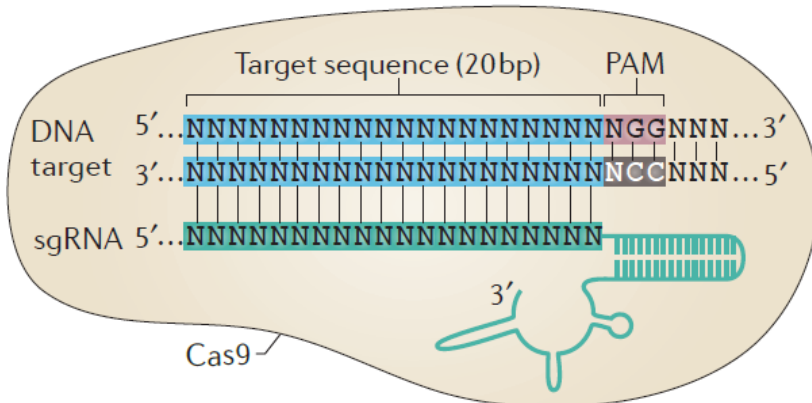
Cas9 cleaves invading DNA from virus recognizing the complementary sequence to the stored one.

To distinguish unself (virus) and self, Cas9 recognizes preceding PAM sequence.



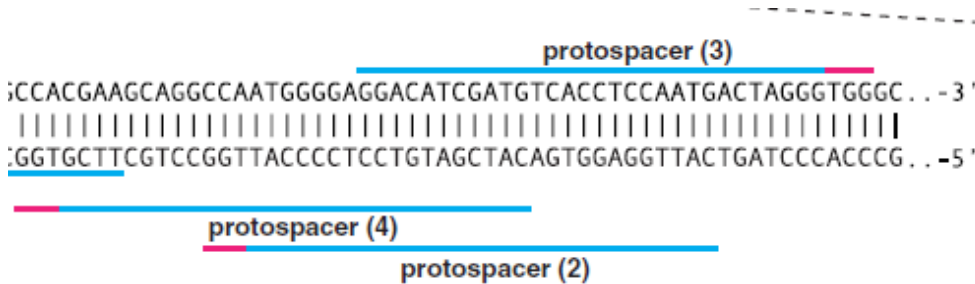
# Design and structure of CRISPR/Cas9

Restraint of PAM sequence is not a problem for practical use



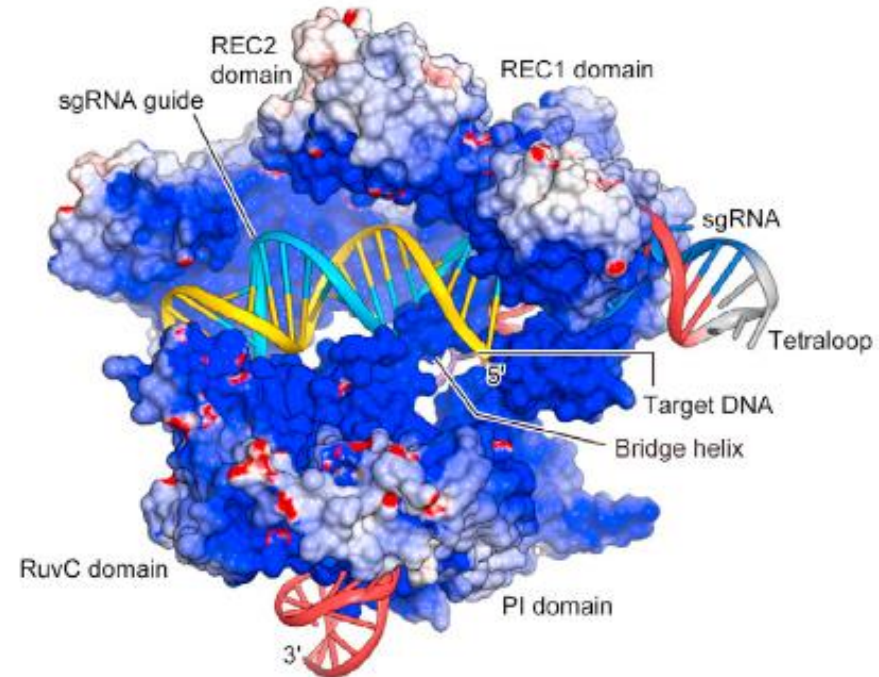
Jacks, T., et al., *Nat Rev Cancer*. 2015 Jul;15(7):387-95.

gRNA must include PAM (NGG) but often several can be designed for 1 gene



Cong, L., et al. (2013) *science*, 339, 819-22

sgRNA; DNA; Cas9 ternary complex



Nishimatsu, H., et al. (2014) *Cell*, 156, 935-49

positively charged groove of CRISPR/Cas9 interacts with negatively charged sgRNA; DNA complex

[See appendix for detailed interactions](#)



# Specificity of CRISPR/Cas9

## Drawback? of CRISPR/Cas9

TALEN is often better than CRISPR/Cas9 in **Specificity**  
**Number of off-target sites of CRISPR/Cas9**

sgRNA name	Targeted chromatin state	sgRNA guiding sequence	PAM	No. of binding sites		
				10	100	1,000
sgRNA 12	Active promoter	GGGGACGCGCTGGCTTCCCG	GGG	13		
sgRNA 11	Active promoter	GGACCGGCTCCCTGGCGGTC	GGG	15		
sgRNA 2	DNase1(+)/intergenic	GAACACAAAGCATAGACTGC	GGG	16		
sgRNA 8	DNase1(-)/intergenic	GCCACTTCTAAGCCCTTGAT	GGG	42		
sgRNA 3	DNase1(+)/intergenic	GGCCCAGACTGAGCACGTGA	TGG	43		
sgRNA 1	DNase1(+)/intergenic	GGGAAAGACCCAGCATCCGT	GGG	49		
sgRNA 5	Active promoter	GCGGTACGCCGCTTCAGTGA	GGG	51		
sgRNA 9	DNase1(-)/intergenic	GAAACTGGTCCCGTTACAG	GGG	120		
sgRNA 10	DNase1(-)/intergenic	GATGAGATAATGATGAGTCA	GGG	201		
sgRNA 7	Inactive promoter	GCCTAGGCAGTGGGGGTGCA	GGG	284		
sgRNA 4	Active promoter	GGCACTGCGGCTGGAGGTGG	GGG	483		
sgRNA 6	Inactive promoter	GGCCCTGCAATGTCAAGGGA	GGG	1,281		

Kuscu, C., et al. (2014) *nature*, 32, 677-83

Depending on sgRNA, 10-1,000 off-target sites exist.

**CRISPR/Cas9 and TALEN are not so different in terms of mismatch tolerance per recognition bases**

CRISPR/Cas9 : 1-3 mismatches / 20 bases

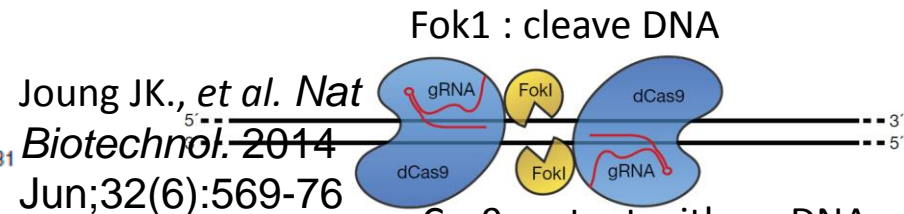
TALEN : 1-2 mismatches / 18 repeats

Mali, P., et al. (2013) *nature biotechnology*, 31, 833-38

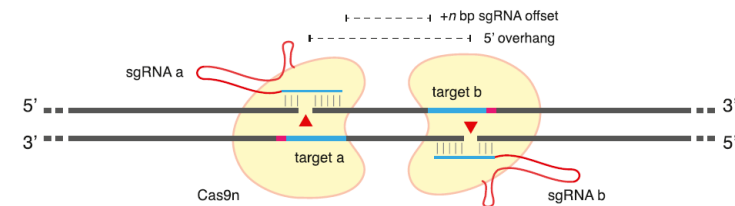
## Strategies to improve specificity of CRISPR/Cas9

1. **Truncate sgRNA** target sequence by 2 or 3 nucleotides (bases which weakly recognize target sequence)
2. **two Cas9s** for one target gene.
3. Cas9 mutants

### 2-1. Cas9-Fok1



- 2-2. Cas9 nickase (cleave only one of the DNA double strand)



Ran, F. A. et al., *Cell* 154, 1380–1389 (2013).

# CRISPR/Cas9 mutant with high specificity

## Construction of highly specific Cas9 mutant

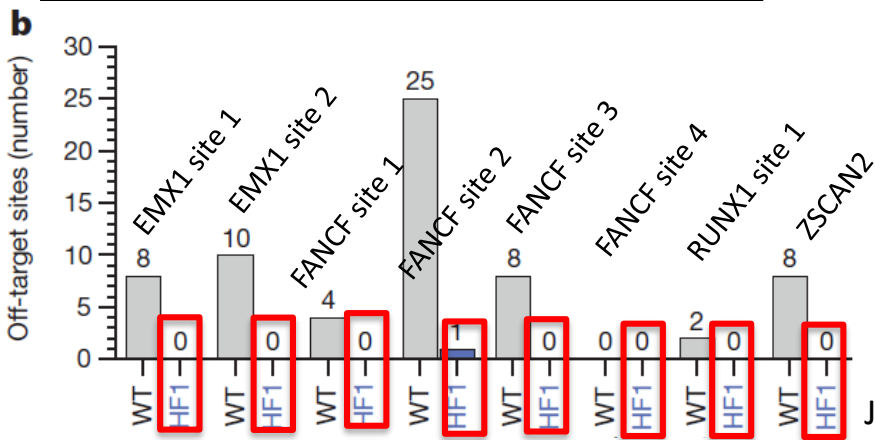
Problems about former attempts to increase specificity

- **unproven efficacies** on genome-wide scale
- potential to create **more new off-target**
- need for fusion of functional domains to Cas9
- reduction in **targeting range**
- more challenges for **delivery**



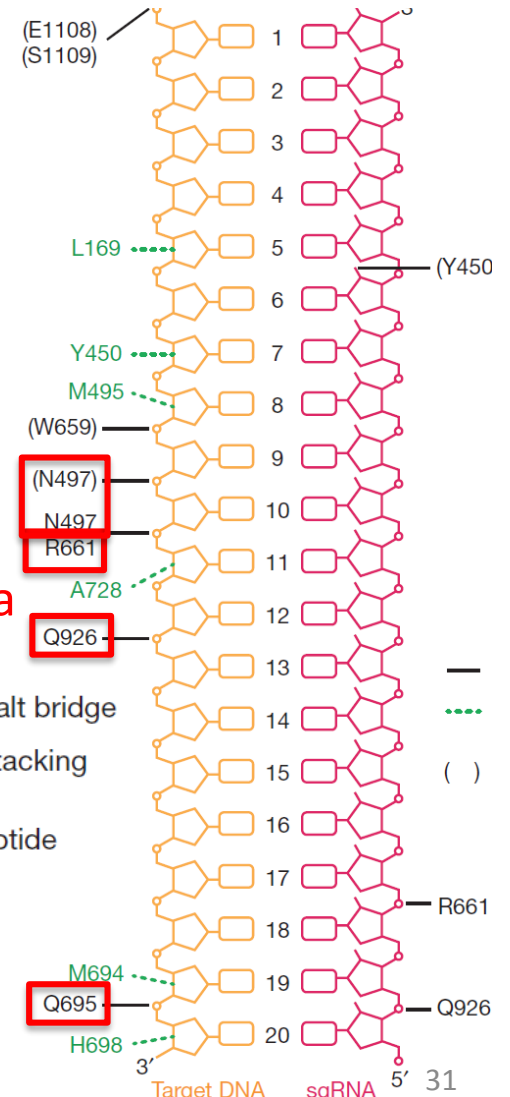
**Necessity for the development of a robust and easily used strategy**

**Mutations to reduce Cas9-DNA interactions lead to the increase in specificity**



**Mutated to Ala**

- Hydrogen bond/salt bridge
- Hydrophobic or stacking interaction
- ( ) Interaction via peptide backbone



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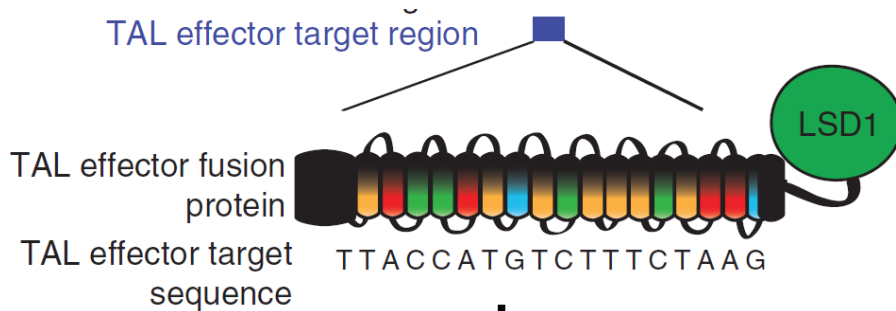
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# Epigenome editing tools

## two ways to deliver effectors to specific DNA locus

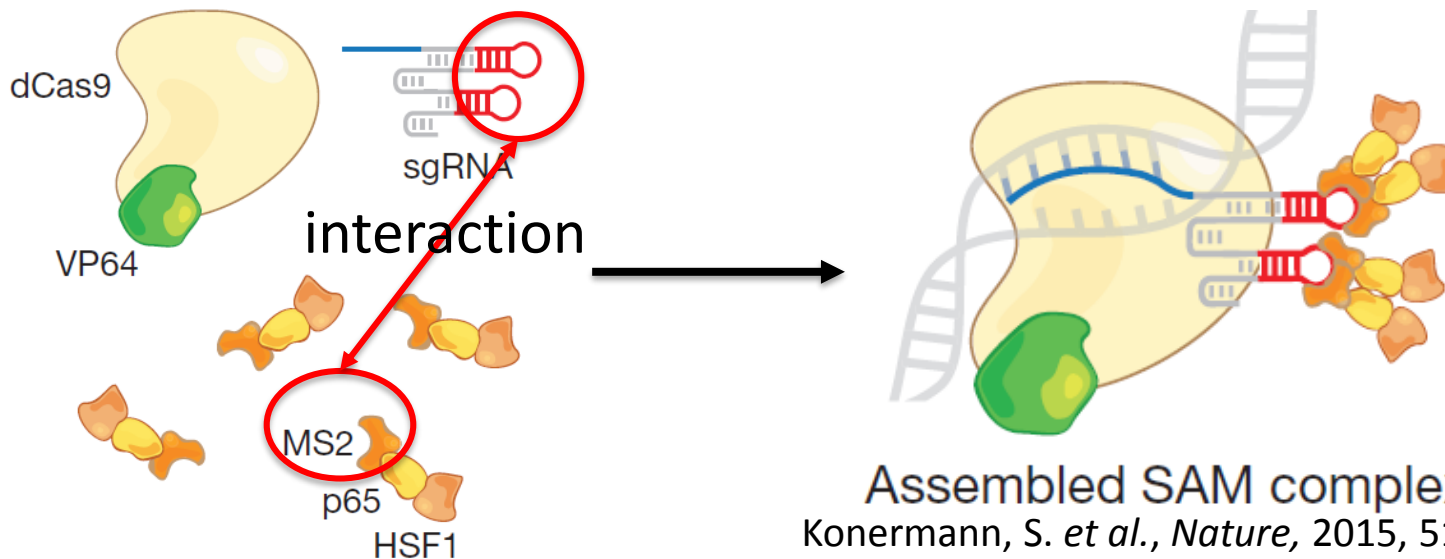
### 1. Directly fuse effector domains to DNA binding protein



LSD1 : histone demethylase

Bernstein BE., *et al.*, *Nat Biotechnol.*, 2013 Dec;31(12):1133-6.

### 2. Use of RNA binding protein



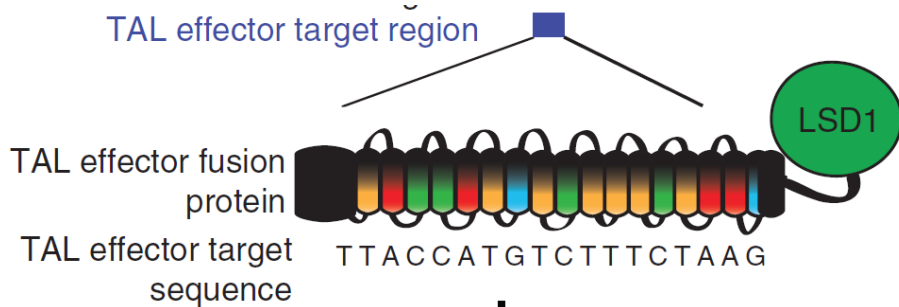
Assembled SAM complex

Konermann, S. *et al.*, *Nature*, 2015, 517, 583-8

# Use of histone demethylase fused TALE

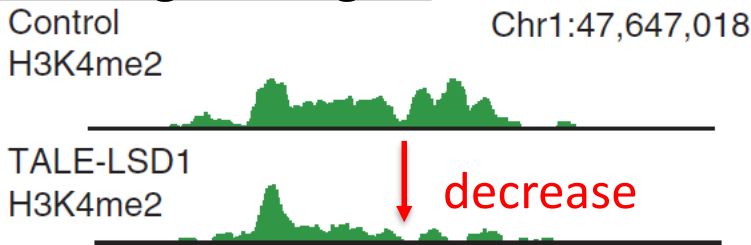
two ways to deliver effectors to specific DNA locus

## 1. Directly fuse effector domains to DNA binding protein



LSD1 : histone demethylase

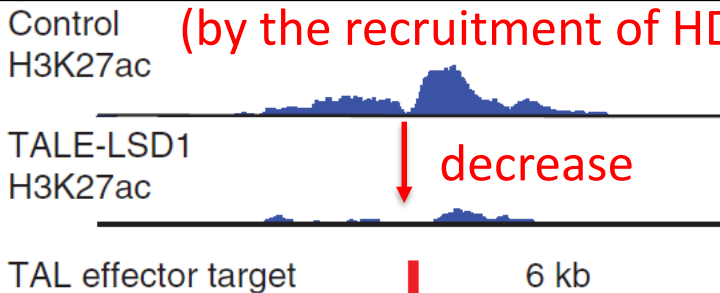
## LSD1 catalyzed the removal of H3K4 methylation around targeted region



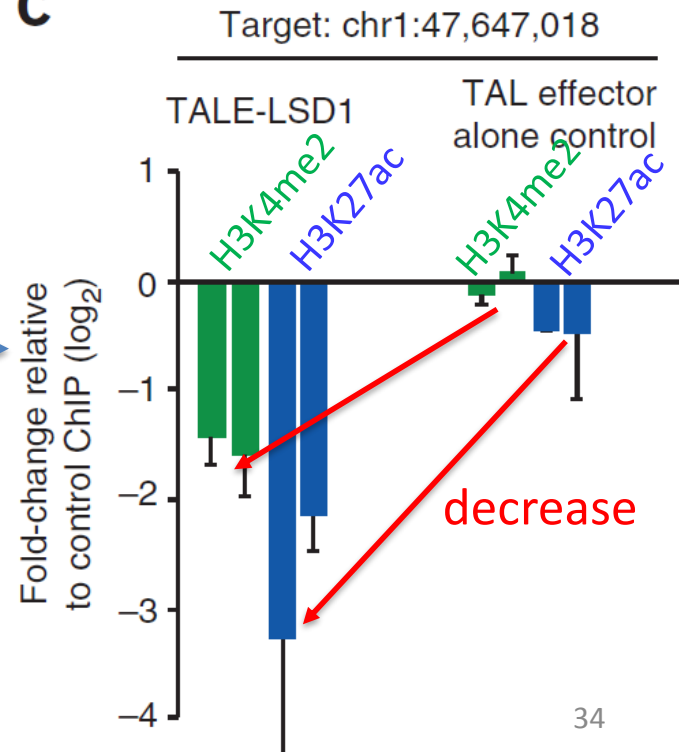
Quantitative data

## Also, removal of H3K27ac was observed

(by the recruitment of HDAC)



C

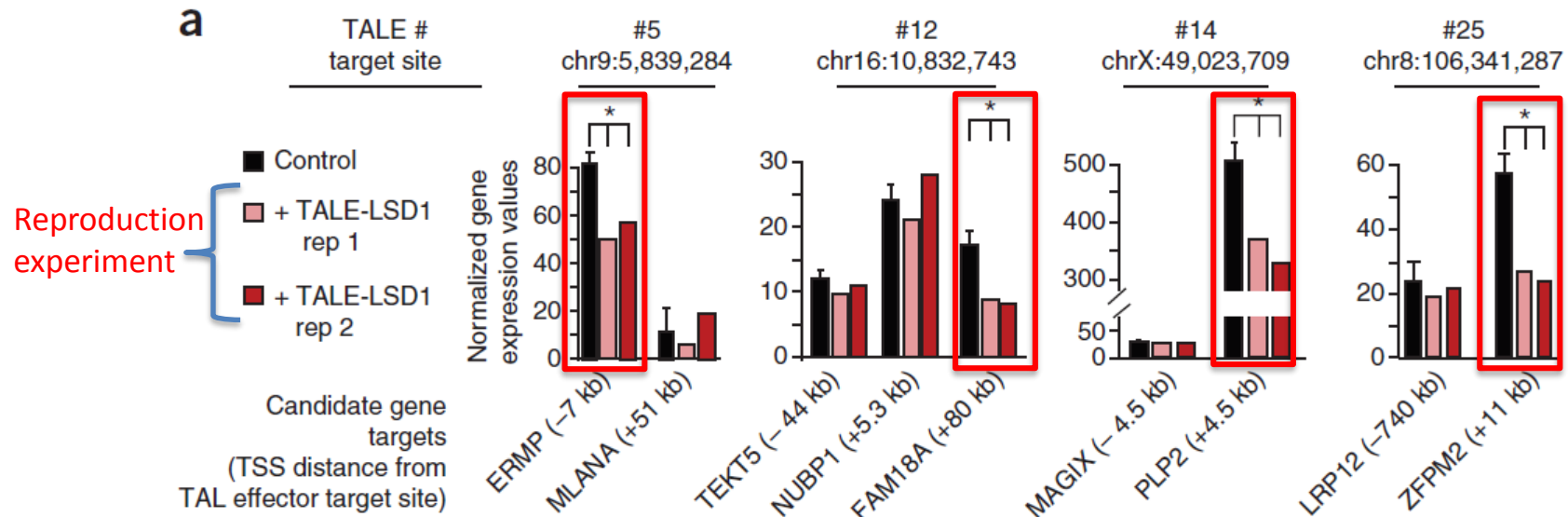


# Epigenetics study using TALE-LSD1①

Prior study **indicate** that sequence elements enriched for H3K27ac and H3K4me2 exhibit enhancer activity

TALEN-LSD1 enables post-translational modifications to be artificially introduced and their effects to be evaluated.

Four of nine enhancers tested, nearby genes were inactivated



Why did the other show little change?

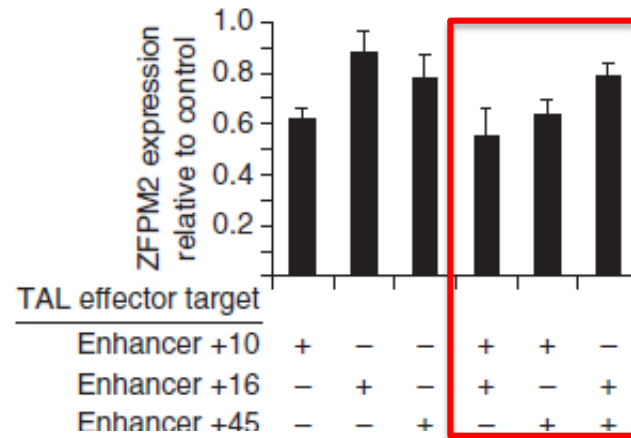
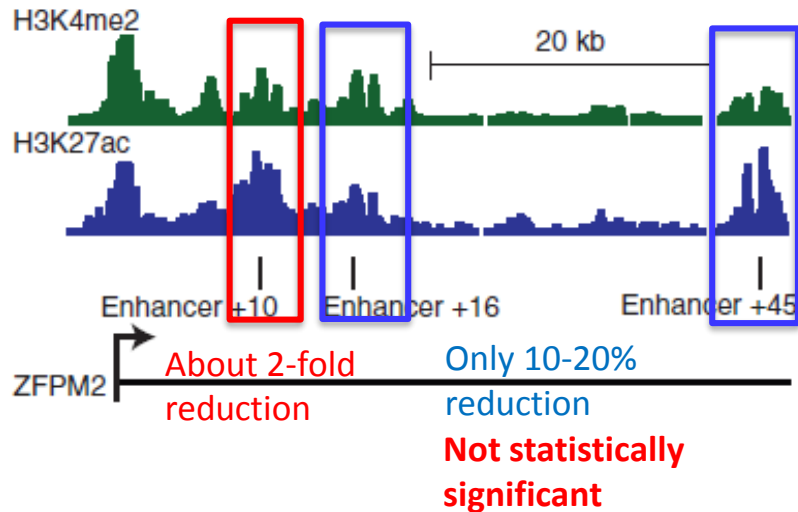
Weak transcriptional effect under detection threshold?

No function as an enhancer in the cell tested?

# Epigenetics study using TALE-LSD1②

In the same enhancer, the region where TALEN-LSD1 is targeted may change the degree of effect

Three enhancers for ZFPM2 were tested, yielding different inactivation of gene expression



Effect of targeting paired enhancers is less than the sum of their individual effects

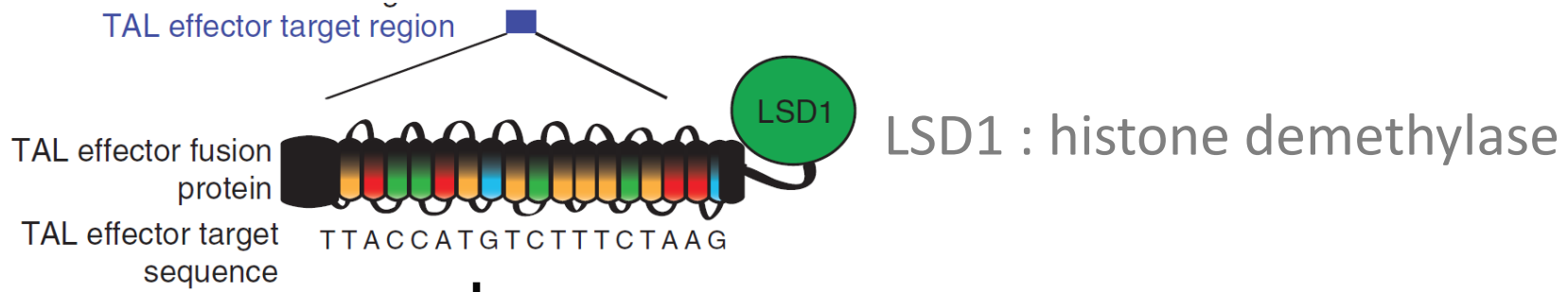
Redundant effect of the multiple enhancers

TALE-LSD1 fusions are valid tools to study complex regulatory interactions among multiple enhancers and genes in a locus

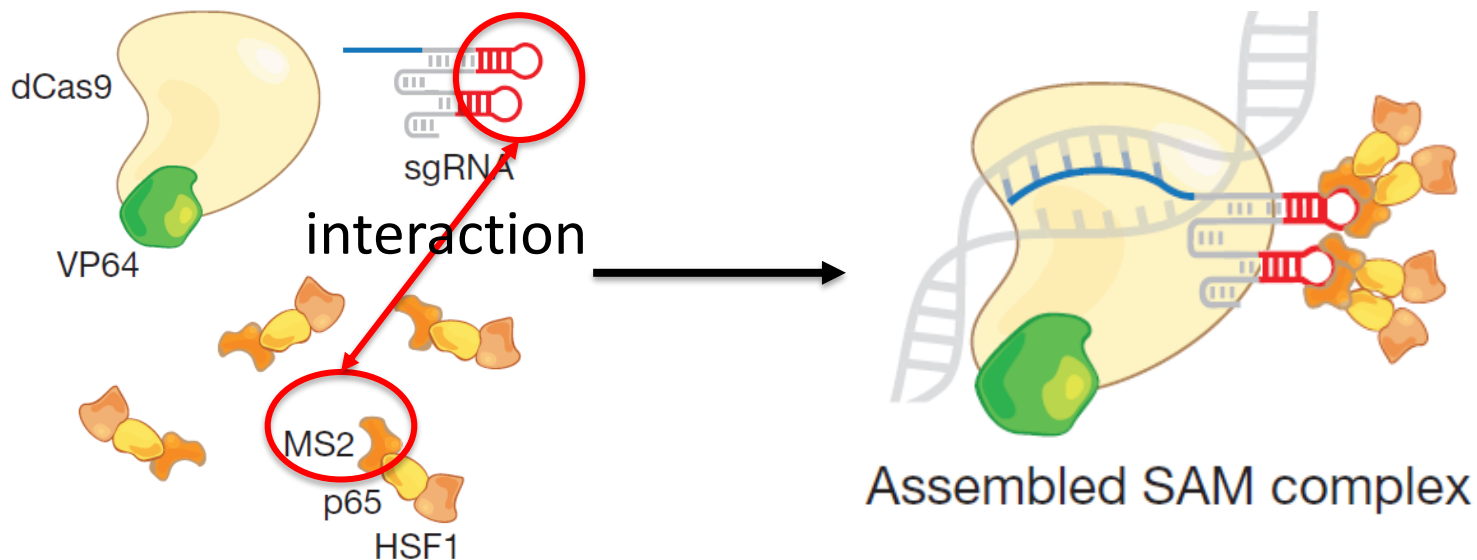
# Specific gene activation tool using CRISPR/Cas9

## two ways to deliver effectors to specific DNA locus

### 1. Directly fuse effector domains to DNA binding protein



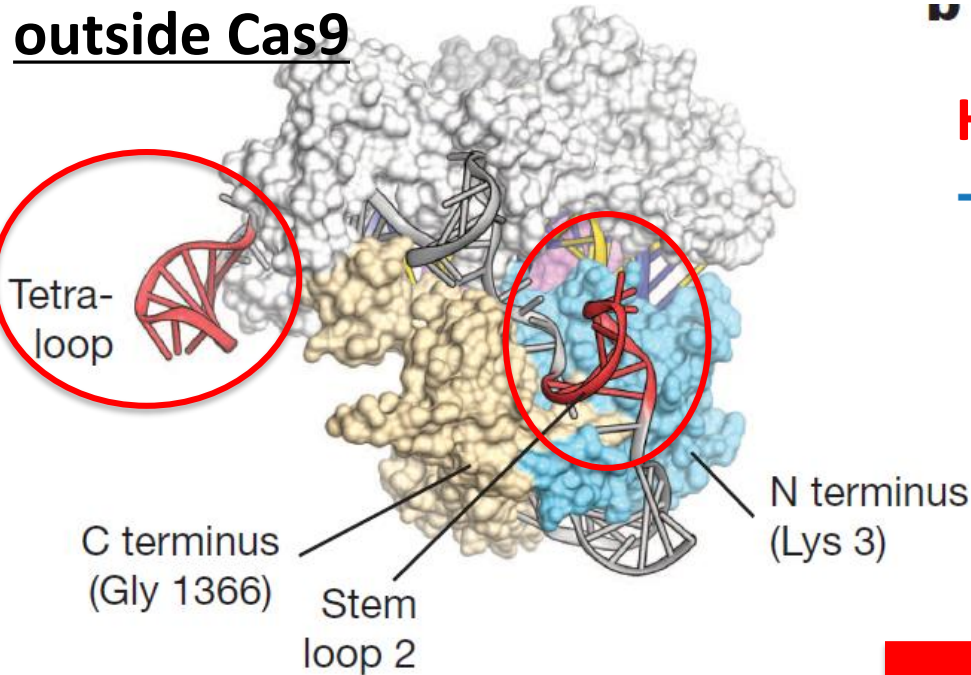
### 2. Use of RNA binding protein



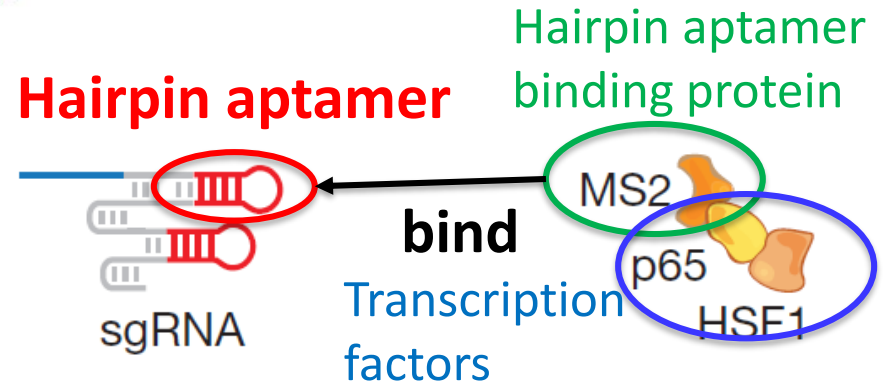
# Structure based design of SAM system

## Structure based design for construction of RNA aptamer

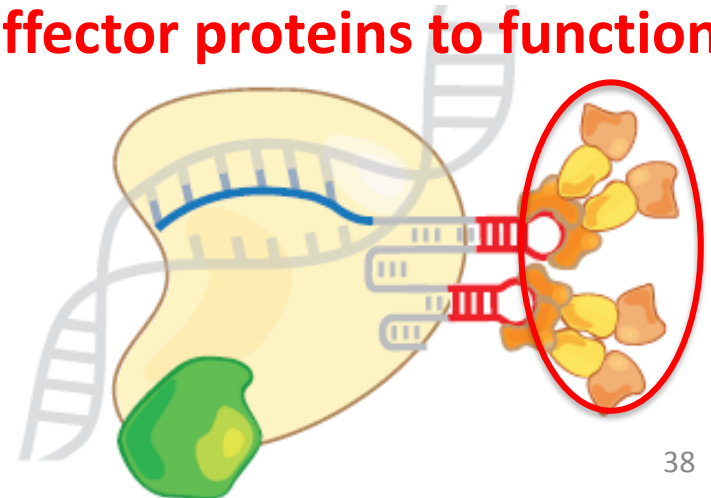
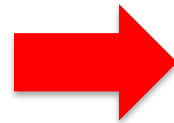
### Tetra loop and stem loop2 is exposed outside Cas9



→ appropriate for scaffold for effector proteins



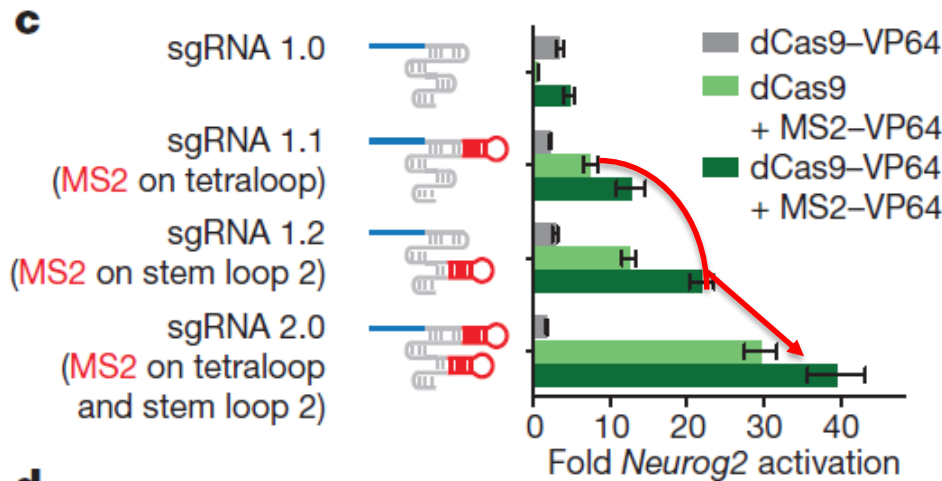
**Good environment for effector proteins to function**



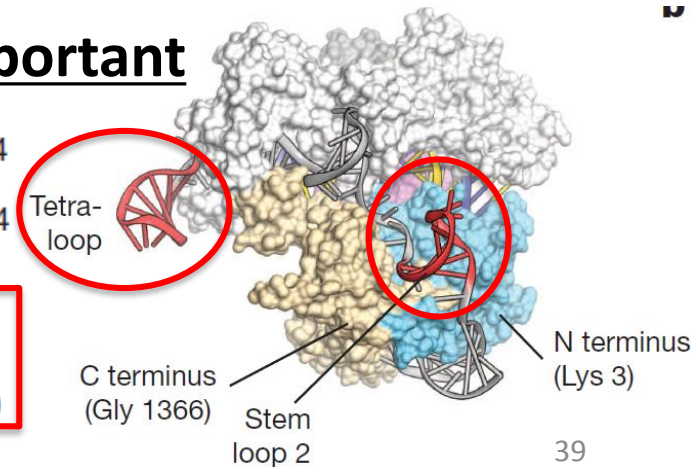
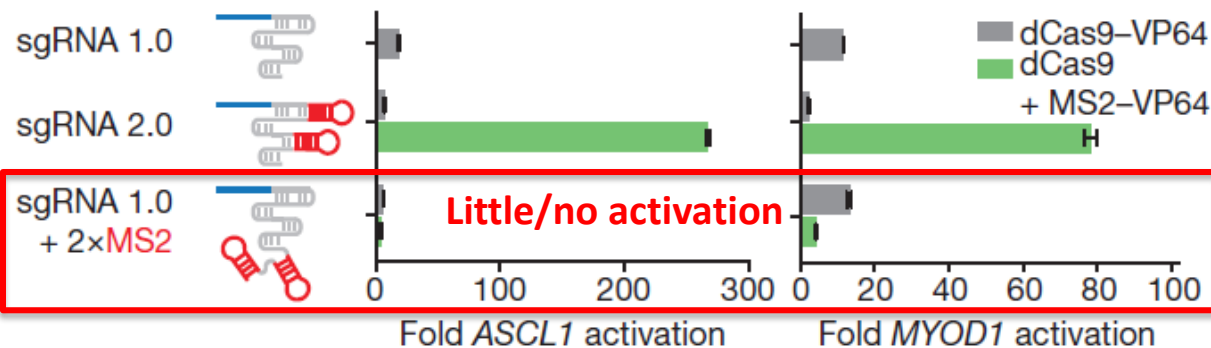
# Activation of specific gene achieved with SAM

## Confirmation of the validity of the design

## Efficient gene activation was achieved dependent on the number of hairpin aptamers



## Structural position of hairpin aptamers is important



**Not exposed on the surface of Cas9**

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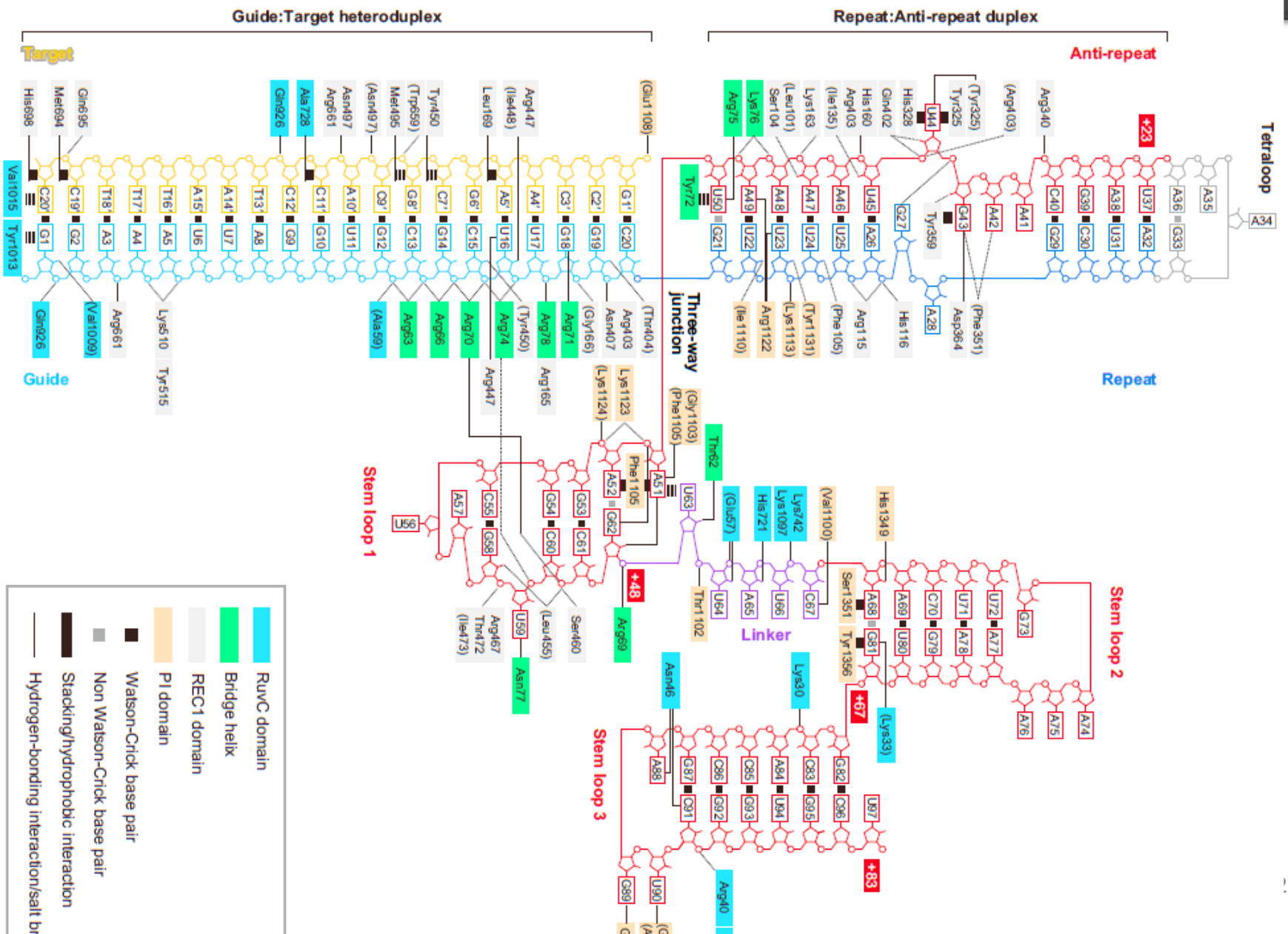
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# Summary

- genome editing tools facilitated the study of gene functions
- **ZFN** was the first to be invented but has not been widely used due to **its difficulty in designing**
- **TALEN** overcame the drawback of ZFN due **to its simplicity in recognizing DNA sequence.**
- **CRISPR/Cas9**, using gRNA for recognition of DNA sequence, rendered **genome editing much easier** and is now intensively studied
- genome editing tools are applicable to **edit epigenome**, enabling **more direct study of epigenetics.**

# Appendix; detailed interactions of Cas9-gRNA-DNA

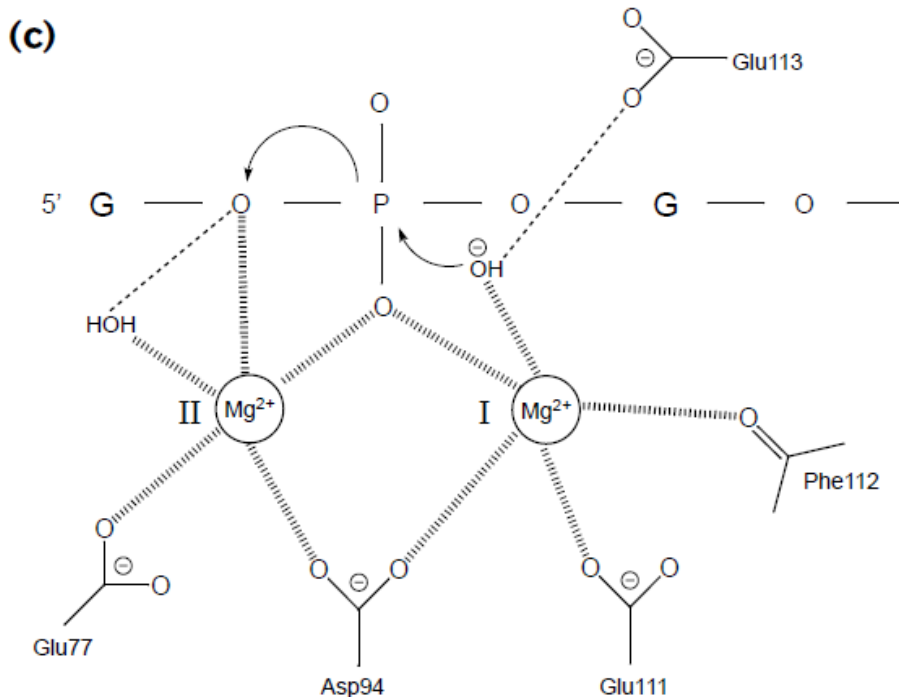


# ZFN (Zinc-finger nuclease)

ZFN consists of Fok1 endonuclease cleavage domain and ZF protein.

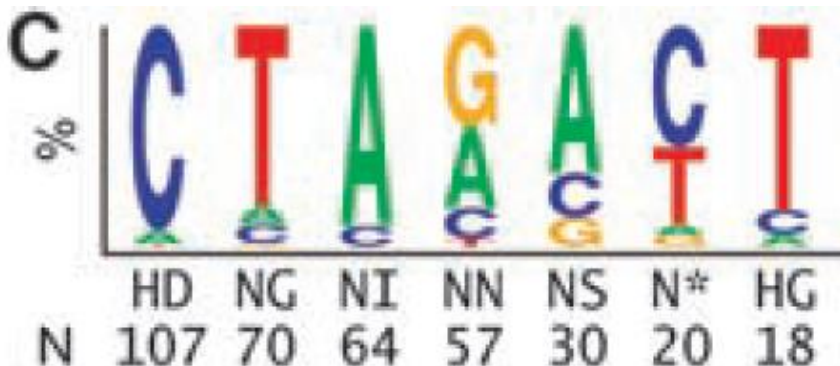
Fok1 endonuclease consists of Recognition domain and Cleavage domain

Recognition domain binds 5'-GGATG-3'

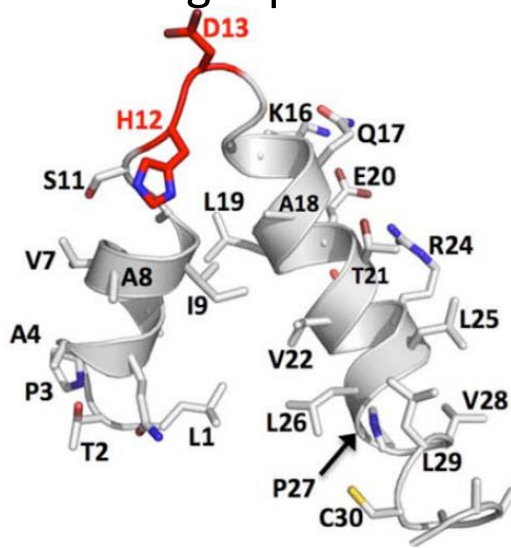


# TALEN (transcription-activator-like effector nuclease)

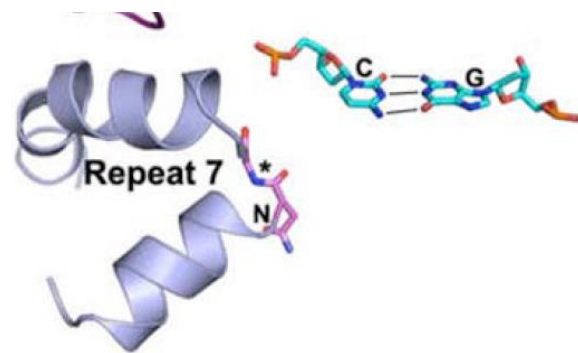
## RVDs and their specificity



Structure of the DNA binding repeat



'N\*' extends less deeply into the major groove



# CRISPR/Cas9

## Application to genome-scale gene activation screen

