Transition Metal Catalysis in Living Systems

Literature Seminar #3
2022/4/15 (Fri)
Wataru Atsumi (B6)

- **Introduction**
- >Applications in medicine and chemical biology
 - Cu-triggered ADC linker cleavage and reversible modification
 - Synthetic prodrug strategy for cancer treatment
 - Perspective
- **≻**Summary

[Nobel prize in chemistry]

Olefin metathesis (2005)

$$R^1 / + / R^2 \stackrel{Ru}{\longrightarrow} R^1 / M_{R^2} + =$$

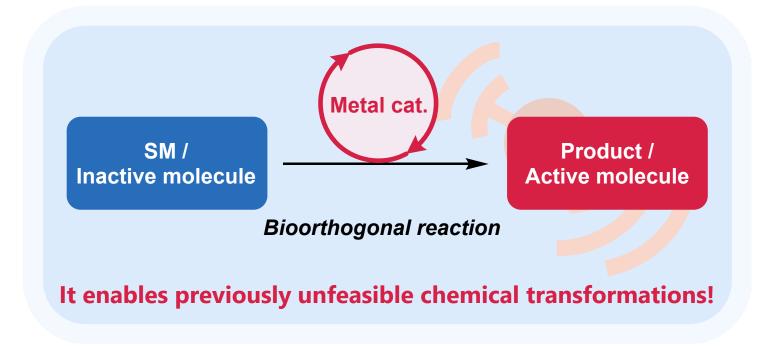
Pd-catalyzed cross coupling (2010)

- ✓ Transformation which has been difficult to perform by classical ways
- ✓ Wider range of reactions
- ✓ Very efficient synthesis

Nobel prize!!



TM Catalysis in Living Systems



[Example: Cu-catalyzed alkyne-azide cycloaddition (CuAAC)]

$$R^{1} \stackrel{\oplus}{N=N=N} \rightarrow + = R^{2}$$

$$R^{1} \stackrel{\oplus}{N=N=N} \rightarrow R^{2}$$

$$R^{2} \stackrel{\bigoplus}{R^{2}}$$

A powerful tool for chemical biology and pharmaceutical sciences!!

TM Catalysis in Living Systems

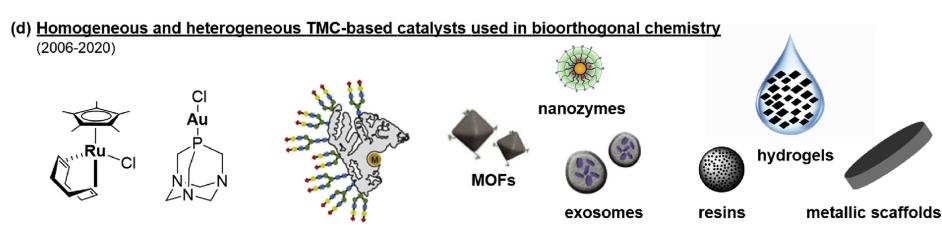
Problems

- **X** Toxicity
- **X** Water solubility
- **X** Stability in water

Requirements

- ✓ Soluble in water
- ✓ Protecting active site

*See appendix for more information.



Organometallic complexes

Metalloenzymes

Nanodevices

Micro and millidevices

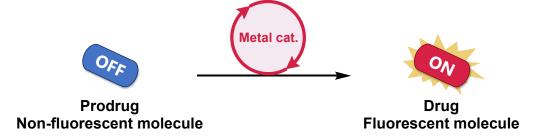
HOMOGENEOUS CATALYSIS

HETEROGENEOUS CATALYSIS

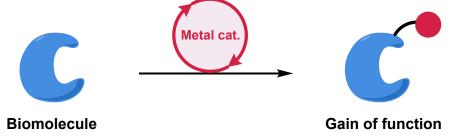
Curr. Opin. Chem. Biol. 2021, 61, 32-42.

Applications of TM Catalysis

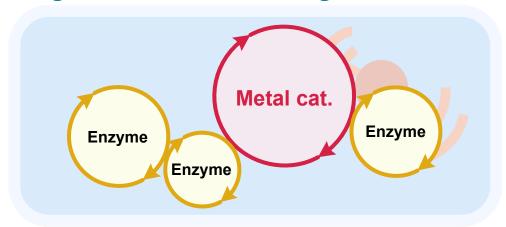
✓ Prodrug activation / Bioimaging



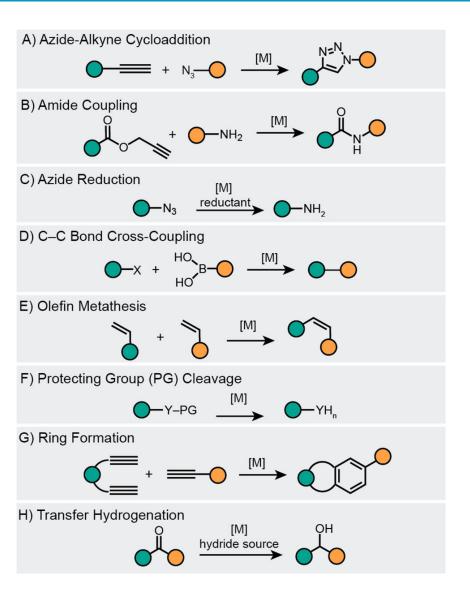
✓ Modification of biomolecules

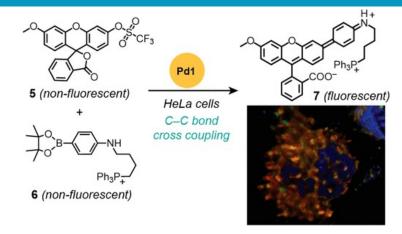


✓ Regulating biological reactions / Creating new cascades



Examples of TM Catalysis in biological conditions 7





Yusop, R. M. et al. Nat. Chem. 2011, 3, 239-243.



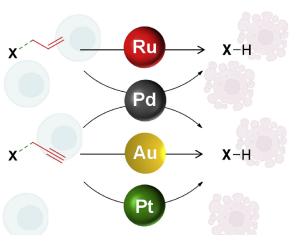
B. G. Davis et al. J. Am. Chem. Soc. 2008, 130, 9642-9643.

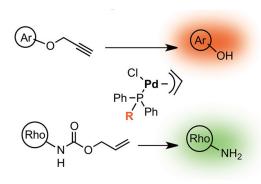
Yugang Bai et al. J. Am. Chem. Soc. 2016, 138, 11077–11080.

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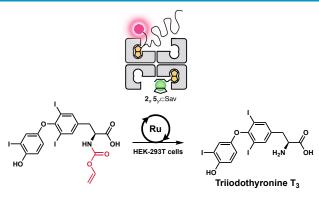
Bioorthogonal Allyl/Propargyl Cleavage

[Previous works]





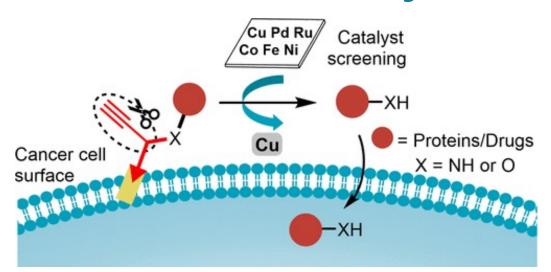
Martínez-Calvo, M. *et al. ACS Catal.* **2018**, *8*, 6055–6061.



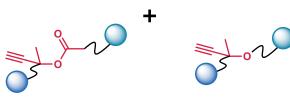
Y. Okamoto et al. Nat. Commun. 2018, 9, 1–7.

TM-catalyzed cleavage reactions have been mainly focused on <u>terminal</u> decaging.

(This work: Internal bond cleavage)



Cu(I) complex



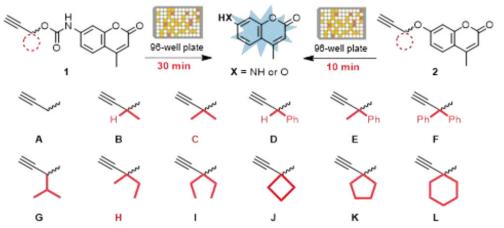
dual-substituted propargyloxycarbonyl (dsProc)

dual-substituted propargyl (dsPra)

- ✓ ADCs linker cleavage
- Reversible cell modification
- ✓ Protein manipulation

Screening and Optimization

(Systematic screening)

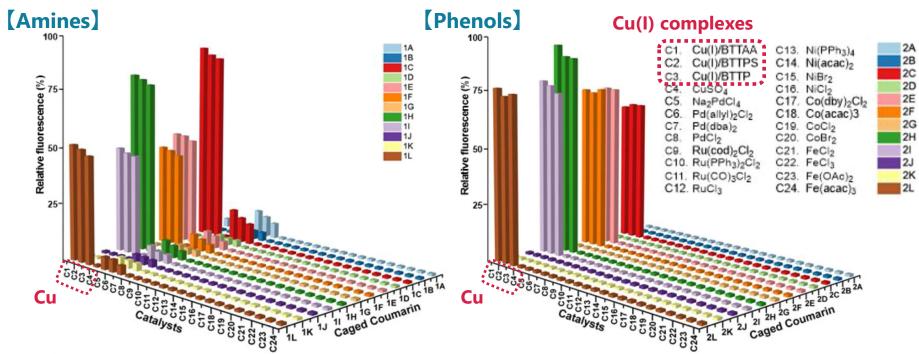


Screening using fluorogenic coumarins

- **24** different transition metal species (C1–C24)
- 12 different substrates (A–L)



- ✓ Cu complexes (except for CuSO₄) showed highly efficient cleavage.
- ✓ Other metals showed lower or no activity.
- ✓ 1D and 2H gave the highest reactivity respectively.



Xin Wang et al. J. Am. Chem. Soc. 2019, 141, 17133-17141.

Action of Cu(I) Complex

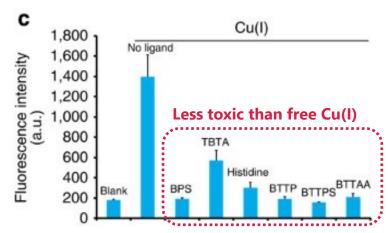
[Proposed catalytic cycle]

$$= \bigcap_{O-Ar} \bigcap_{[Cu]} \bigcap_{O-Ar} \bigcap_{H^+} \bigcap_{O-Ar} \bigcap_{H^2O} \bigcap_{O+Ar} \bigcap_{O-Ar} \bigcap_{O-Ar}$$

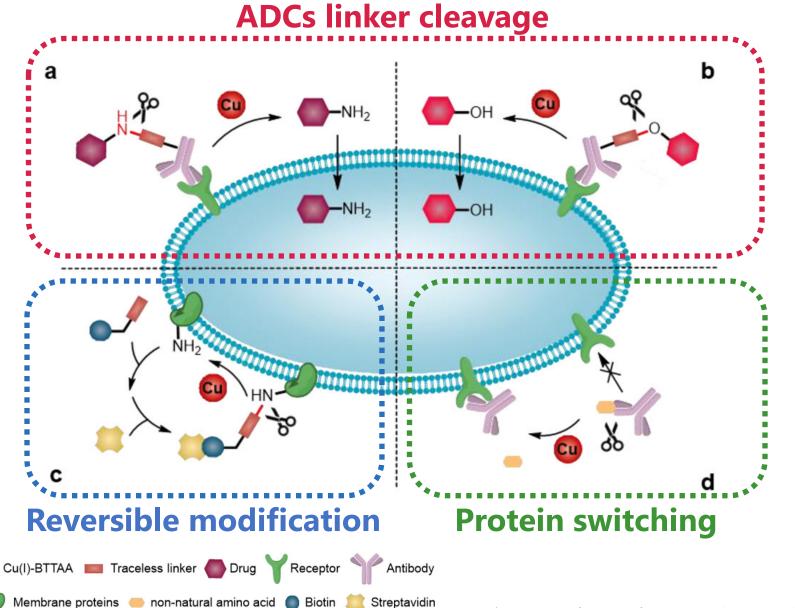
[Cu(I) stabilizing ligands]

$\textbf{Details} \, \rightarrow \, \textbf{See appendix}$

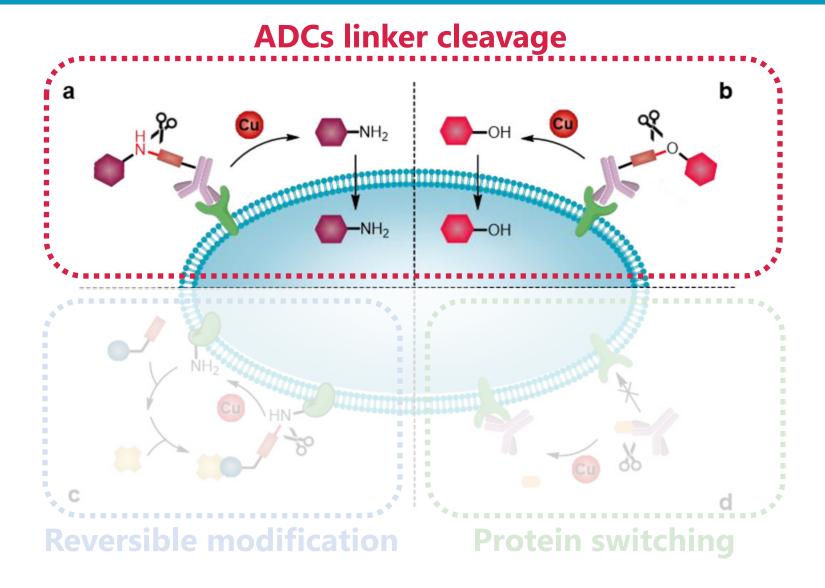
[ROS generation of Cu(I) complexes]



Xin Wang et al. J. Am. Chem. Soc. **2019**, 141, 17133–17141. Yang M. et al. Nat. Commun. **2014**, 5, 4981.

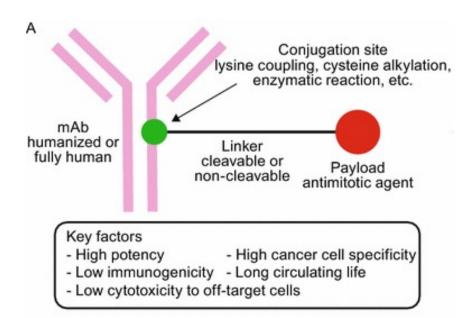


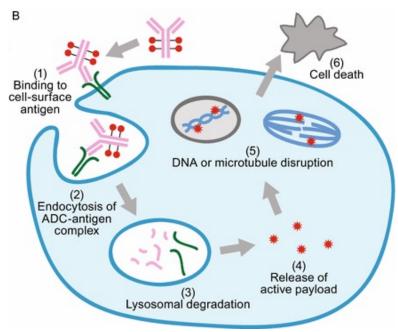
Cu-Triggered ADCs Linker Cleavage



Antibody-Drug Conjugates (ADCs)

(Structure and mechanism of action of ADC)





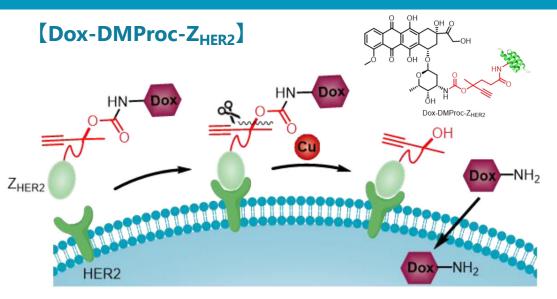
Drawback of noncleavable ADCs

- **X** Inefficient internalization
- **X** Limited cytotoxicity

Catalytically cleavable ADCs

- ✓ Higher cytotoxicity
- ✓ Catalytic amount of triggers (other ways: stoichiometric)

Cu-Triggered ADCs Linker Cleavage (Dox)



Doxorubicin (Dox)

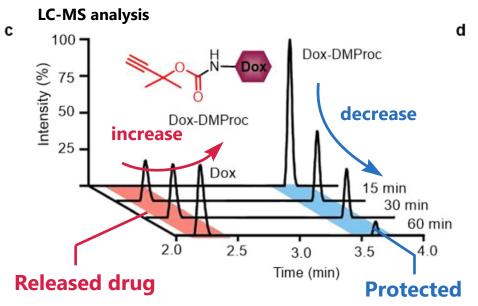
- Anticancer drug
- Forming intercalation with DNA
- **X** Lack of target selectivity
- X High level of side effect (e.g. cardiotoxicity)

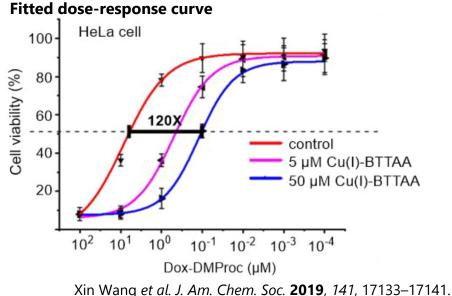
\mathbf{Z}_{HER2}

- Affibody targeting the HER2 receptor
- HER2 overexpresses in certain types of breast cancer → Targeting cancer cells

*HER2: <u>H</u>uman <u>E</u>pidermal grow factor <u>R</u>eceptor 2

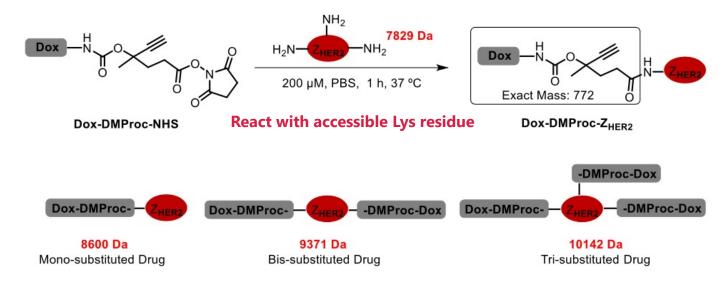
(Dox-DMProc as a model substrate)



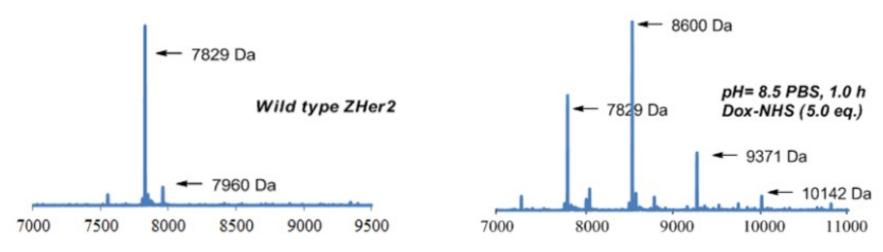


Cu-Triggered ADCs Linker Cleavage (Dox)

(Synthesis of Dox-DMProc-Z_{HER2})



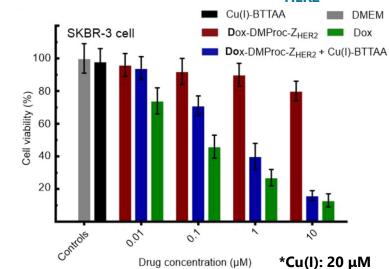
[LC-MS/MS analysis]

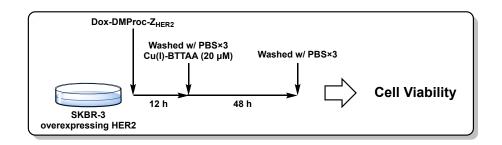


Xin Wang et al. J. Am. Chem. Soc. 2019, 141, 17133–17141.

Cu-Triggered ADCs Linker Cleavage (Dox)

[Effective of Dox-DMProc-Z_{HER2}]

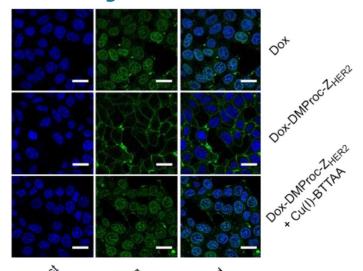




- ✓ DMProc-Z_{HER2} conjugation effectively blocked the toxicity of Dox (— and —)
- ✓ Dox-DMProc-Z_{HER2} restored its toxicity by Cu(I)
 (and)

(Fluorescent images)

nucleus →



Free Dox Bright flu

Bright fluorescent in the nucleus (: Intercalation with double strand DNA)

ADCs

Fluorescence at the membrane areas

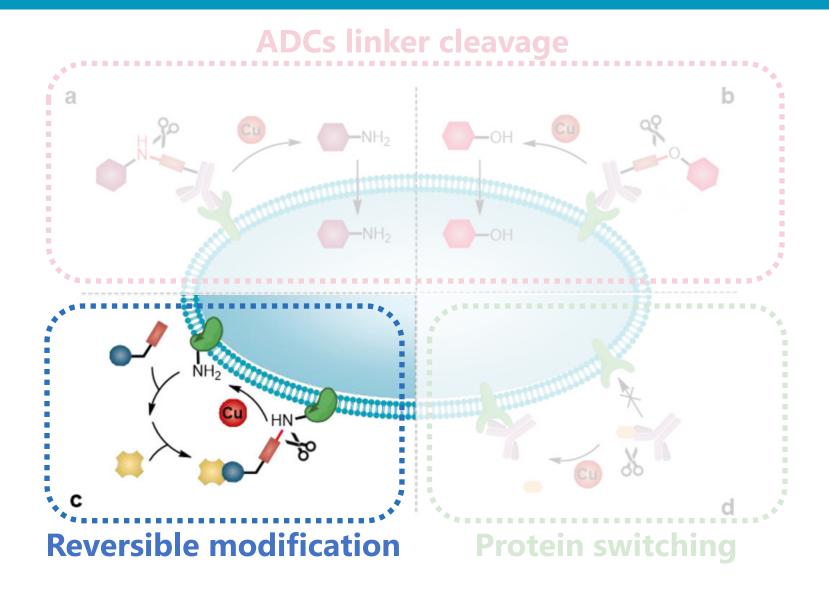
→ Accumulation at cell surface

Free Dox from ADCs

Bright fluorescent within the nucleus

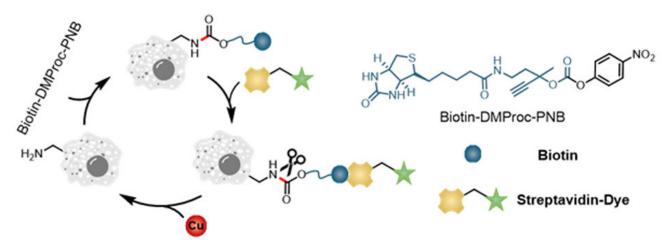
→ Cu-triggered cleavage was demonstrated!

Xin Wang et al. J. Am. Chem. Soc. 2019, 141, 17133-17141.

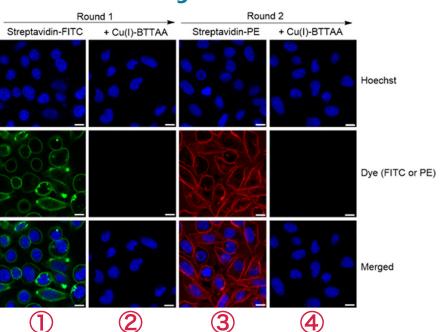


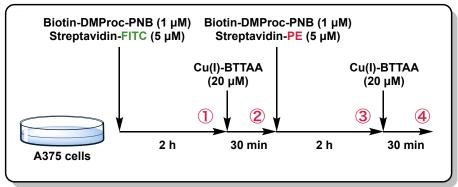
Reversible Cell Surface Modification

(Schematic view of Cu-controlled reversible cell modification)



[Fluorescent images]



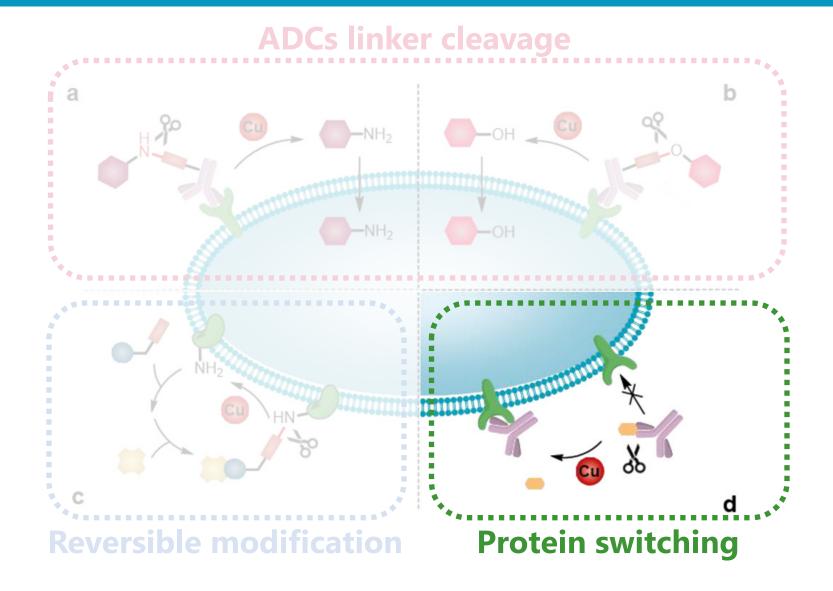


*FITC, PE: Fluorescent dye

Cu-catalyzed linker cleavage allowed the regeneration of native Lys residues on the cell surface.

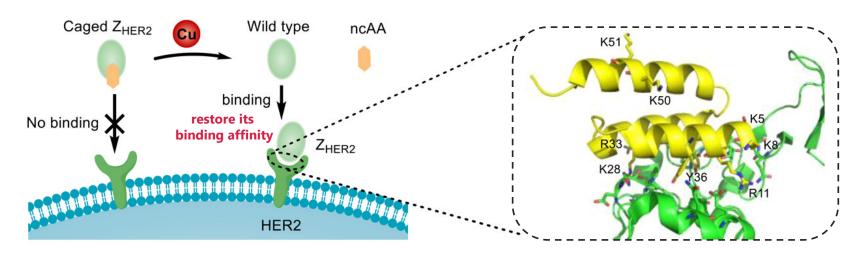
Xin Wang et al. J. Am. Chem. Soc. 2019, 141, 17133-17141.

Cu-Mediated Protein Switching

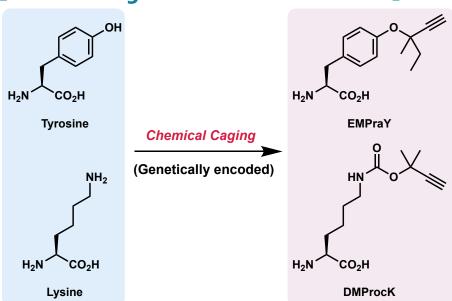


Cu-Mediated Protein Switching

(Schematic view of Cu-controlled protein activation)



[Chemical caged unnatural amino acids]



Incorporation of unnatural amino acids on the binding face of Z_{HER2}

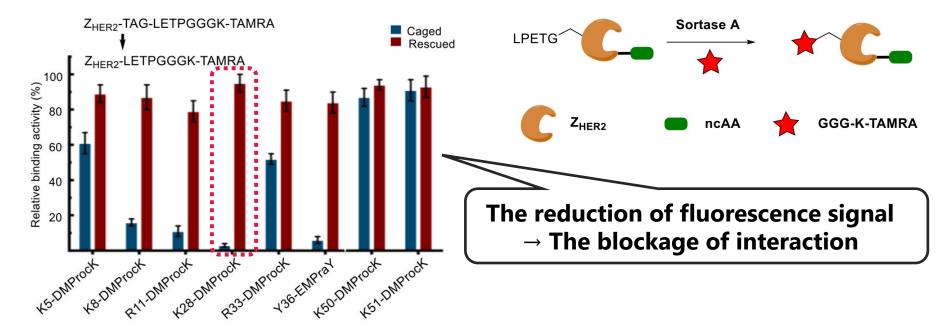


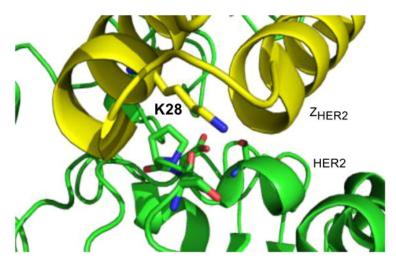
Regulation of its binding affinity by Cu-catalyzed reaction!

Xin Wang et al. J. Am. Chem. Soc. 2019, 141, 17133-17141.

Cu-Mediated Protein Switching

(Binding activity before and after Cu catalysis)





✓ Chemical caging through K28 almost completely blocked the interaction between Z_{HER2} and HER2

Manipulation of protein-protein interaction *in situ*.

Short Summary & Perspective

>ADC linker cleavage

It allows extracellular release of payloads that can overcome the drawback of noncleavable ADCs.

Reversible cell surface modification
It can be applied for the cell capture, which could be quite useful in cancer cell diagnosis.

> Protein manipulation

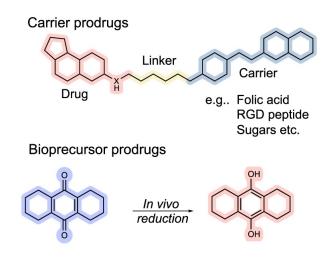
It has the potential to facilitate the development of **protein-based prodrug therapy**.

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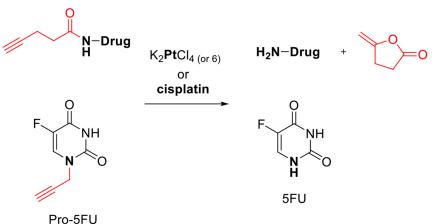
TM-Catalyzed Prodrug Activation

Types of prodrug

Physiologically activated



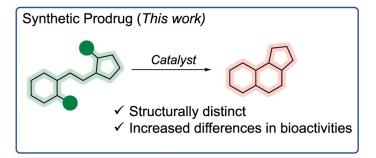
[Decaging prodrug by TM catalysis]



Externally activated

Decaging Prodrug





Nasibullin, I, Tanaka K. et al. Nat. Commun. 2022, 13, 1–12.

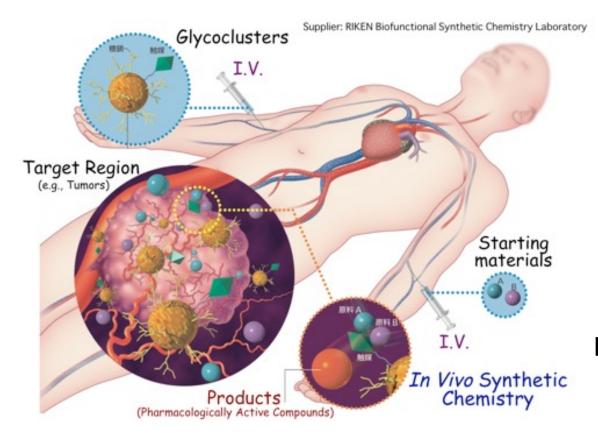
Decaging prodrug activation strategy by TM catalysis has been demonstrated in many papers.

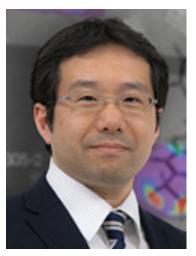


How about synthetic prodrug?

B. L. Oliveira et al. J. Am. Chem. Soc. 2020, 142, 10869-10880.

Therapeutic In Vivo Synthetic Chemistry



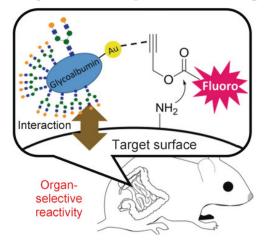


Prof. Katsunori Tanaka Tokyo Tech & RIKEN

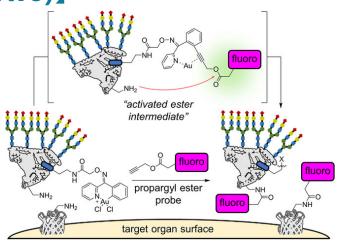
Project

The development of a model system where bioactive compounds can be synthesized within living animals.

[Au catalysis at specific organs (in vivo)]

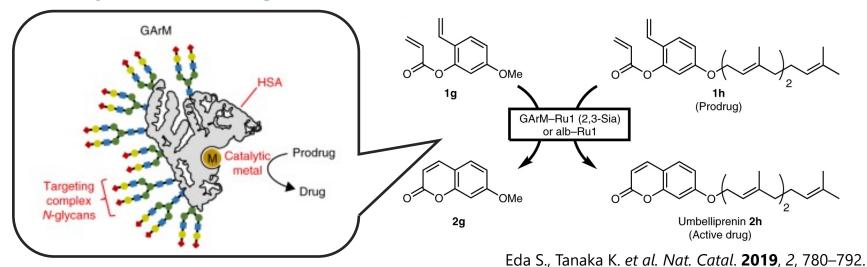


Tsubokura K., Tanaka K. *et al. Agew. Chem., Int. Ed.* **2017**, *56*, 3579–3584

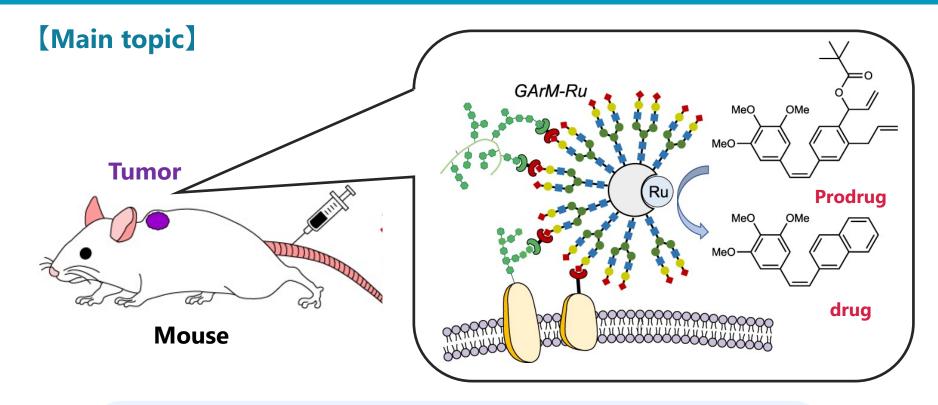


Lin Y., Tanaka K. et al. Chem. Eur. J. **2018**, 24, 10595–10600.

[Ru catalysis the biological condition (in cell)]



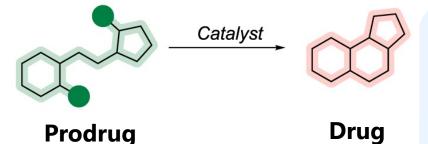
Synthetic Prodrug Strategy



- ✓ Design of synthetic prodrug to maximize its activation
- ✓ Catalytic activation of prodrug in mice
- ✓ Albumin-based artificial metalloenzyme
- ✓ N-glycosylation to target cancer cells

Retrosynthetic Prodrug Design

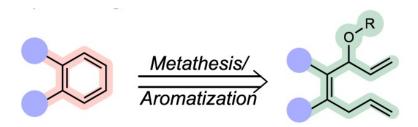
[Advantage of synthetic prodrug]



- ✓ Activation via bond forming reactions to construct a pharmacophore (drug's backbone)
- ✓ No pharmacophore in the prodrug structure
 - → Less adverse effect

[Design and optimization]

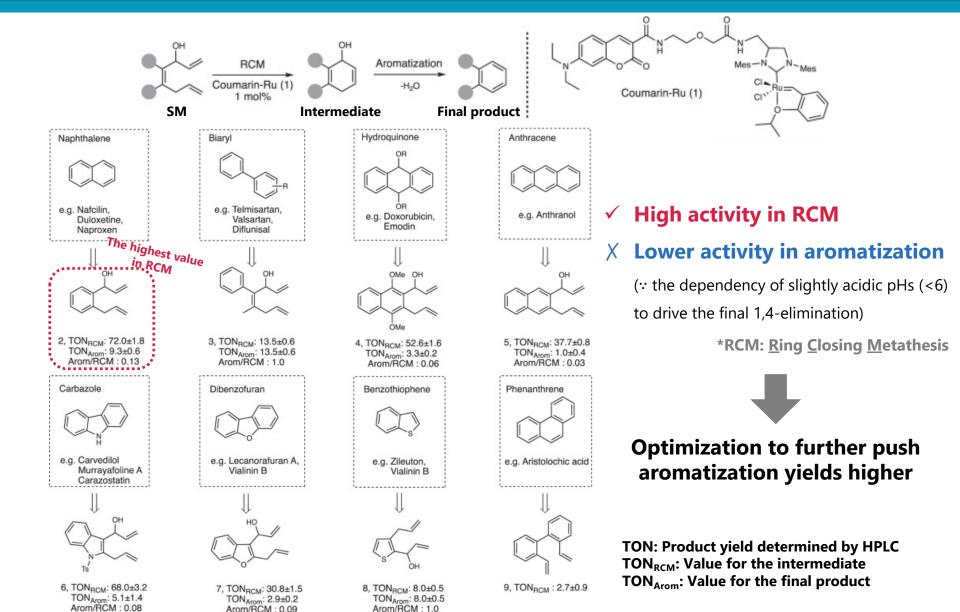
1) Finding a suitable reaction



2) Structure optimization

- Increase cascade reactivity
- Increase activity with biocatalyst
- Decrease prodrug effects
- Increase hydrolytic stability

Pharmacophore Backbone Screening

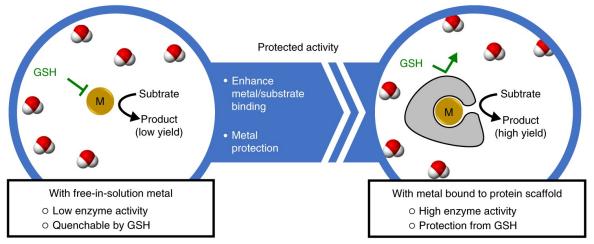


Optimization of Leaving Group

The most acidic leaving group (ester-containing precursor 12) showed both excellent RCM activity and full aromatization.

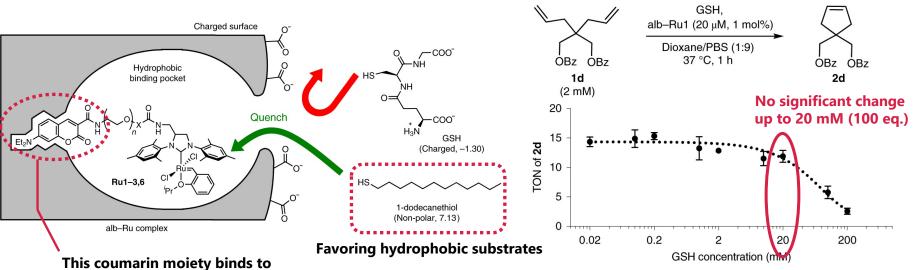
Strategy for RCM Reaction In Vivo

[Albumin-based artificial metalloenzyme (ArM)]



[Structure of metalloenzyme]

[Deactivation by glutathione]



the hydrophobic pocket of albumin. Eda S., Tanaka K.

Eda S., Tanaka K. et al. Nat. Catal. 2019, 2, 780-792.

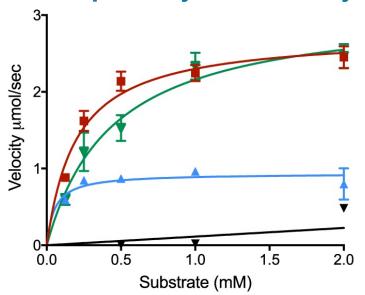
Hoveyda-Grubbs Type ArM Activity

[Reactivity in physiological conditions]

Entry	Solvent	12 conc. (mM)	Alb-Ru (mol%)	TON
1	PBS	4	1	31.1±1.1
2ª	PBS	4	1	30.8±0.1
3	PBS	2	1	34.2±0.5
4	PBS	1	1	36.3±3.7
5	D-MEM media	4	1	12.0±0.4
6	D-MEM media	2	1	5.6±0.4
7	D-MEM media	1	1	3.4±0.2

^a 20 mol% of GSH

(Substrate specificity and reactivity)

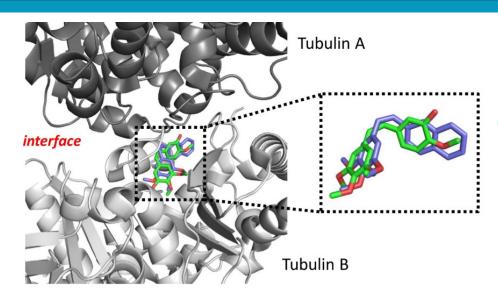


- ✓ Relatively high yield in the presence of GSH
- ✓ No adverse effect under lower conc. of substrate
- △ Lower yield in the cell growth media

14 showed the highest catalytic efficiency.

Due to the introduction of C10 terpene chain, which mimics fatty acid (good albumin ligand).

Prodrug Design

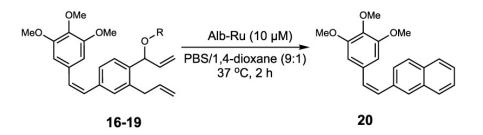


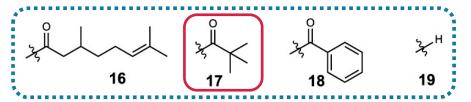
Combretastatin derivatives

- \checkmark Binding into colchicine site of β-tubulin
- ✓ Microtubule polymerization inhibitor
- Disruption of tumor growth

Prodrug Reactivity and Stability

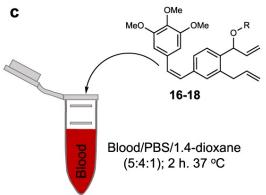
[Prodrug reactivity]





Substrate	Product yield (%)				
conc. (µM)	19	16	17	18	
5	0	28.0±1.4	12.8±0.4	4.6±0.9	
10	0	35.5±0.1	37.5±2.9	28.5±1.0	
50	3.3±0.2	47.4±0.3	57.0±0.6	44.1±2.3	
100	38.2±1.6	58.3±2.3	69.7±1.7	50.4±0.8	
250	36.7±0.5	54.6±0.7	77.0±0.1	31.4±1.6	

[Hydrolytic stability]



Substrate	Hydrolysis (%)
16	22.3±1.3
17	9.8±0.7
18	31.5±1.7

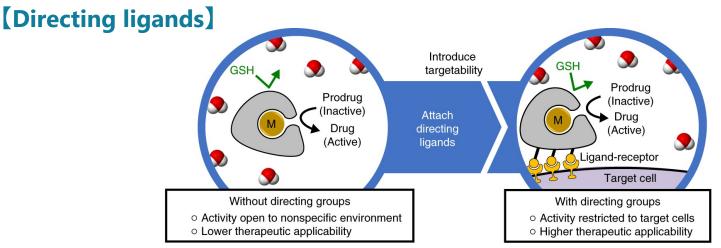
From these results, prodrug 17 (pivalate) is the best candidate moving forward.

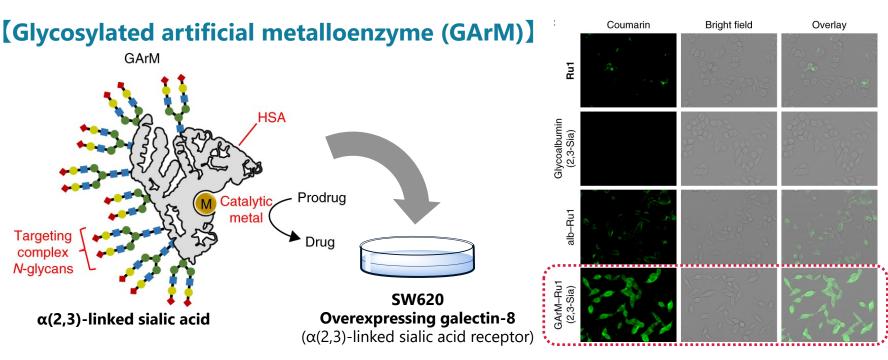


Next

Evaluation of therapeutic effect in cellulo and in vivo.

Targeting Cancer Cells

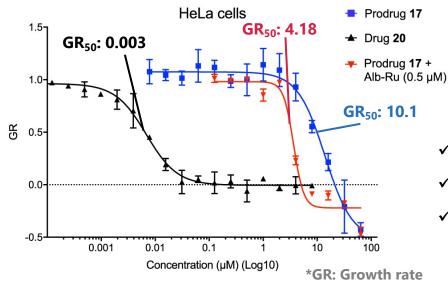


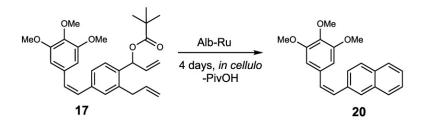


Eda S., Tanaka K. et al. Nat. Catal. 2019, 2, 780–792.

In Cellulo Study

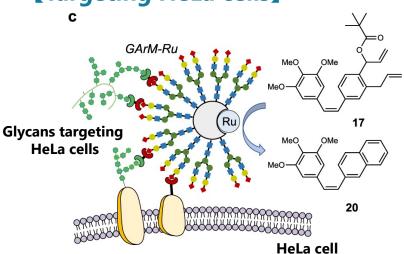
[Prodrug efficacy]

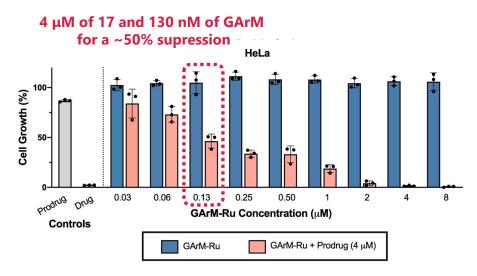




- Drug 20 showed excellent cytotoxicity (nM range)
- ✓ Prodrug 17 was less toxic (mM range)
- ✓ Prodrug activation by GArM was observed



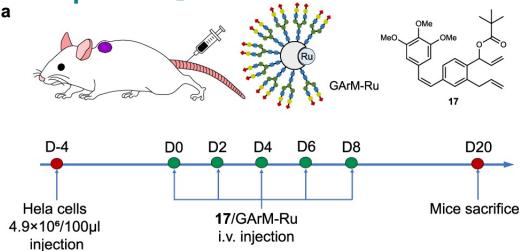




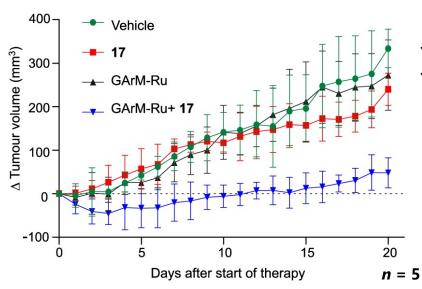
Nasibullin, I, Tanaka K. et al. Nat. Commun. 2022, 13, 1–12.

Prodrug Activation in Mice





(Tumor size in mice)



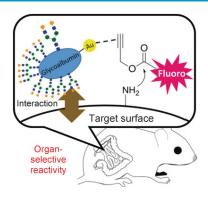
- √ 17 or GArM-Ru: No activity
- ✓ GArM-Ru + 17: Suppression of tumor growth

Pictures and the weight of tumor tissue → See appendix



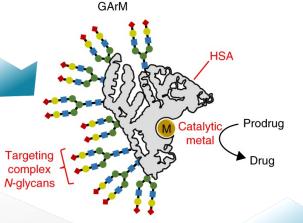
Prodrug activation via Ru-catalyzed RCM was achieved in mice!

Short Summary



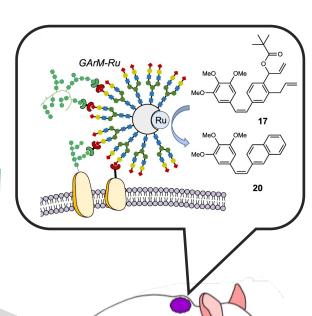
In vivo Au catalysis Targeting specific organs

Angew. Chem., Int. Ed. **2017.** Chem. Eur. J. **2018.**



Ru-catalyzed RCM in cells GArM targeting cancer cells

Nat. Catal. 2019.



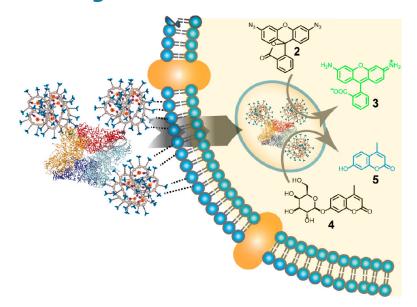
Tumor suppression via prodrug activation in vivo!!

Nat. Commun. 2022.

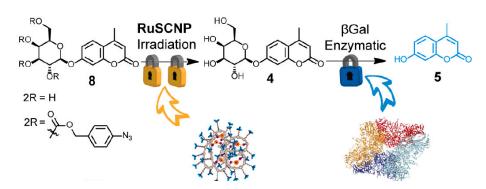
- **Introduction**
- >Applications in medicine and chemical biology
 - Cu-triggered ADC linker cleavage and reversible modification
 - Synthetic prodrug strategy for cancer treatment
 - Perspective
- **≻**Summary

Perspective

[Creating new tandem reactions]



[TM catalysis in vivo in the future...]



Tandem catalysis by Ru cat. and enzyme to provide bioactive agents in cells

J. Chen. et al. J. Am. Chem. Soc. **2020**, 142, 4565–4569.

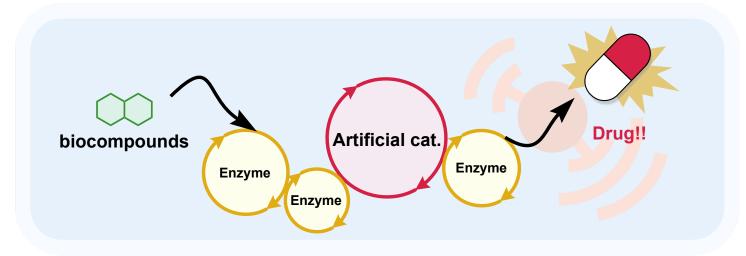
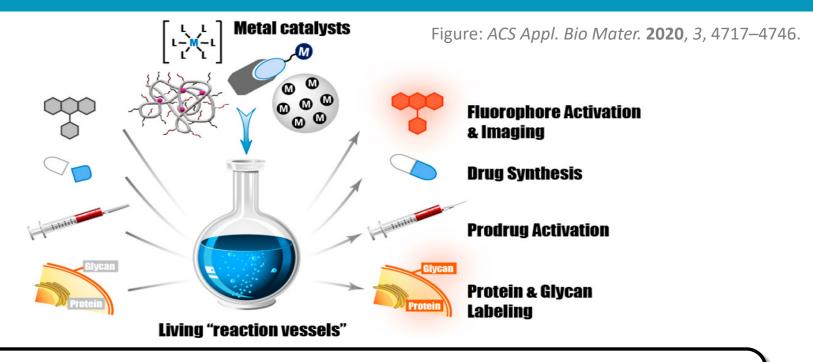


Table of Contents

- >Introduction
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 - Perspective
- **>**Summary



TM catalysis allows a great variety of reactions in vivo.



It leads to the further development of biological tools and the creation of new therapeutic modalities.

Appendix

Toxicity of Metal Compounds

[Myths of metal compounds]

- ➤ Heavy metals correspond to more toxic compounds in comparison to lighter metals
 - →This brief is ungrounded as toxic features of a metal depend on its oxidation state, ligands, etc.
- >Toxicity can be directly correlated with the structure of metal compounds.
 - →It is hardly possible to draw a direct rule for correlating the structure with toxicity.
- >All nanoparticles are toxic.
 - →As their transformations in the environment are often intricate and difficult to predict, it is hard to establish the rule of their toxicity.

^{*}For detailed information, please see the review cited below.

Toxicity of Metal Compounds

A All available compounds

21	22	23	24	manganese 25	26	27	28	29	30
Sc	Ti	V	Cr	Mn	Fe	Co	Ni	Cu	Zn
44.956	47.867	50.942	51.996	54.938	55.845	58.933	58.693	63.546	65.38
yttrium 39	zirconium 40	niobium 41	molybdenum 42	technetium 43	ruthenium 44	rhodium 45	palladium 46	silver 47	cadmium 48
Υ	Zr	Nb	Mo	Tc	Ru	Rh	Pd	Ag	Cd
88.906	91.224	92.906	95.96	[98]	101.07	102.91	106.42	107.87	112.41
	hafnium 72	tantalum 73	tungsten 74	rhenium 75	osmium 76	iridium 77	platinum 78	gold 79	mercury 80
	Hf	Ta	W	Re	Os	lr	Pt	Au	Hg
	178.49	180.95	183.84	186.21	190.23	192.22	195.08	196.97	200.59
	rutherfordium 104	dubnium 105	seaborgium 106	bohrium 107	hassium 108	meitnerium 109	darmstadtium 110	roentgenium 111	
	Rf	Db	Sg	Bh	Hs	Mt	Ds	Rg	
	[261]	[262]	[266]	[264]	[277]	[268]	[271]	[272]	

C Oxides

scandium	titanium	vanadium	chromium	manganese	iron	cobalt	nickel	copper	zinc
21	22	23	24	25	26	27	28	29	30
Sc	Ti	V	Cr	Mn	Fe	Co	Ni	Cu	Zn
44.956	47.867	50.942	51.996	54.938	55.845	58.933	58.693	63.546	65.38
yttrium 39	zirconium 40	niobium 41	molybdenum 42	technetium 43	ruthenium 44	rhodium 45	palladium 46	silver 47	cadmiur 48
Υ	Zr	Nb	Mo	Tc	Ru	Rh	Pd	Ag	Co
88.906	91.224	92.906	95.96	[98]	101.07	102.91	106.42	107.87	112.41
	hafnium 72	tantalum 73	tungsten 74	rhenium 75	osmium 76	iridium 77	platinum 78	gold 79	mercur 80
	Hf	Ta	W	Re	Os	lr	Pt	Au	Ho
	178.49	180.95	183.84	186.21	190.23	192.22	195.08	196.97	200.59
	rutherfordium 104	dubnium 105	seaborgium 106	bohrium 107	hassium 108	meitnerium 109	darmstadtium 110	roentgenium 111	
	Rf	Db	Sg	Bh	Hs	Mt	Ds	Rg	
	[261]	[262]	[266]	[264]	[277]	[268]	[271]	[272]	

B Chlorides

scandium 21	titanium 22	vanadium 23	chromium 24	manganese 25	iron 26	cobalt 27	nickel 28	copper 29	zinc 30
Sc	Ti	V	Cr	Mn	Fe	Co	Ni	Cu	Zn
44.956	47.867	50.942	51.996	54.938	55.845	58.933	58.693	63.546	65.38
yttrium 39	zirconium 40	niobium 41	molybdenum 42	technetium 43	ruthenium 44	rhodium 45	palladium 46	silver 47	cadmium 48
Y	Zr	Nb	Mo	Tc	Ru	Rh	Pd	Aq	Cd
88.906	91.224	92.906	95.96	[98]	101.07	102.91	106.42	107.87	112.41
	hafnium 72	tantalum 73	tungsten 74	rhenium 75	osmium 76	iridium 77	platinum 78	gold 79	mercury 80
	Hf	Ta	W	Re	Os	Ir	Pt	Au	Hg
	178.49	180.95	183.84	186.21	190.23	192.22	195.08	196.97	200.59
	rutherfordium 104	dubnium 105	seaborgium 106	bohrium 107	hassium 108	meitnerium 109	darmstadtium 110	roentgenium 111	
	Rf	Db	Sg	Bh	Hs	Mt	Ds	Rg	
	[261]	[262]	[266]	[264]	[277]	[268]	[271]	[272]	

D

Rating	Common description	LD ₅₀ (single oral dose for rats, mg kg ⁻¹)
1	Extremely toxic	≤1
2	Highly toxic	1-50
3	Moderately toxic	50-500
4	Slightly toxic	500-5000
5	Practically non-toxic	5000-15000
6	Relatively harmless	>15000

 Compound
 Oral LD₅₀, mg×kg⁻¹ (rat)

 CrO₃
 52

 CrCl₃
 440

 CrCl₂
 1870

 Cr(NO₃)₃×9H₂O
 3250

 Cr(acac)₃
 3360

 Cr₂O₃
 >15000

The toxicity of metal compounds can differ greatly because of differences in **solubility**, **oxidation state**, **ligands** etc.

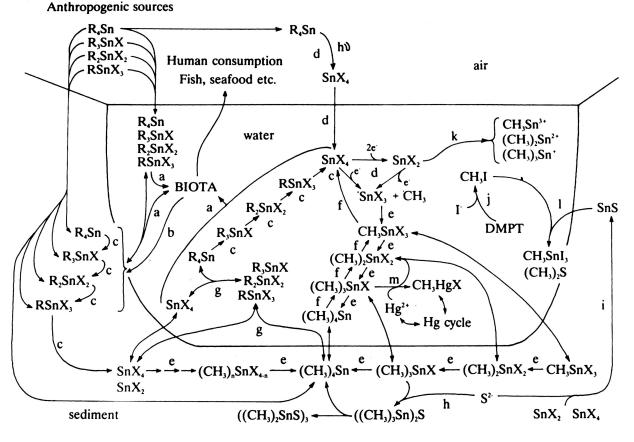


Heavy metals are not necessarily toxic than light metals.

Egorova K. S., Ananikov V. P., *Organometallics*, **2017**, *36*, 4071–4090.

Toxicity of Metal Compounds

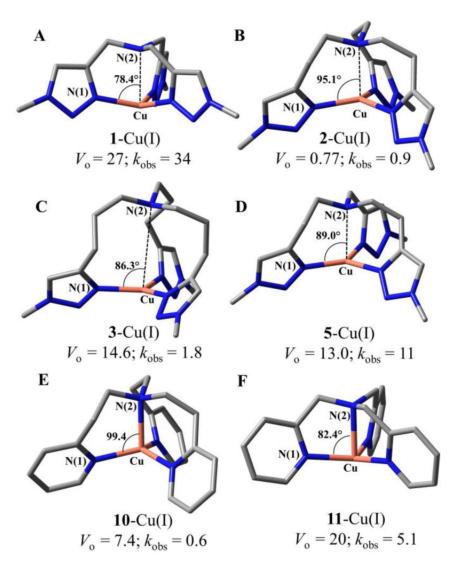
(An example: Possible biochemical transformations of organotin compounds)



Due to liability and facile reactivity of metal-containing compounds, they can undergo profound structural transformations before affecting living organisms.

→ It is difficult to identify the structure that causes the most pronounced toxicity.

Tripodal Amine Ligands for Cu(I) Complexes



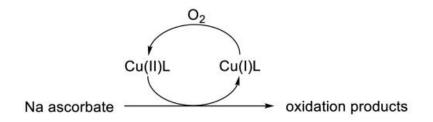
 V_o : oxidation rate

 k_{obs} : CuAAC second-order rate constant

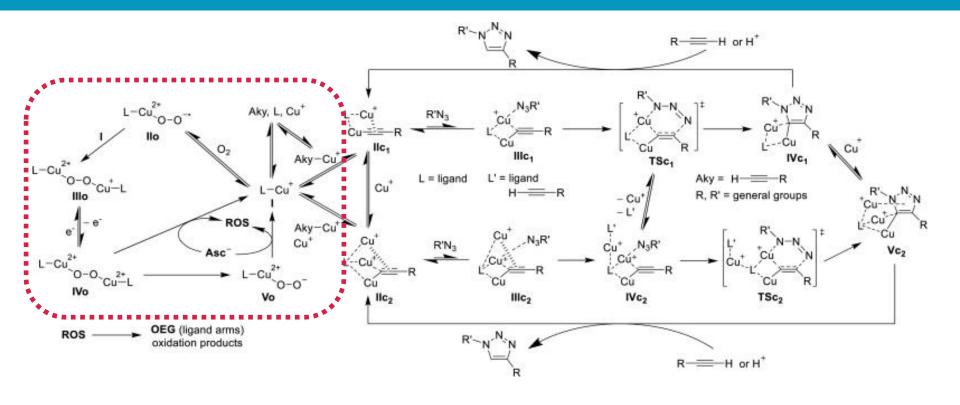
Chelate arms length of Cu(I) complexes mainly influences the angles between N(1)-Cu(I)-N(2).



Larger N(1)-Cu(l)-N(2) angle gives better protection of Cu(l) against oxidation.



Cu(I)-Triggered ROS Production



The toxicity of Cu(I)

→ Due to the production of reactive oxygen species (ROS) (Fenton's reaction is also a plausible mechanism of its toxicity.)

Biological Effects of Grubbs Catalysts

Table 1. Inhibition of TrxR, GR, trypsin, and catB by Grubbs-type catalysts. ^[a]						
	EC ₅₀ [μм]					
Compd	TrxR	GR	trypsin	catB		
G1	> 100 (53 ± 5 %)	$>$ 100 (78 \pm 7%)	> 100 (83 ± 3 %)	> 100 (75 ± 2 %)*		
G2	16.4 ± 2.3	$>$ 100 (79 \pm 5 %)	$>$ 100 (87 \pm 3%)	$>$ 100 (62 \pm 3 %)*		
HG1	3.4 ± 0.9	$>$ 100 (71 \pm 2%)	$>$ 100 (90 \pm 4%)	$\textbf{29.3} \pm \textbf{5.2*}$		
HG2	2.5 ± 0.3	$>$ 100 (80 \pm 4%)	$>$ 100 (96 \pm 2%)	$\textbf{8.0} \pm \textbf{1.2*}$		

[a] Results are expressed as mean (\pm SD) of three independent experiments. If no EC₅₀ value could be calculated, the residual enzymatic activity at 100 μ m is given in parentheses. *: insufficient solubility at 100 μ m.

Table 2. Antiproliferative effects of Grubbs-type catalysts in MCF-7 and HT-29 cells. $^{\rm [a]}$

		IC ₅₀ [µм]
Compd	MCF-7	HT-29
Cisplatin ^[b]	2.0	7.0
G1	54.8 ± 2.1	$>$ 100 (69 \pm 4%)
G2	$>$ 100 (60 \pm 3%)	$>$ 100 (94 \pm 2%)
HG1	27.8 ± 1.4	$>$ 100 (76 \pm 3%)
HG2	9.9 ± 3.7	13.4 ± 4.4

[a] Results are expressed as mean (\pm SD) of three independent experiments. If no IC₅₀ value could be calculated, the percentage of cell biomass (compared with an untreated control) at 100 μ M of the compound is given in parentheses. [b] IC₅₀ values for cisplatin in the same assay are given as a reference (data taken from Ref. [25, 30]).

Grubbs-type catalysts have potential as inhibitors of tumor-relevant enzymes, exhibit antiproliferative effects in cultured tumor cells, and influence cell metabolism.

ADCs Linker Cleavage

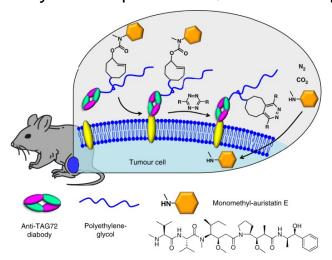
Cleavable linkers so far

Current cleavable linkers rely on...

- Redox cleavage
- Enzyme-mediated cleavage
- photocleavage

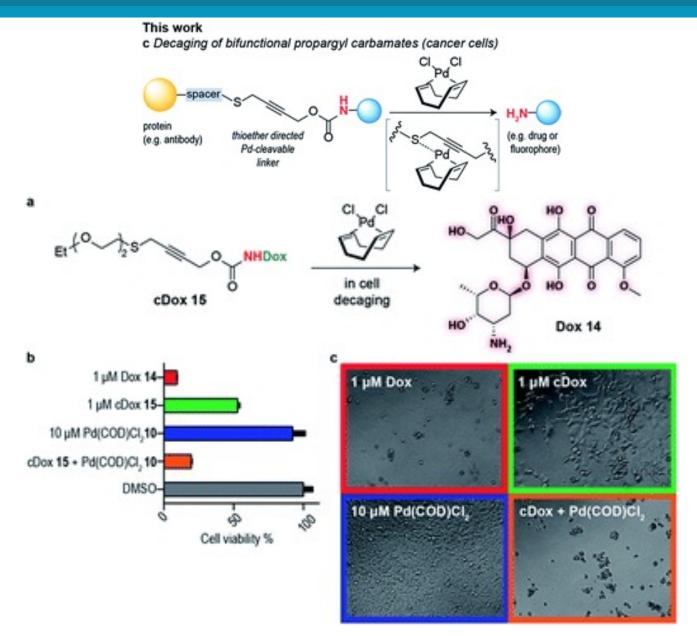
These are not amendable for bioorthogonal control over the activation process.

- Transcyclooctene (TCO)-based linker
 - →It requires a sophisticated synthesis procedure, which hampers large-scale production.



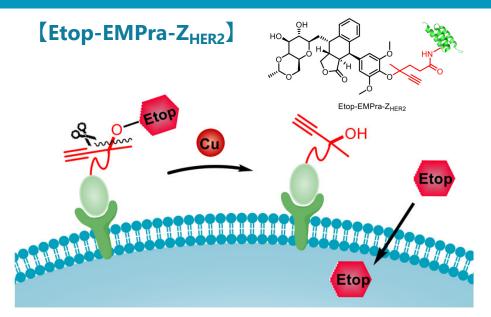
(TCO)-based linker

Another Example of ADC Cleavage by TM



B. J. Stenton, et al. Chem. Sci. 2019, 9, 4185–4189.

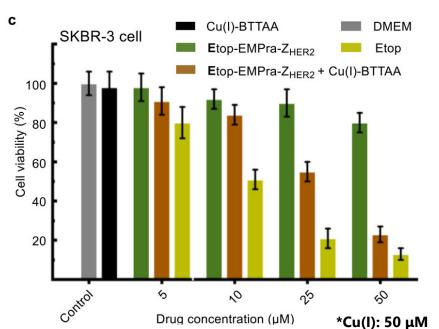
Cu-Triggered ADCs Linker Cleavage (Etop)



Etoposide

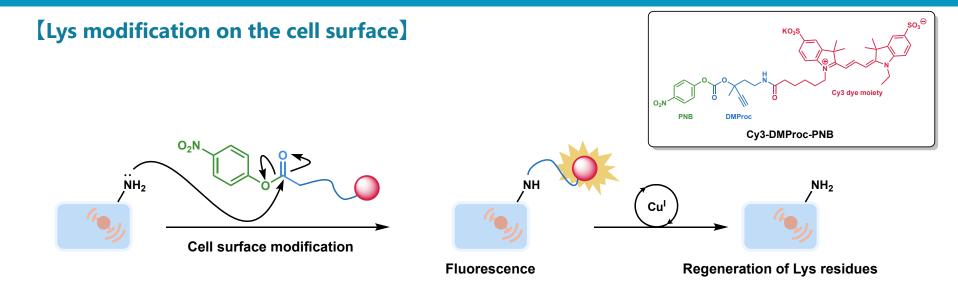
- Anticancer drug
- Topoisomerase II inhibitor → DNA damage
- X Side effects due to the lack of selectivity

Cu-mediated release of phenols instead of amines



Cu-catalyzed, on demand and on target ADC activation strategy was also demonstrated in Etop-EMPra-Z_{HER2}.

Cell Surface Modification



[Fluorescent images]

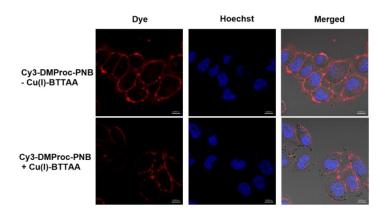
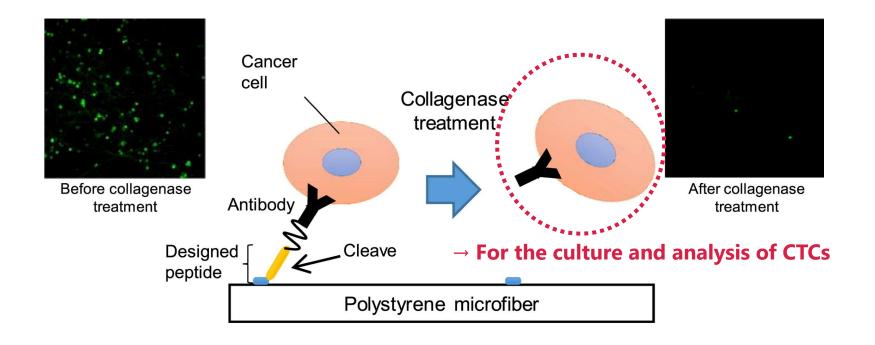


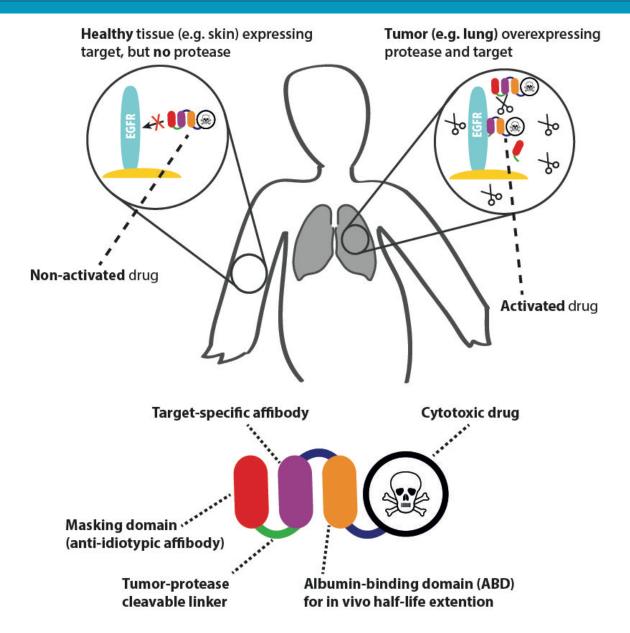
Figure S8. Fluorescent imaging of the release of Cy3 by Cu(I)-BTTAA on living cells. The copper-triggered cleavage of the internal linker resulted in the decreased fluorescence of Cy3. Scale bar: $100 \ \mu m$

Capture and Release of CTCs

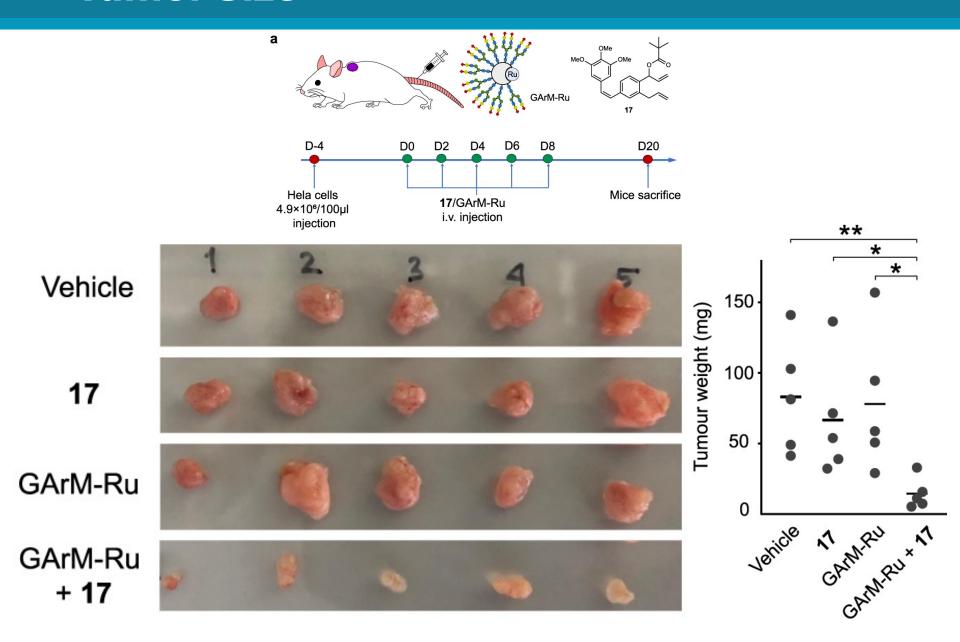


- ✓ CTCs (Circulating Tumor Cells), which are tumor cells present in the blood, have attracted much attention as a new tumor marker.
- ✓ The capture and release system will be useful for further CTCs analysis.
- ✓ This system can be applicable for more accurate cancer diagnosis.

Protein-Based Prodrug



https://www.kth.se/profile/lofblom/page/protein-based-prodrugs



Nasibullin, I, Tanaka K. et al. Nat. Commun. 2022, 13, 1-12.

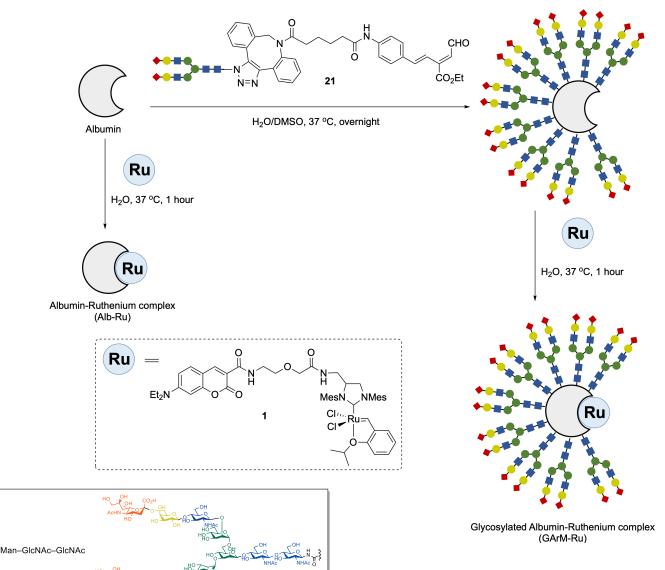
Preparation of ArM

where:

Sia-Gal-GlcNAc-Man

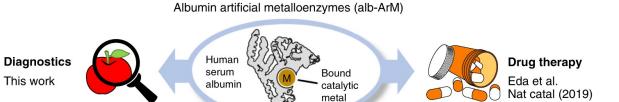
Sia-Gal-GlcNAc-Man-

Supplementary Figure 1. Structure of the $\alpha(2,6)$ -Sia terminated glycan-aldehyde probe 21



Nasibullin, I, Tanaka K. et al. Nat. Commun. 2022, 13, 1–12.

ArM as an Ethylene Sensor



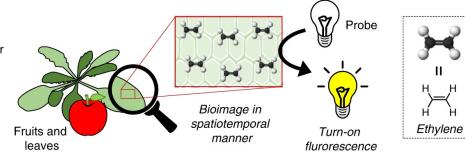
ArM-based ethylene detection in plants:

(AEP)

GOAL

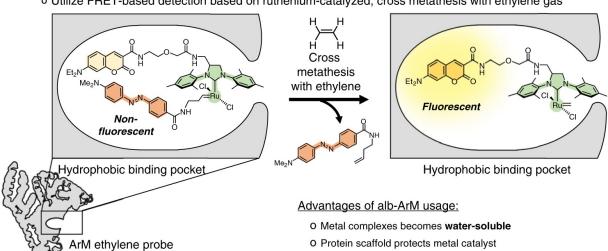
This work

 To create an enzyme biosensor that can detect ethylene in a spatiotemporal manner



STRATEGY

o Utilize FRET-based detection based on ruthenium-catalyzed, cross metathesis with ethylene gas



Bong, K., Tanaka K. et al. Nat. Commun. 2019, 10, 1–15.

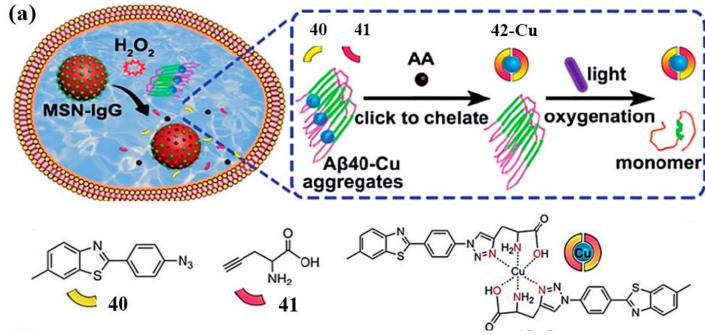
Drugs Possessing Propargyl Group

Transition metal catalyst



Causing a new drug interaction(?)

Cu catalysis in $A\beta$ plaques



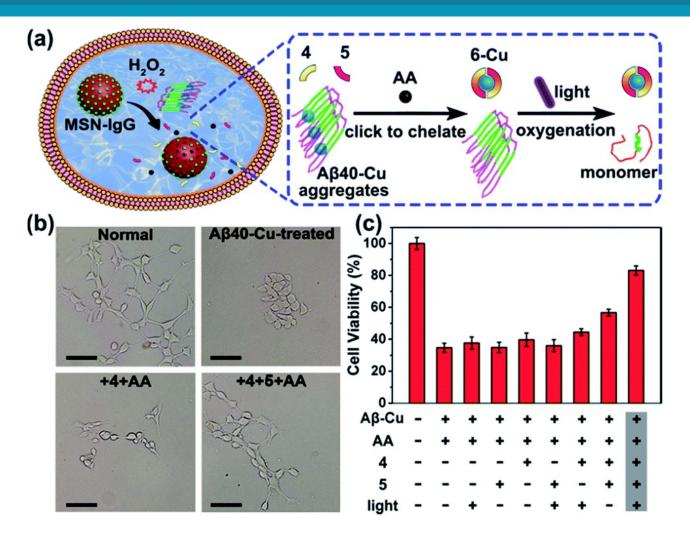
Azide-alkyne cycloaddition catalyzed by endogenous copper accumulated in Aβ plaques.



 $A\beta$ photo-oxygenation & Cu extraction from $A\beta$ -Cu aggregates

→The synergistic effects promoting the disassembly of Aβ-Cu aggregates!!

Cu catalysis in $A\beta$ plaques



- √ 40+41+ascorbic acid(AA) → cell morphology was restored.
- ✓ After UV irradiation (photo-oxygenation), cell viability further improved.