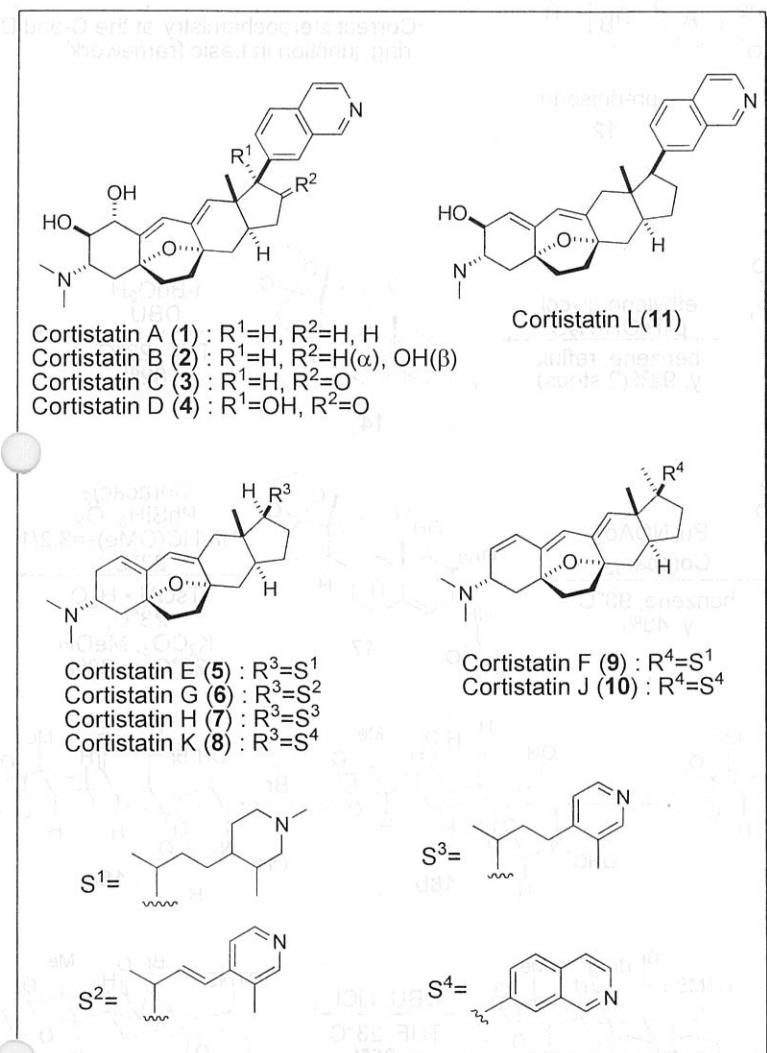
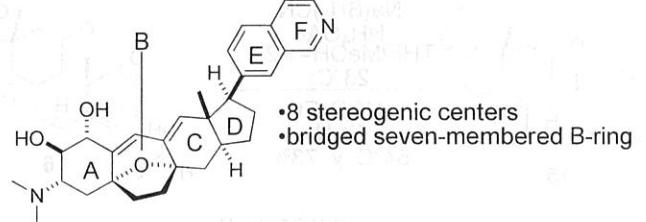


# Cortistatins

## Total Synthesis and Their Structure-Activity Relationship

**Isolation**M. Kobayashi et al. *JACS*, 2006, 128, 3148**Semisynthesis**P. S. Baran et al. *JACS*, 2008, 130, 7241**Total synthesis**M. D. Shair et al. *JACS*, 2008, 130, 16864K. C. Nicolaou et al. *Angew. Chem. Int. Ed.* 2008, 47, 7310**Formal total synthesis**M. Hirama et al. *Tetrahedron Lett.* 2009, 50, 3277R. Sarpong et al. *Tetrahedron* In press**Other studies for the total synthesis**S. J. Danishefsky et al. *Tetrahedron Lett.* 2008, 49, 6610E. J. Corey et al. *Org. Lett.* 2008, 10, 5247P. Magnus et al. *Org. Lett.* 2009, 11, 3938B. W. Gung et al. *Chem. Eur. J.* 2010, 16, 639

**Table 1.** Selective Growth Inhibition of Cortistatins against HUVECs<sup>a</sup>

cell line	1		2		3		4	
	IC <sub>50</sub>	S.I.						
HUVECs	0.0018	1	1.1	1	0.019	1	0.15	1
KB3-1	7.0	3900	120	110	150	7900	55	460
Neuro2A	6.0	3300	160	150	180	9500	>300	nd
K562	7.0	3900	200	180	>300	nd	>300	nd
NHDF	6.0	3300	>300	nd	>300	nd	>300	nd

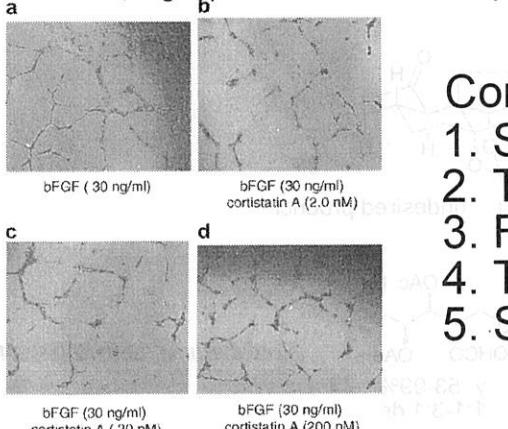
<sup>a</sup> IC<sub>50</sub> =  $\mu\text{M}$ ; nd = not determined; S.I. = selective index: IC<sub>50</sub> against testing cells/IC<sub>50</sub> against HUVECs.

Cortistatins are first isolated from marine sponge *Corticium simplex* in 2006, as a selective inhibitor of the proliferation of HUVECs(human umbilical vein endothelial cells).

The most potent member of the family (IC<sub>50</sub>=1.8nM), cortistatin A (1) demonstrated a selectivity index of more than 3000 against HUVECs in comparison with normal human dermal fibroblast(NHDF) and several other tumor cells(KB3-1, K562, and Neuro2A).

After that, cortistatin was found to inhibit not only proliferation, but also migration of HUVECs, thereby inhibited tubular formation. What outstanding is that cortistatins are not cytotoxic but cytostatic, probably through reversible binding to the target protein.

Cortistatins are thought to be promising drug candidates or leads for the diseases related to angiogenesis. These biological properties coupled with its unprecedented molecular architecture made cortistatin A a target for chemical synthesis. As a result of extensive studies, 5 groups have succeeded in its synthesis so far.

**Contents**

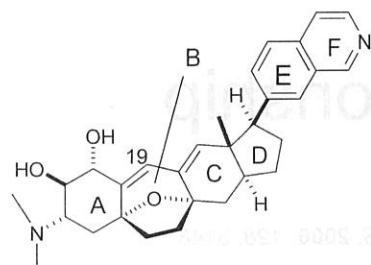
- |                                     |       |
|-------------------------------------|-------|
| 1. Semisynthesis by Baran           | 2-3   |
| 2. Total Synthesis by Nicolaou      | 4-6   |
| 3. Formal Total Synthesis by Himura | 7-8   |
| 4. Total Synthesis by Shair         | 9-10  |
| 5. Structure-Activity Relationship  | 11-14 |

# 1. Semisynthesis by Baran

P. S. Baran et al. JACS, 2008, 130, 7241

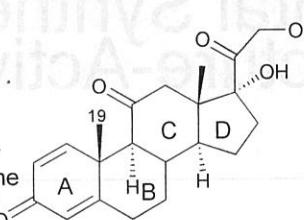
Retrosynthetic Analysis

## Retrosynthesis



Cortistatin A (1)

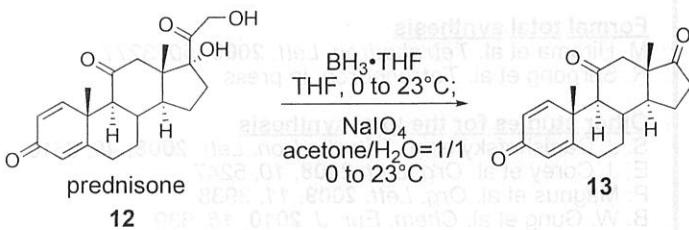
1. A-ring functionalization
2. B-ring expansion
3. Oxidation at C19
4. coupling with isoquinoline
5. functional group manipulation



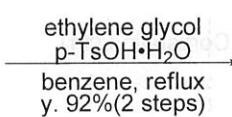
prednisone  
12

- Prednisone is commercially available at \$1.20/g and possesses 70% of carbon atoms.

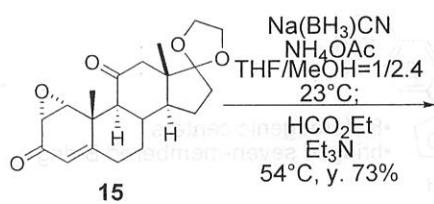
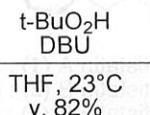
- Correct stereochemistry at the C-and D-ring junction in basic framework.



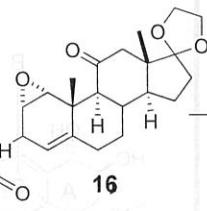
12



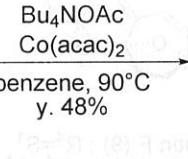
14



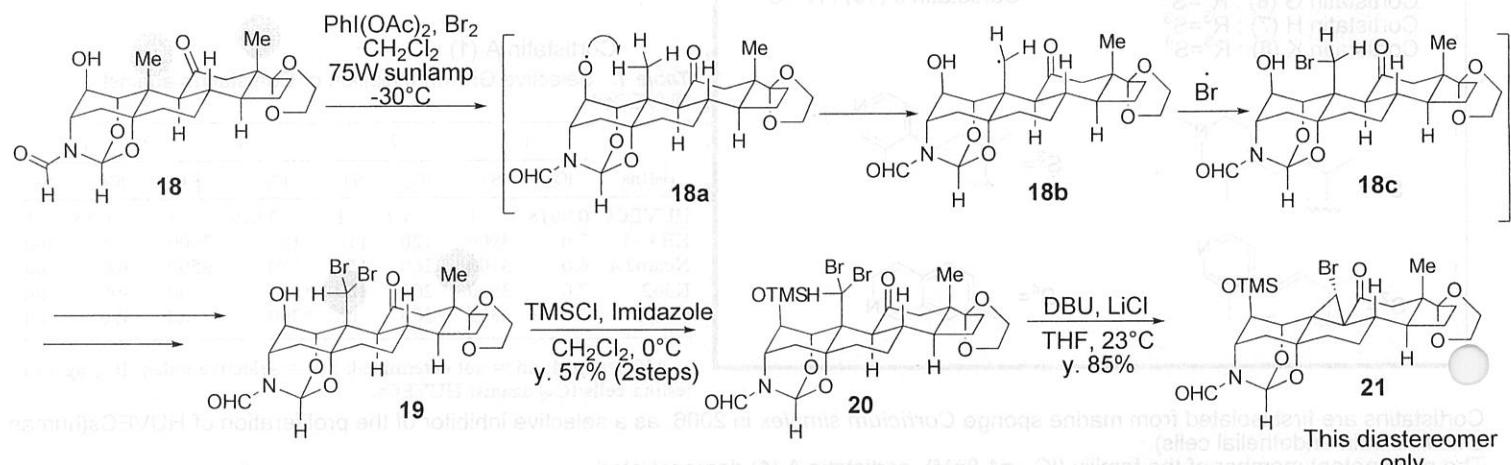
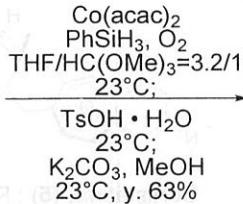
15



16

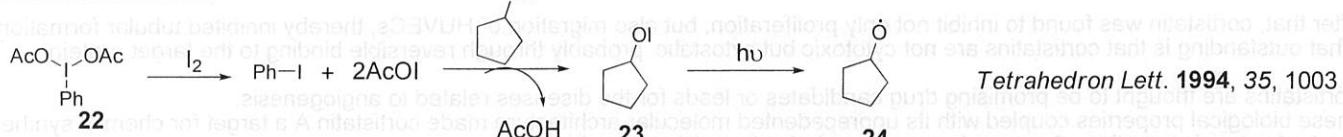


17



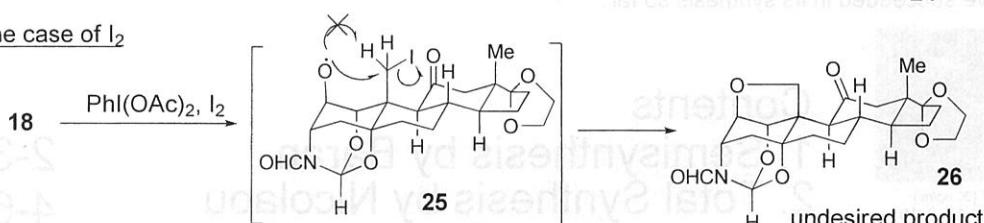
This diastereomer only

## mechanism of radial formation

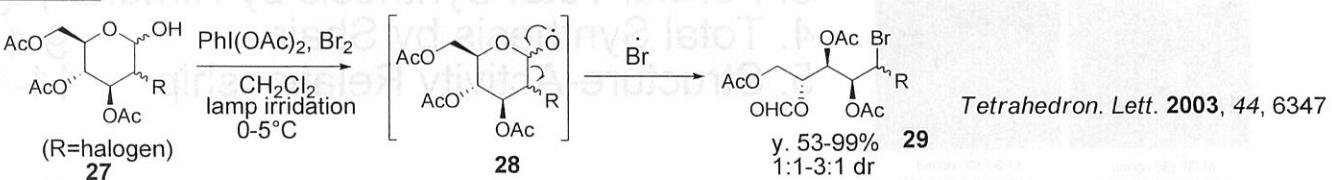


Tetrahedron Lett. 1994, 35, 1003

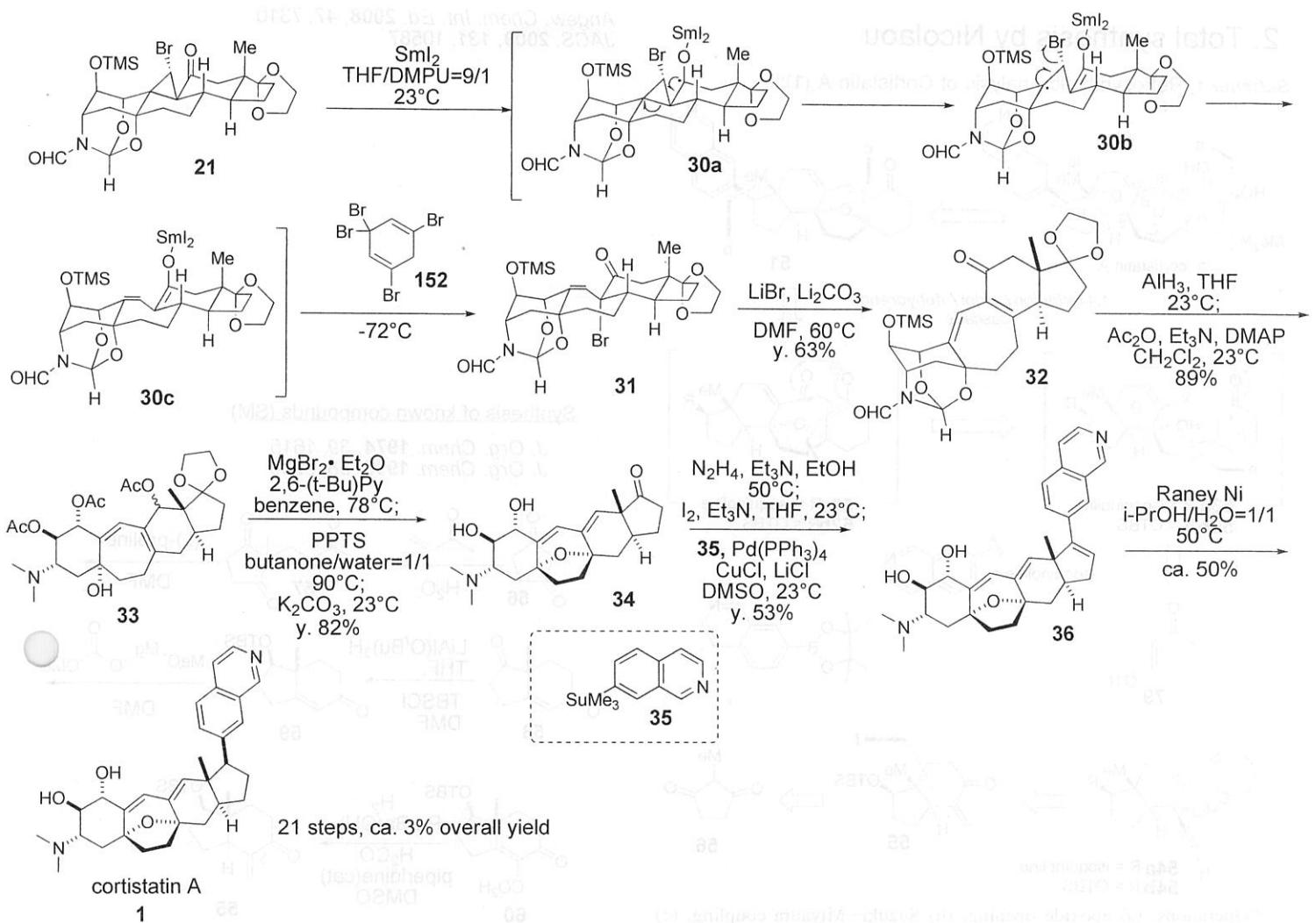
in the case of I2



## a precedent

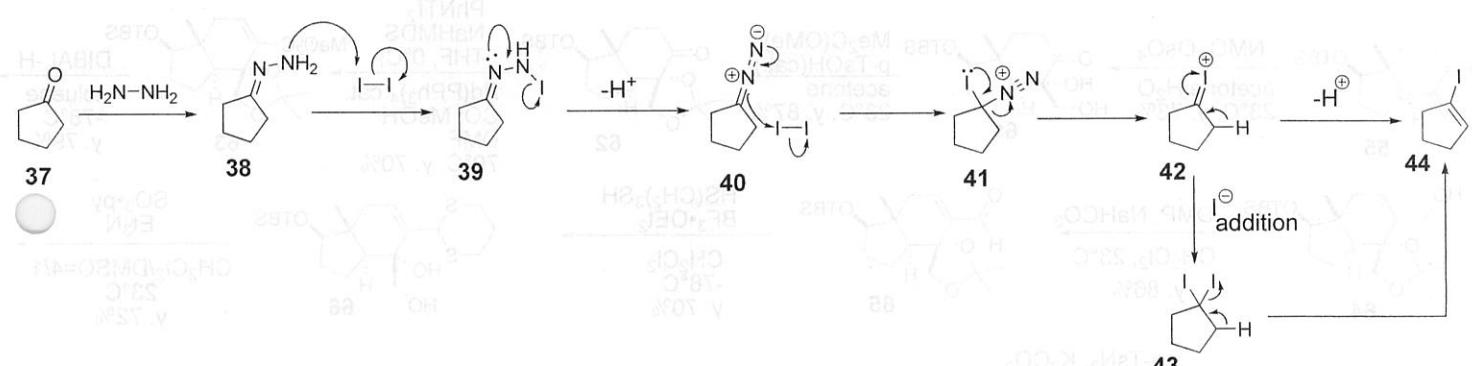


Tetrahedron Lett. 2003, 44, 6347

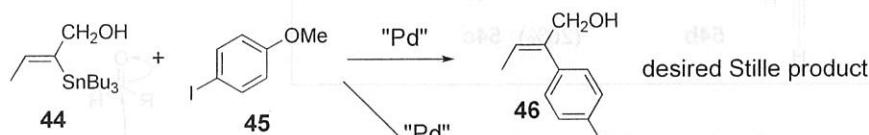


#### Generation of vinyl iodide from ketone

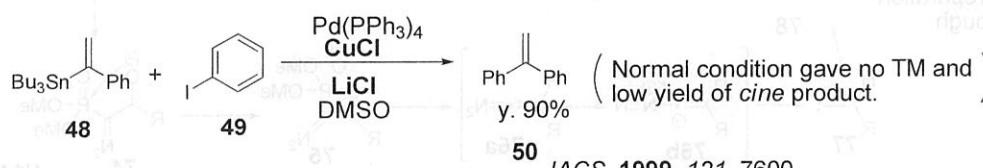
J. Chem. Soc. 1962, 470



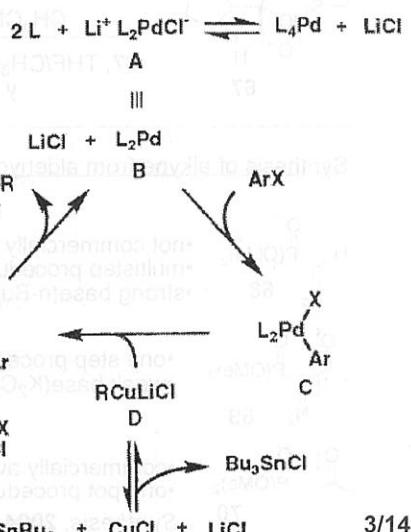
#### Stille coupling with sterically hindered 1-substituted vinylstannanes



Tetrahedron Lett. 1998, 39, 5177



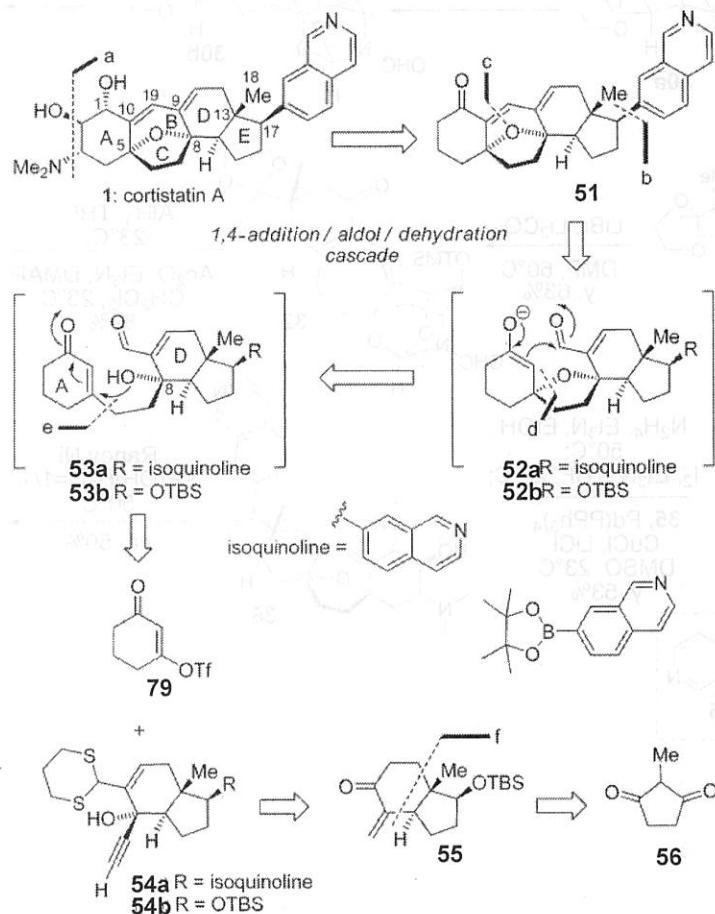
#### Proposed Catalytic Cycle



## 2. Total synthesis by Nicolaou

Angew. Chem. Int. Ed. 2008, 47, 7310  
JACS, 2009, 131, 10587

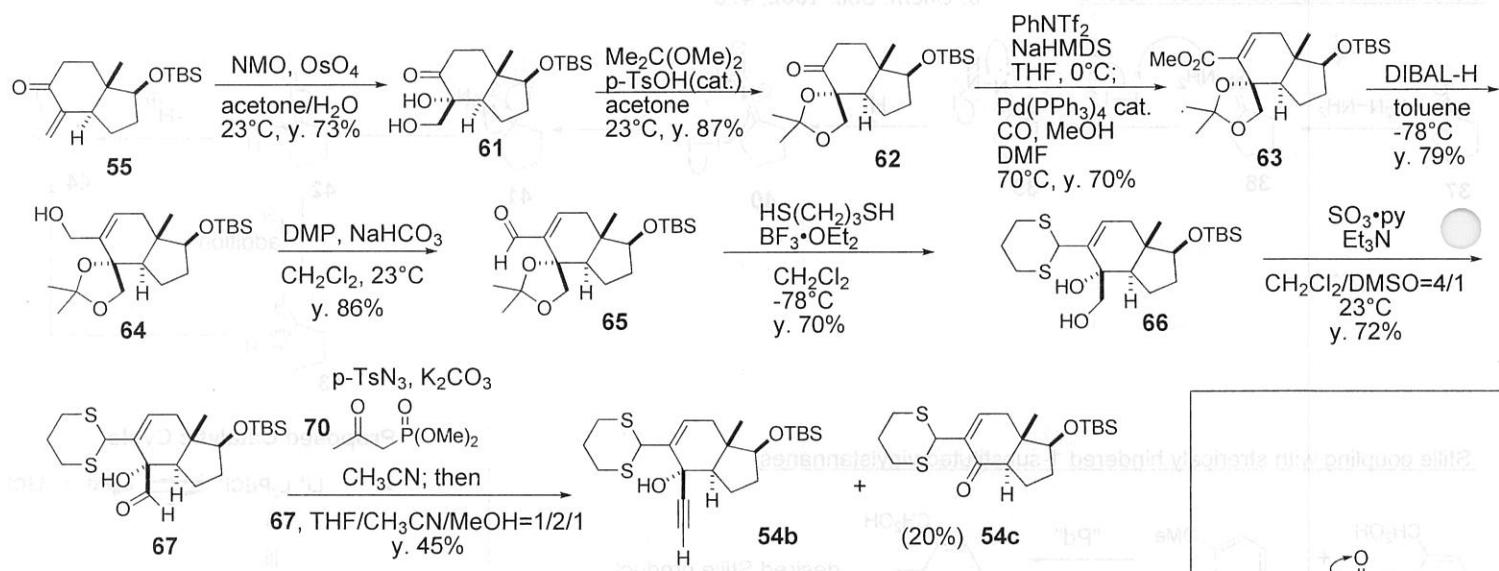
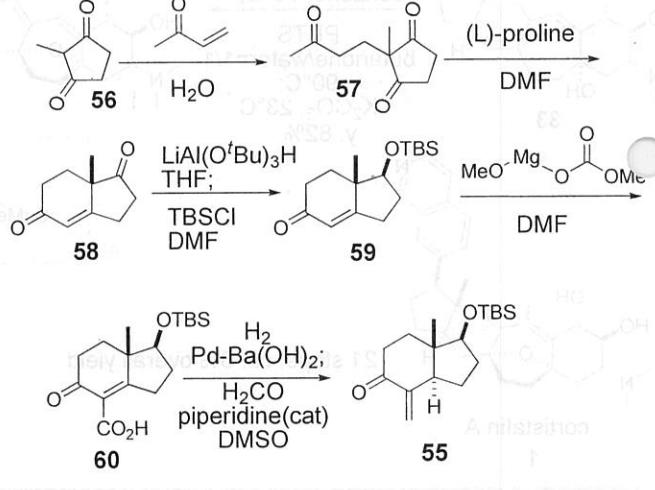
**Scheme 1.** Retrosynthetic Analysis of Cortistatin A (1)<sup>a</sup>



<sup>a</sup> Operations: (a) epoxide opening; (b) Suzuki–Miyaura coupling; (c) aldol condensation; (d) 1,4-addition; (e) Sonogashira coupling; (f) Hajos–Parrish ketone construction.

### Synthesis of known compounds (SM)

J. Org. Chem. 1974, 39, 1615  
J. Org. Chem. 1993, 58, 3938

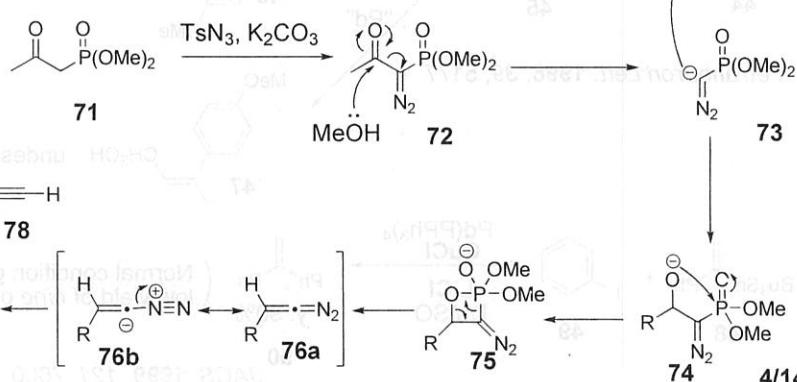


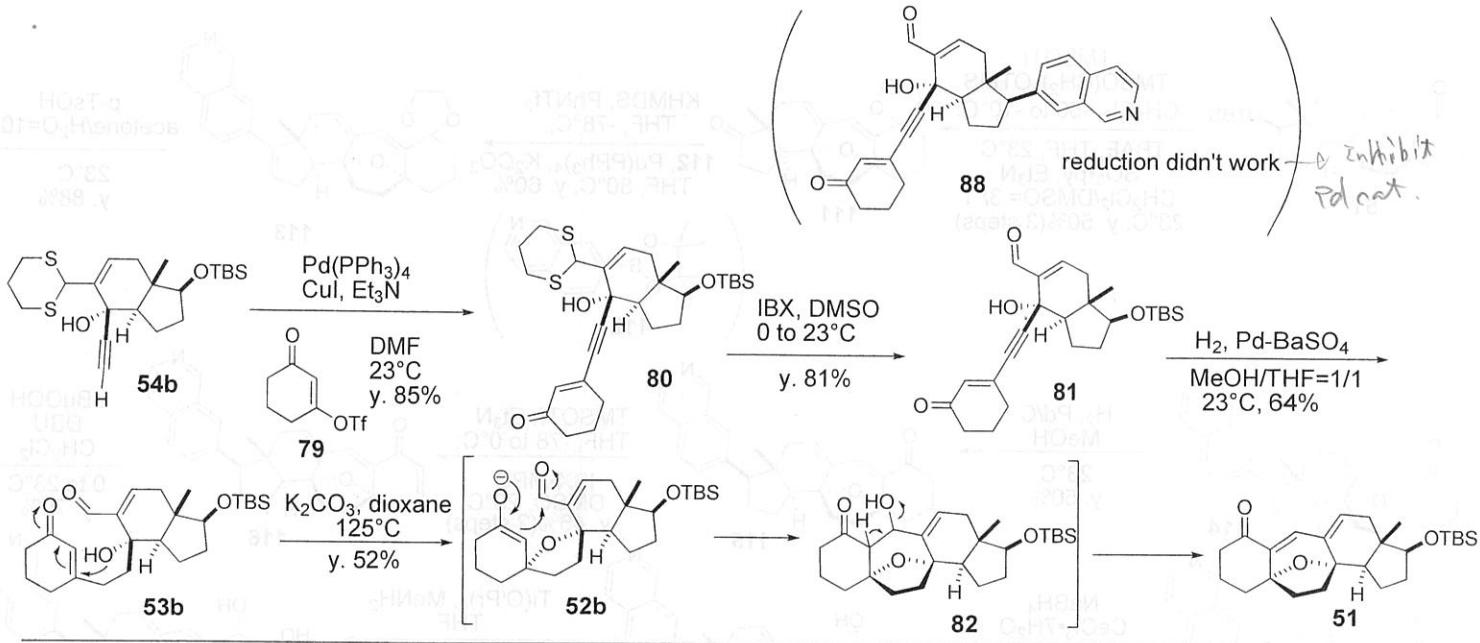
### Synthesis of alkyne from aldehyde

68 •not commercially available  
•multistep procedures for preparation  
•strong base(*n*-BuLi) is required

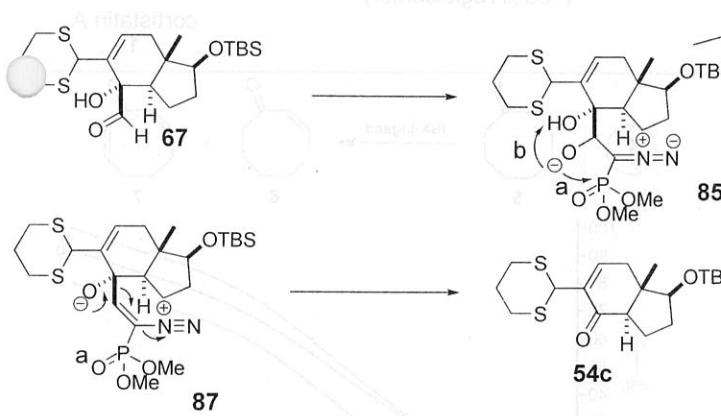
69 •one step procedure for preparation  
•weak base(K<sub>2</sub>CO<sub>3</sub>) is enough

70 •commercially available  
•one pot procedure  
*Synthesis*, 2004, 59

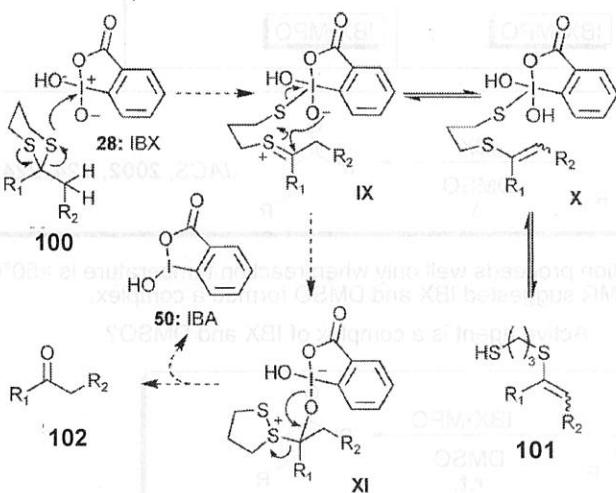




67 → 54c

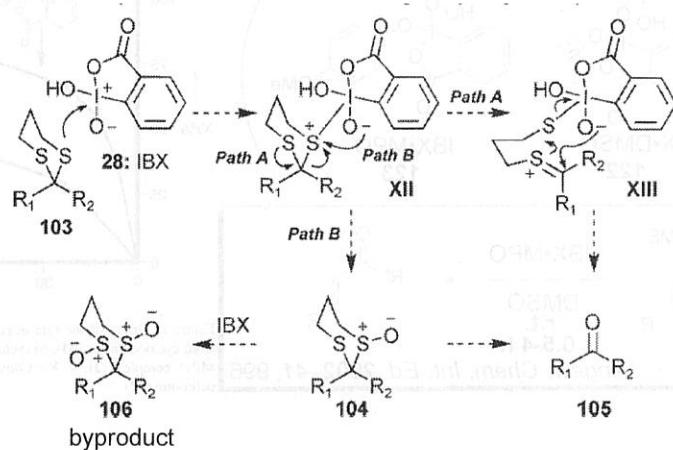
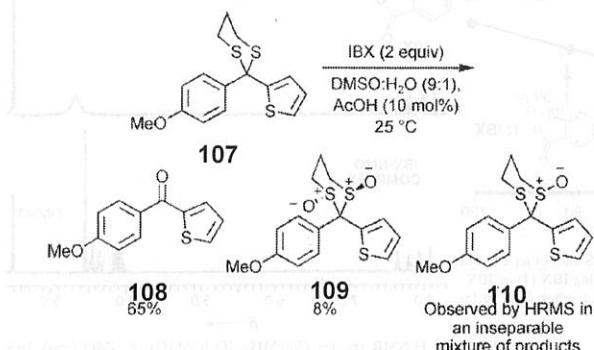


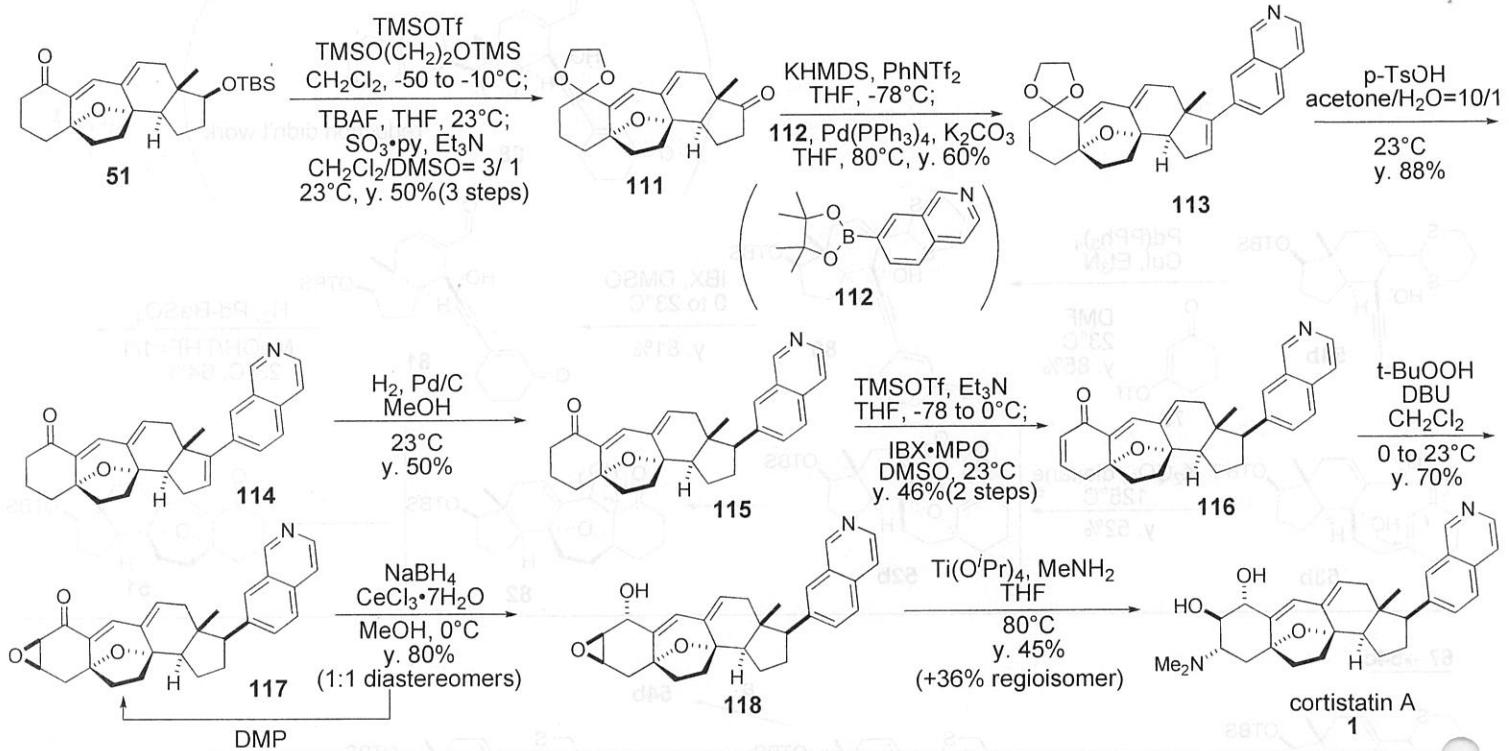
### Mechanism



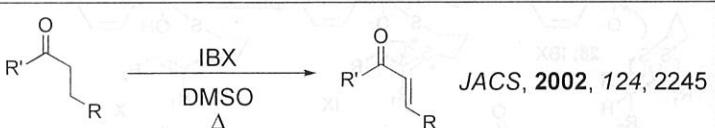
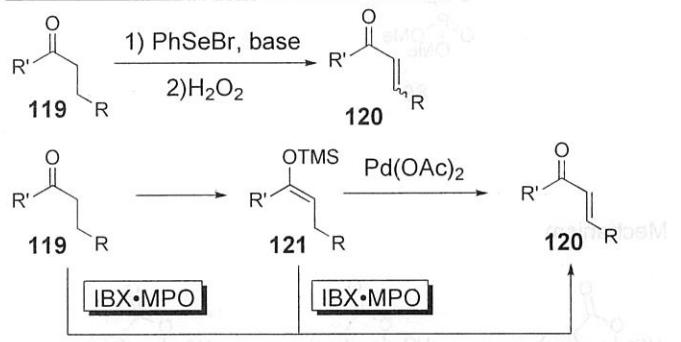
reaction mechanism of sterically hindered substrates

### reactions of sterically hindered substrates (lacking $\alpha$ -protons)





#### Synthesis of enone from saturated ketone



• Reaction proceeds well only when reaction temperature is >50°C  
 • 1H-NMR suggested IBX and DMSO formed a complex.

→ Active agent is a complex of IBX and DMSO?

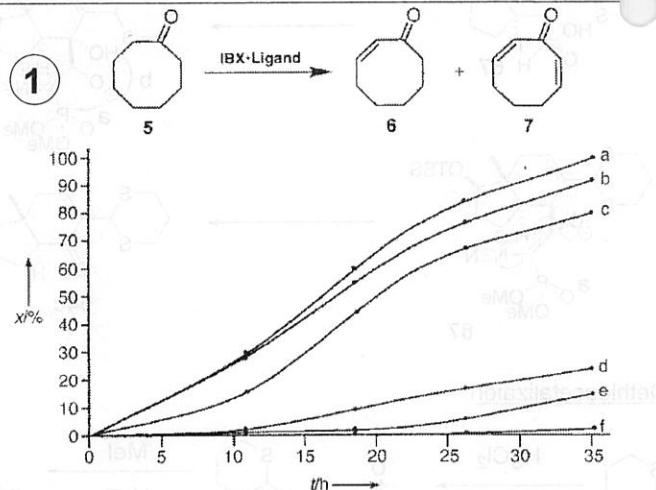
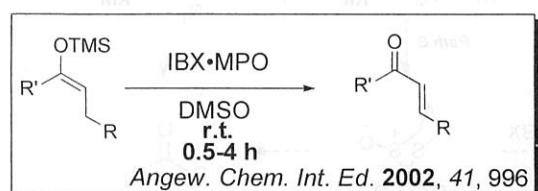
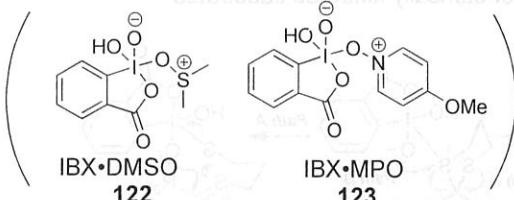
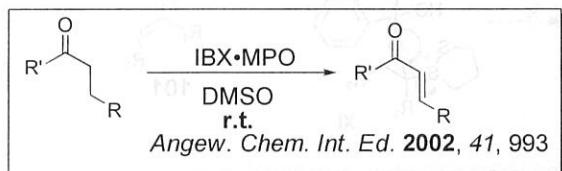


Figure 1. Graph of the rate of conversion of cyclooctanone (5) into dehydrogenated products (*cis* enone 6 and dienone 7) over time, for different IBX-ligand systems: a) IBX·MPO; b) IBX·NMO; c) IBX·tri-methylamine-*N*-oxide; d) IBX·DMSO formed by dissolution at 75°C for 30 min; e) IBX·DMSO formed by dissolution at 25°C; f) IBX·DMSO formed by dissolution at 90°C for 20 min (leads to significant quantities of IBA from the reduction of IBX by DMSO which inhibits the desired reaction<sup>[2]</sup>). Reagents and conditions: IBX/ligand 1:1 (2.2 equiv), [D<sub>6</sub>]DMSO, 25 °C, conversion monitored by means of <sup>1</sup>H NMR spectroscopy. x = Total dehydrogenation.

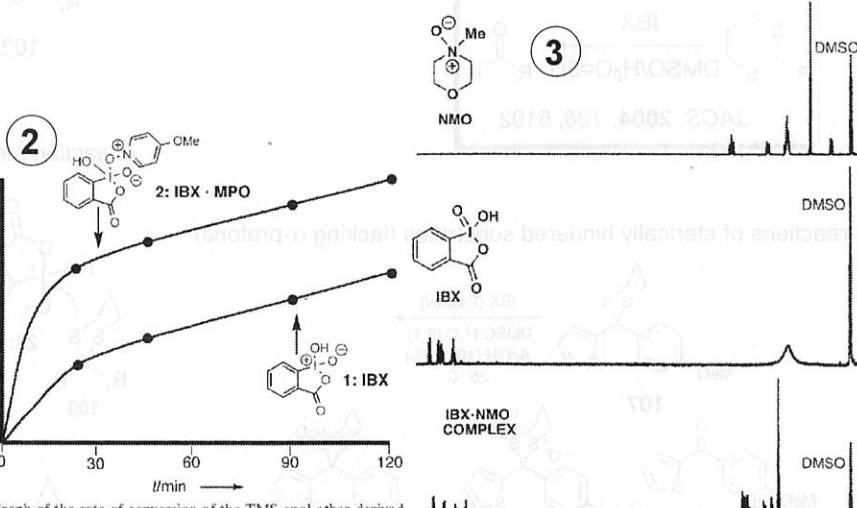
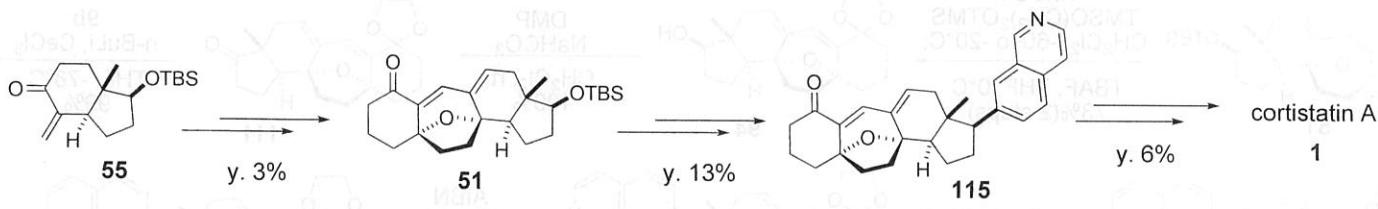


Figure 1. Graph of the rate of conversion of the TMS enol ether derived from cyclooctanone (31), to cyclooctenone (32) by using IBX (1) or IBX·MPO complex (2).<sup>[2]</sup> X = Conversion of TMS enol ether into cyclooctenone (32).

Figure 2. <sup>1</sup>H NMR spectra (500 MHz, [D<sub>6</sub>]DMSO) of NMO (top), IBX (center), and the observed NMO-IBX complex (bottom).

Althouth total synthesis was accomplished, overall yield was only 0.03%\*\*\*

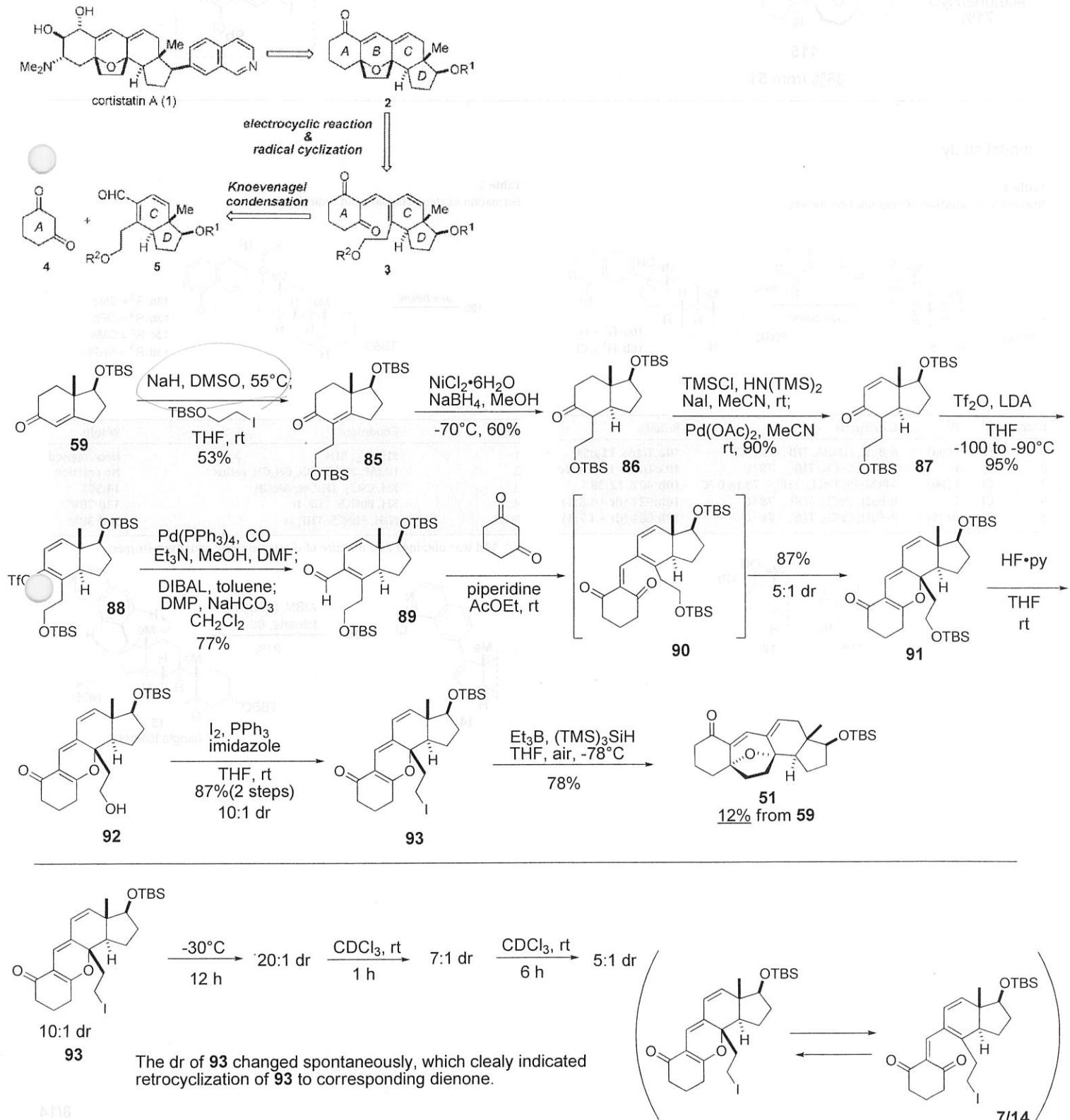


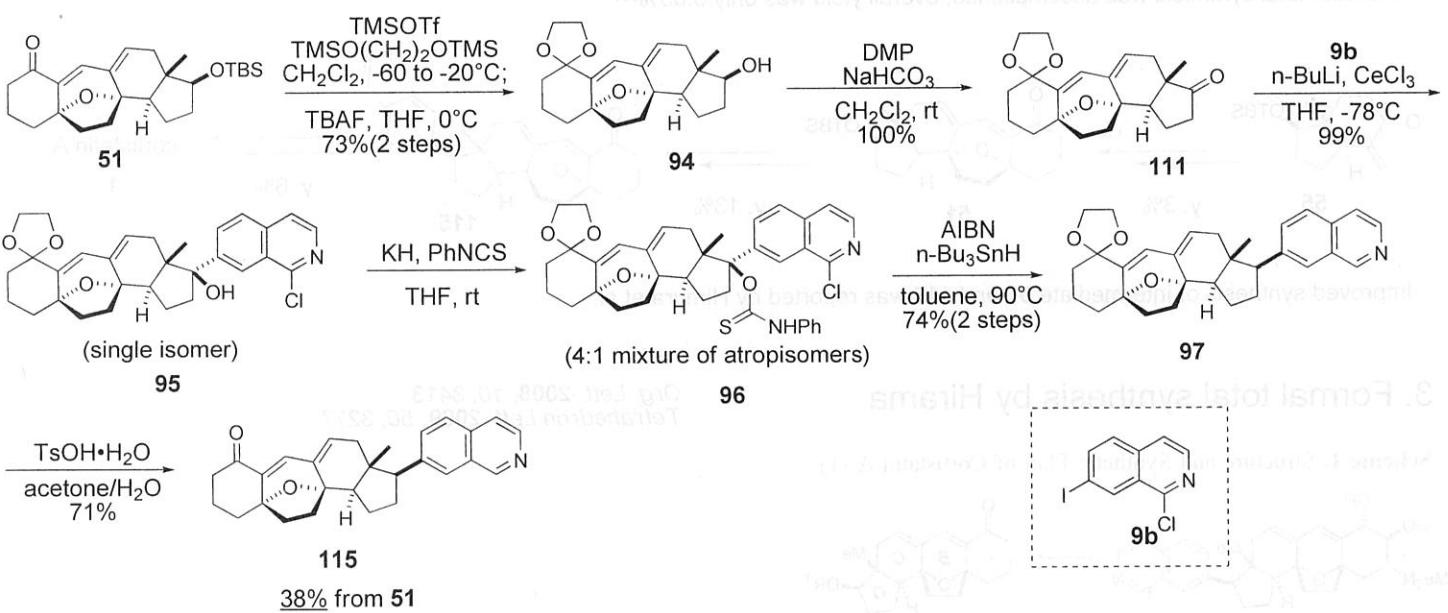
Improved synthesis of intermediate 51 and 115 was reported by Himura et al.

### 3. Formal total synthesis by Hirama

*Org. Lett.* **2008**, *10*, 3413  
*Tetrahedron Lett.* **2009**, *50*, 3277

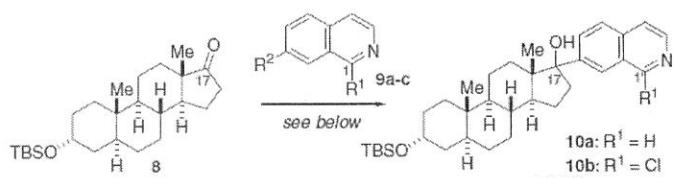
Scheme 1. Structure and Synthetic Plan of Cortistatin A (1)





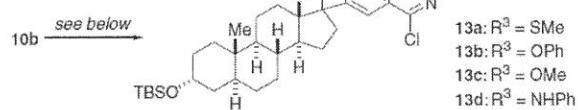
### model study

**Table 1**  
Nucleophilic addition of isoquinoline moiety



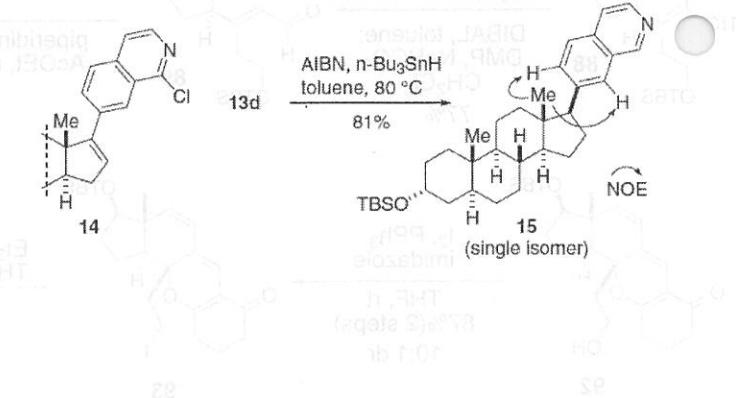
Entry	R <sup>1</sup>	R <sup>2</sup>	Conditions	Results
1	H	I (9a)	n-BuLi, HMPA, THF, -78 °C	10a: Trace, 11a: 5%
2	H	I	n-BuLi, CeCl <sub>3</sub> , THF, -78 °C	10a: Trace, 11a: Trace
3	Cl	I (9b)	i-PrMgBr, CeCl <sub>3</sub> , THF, -78 to 0 °C	10b: 40%, 12: 38%
4	Cl	I	n-BuLi, CeCl <sub>3</sub> , THF, -78 °C	10b: 92% (dr = 1.8:1)
5	Cl	Br (9c)	n-BuLi, CeCl <sub>3</sub> , THF, -78 °C	10b: 68% (dr = 1.7:1)

**Table 2**  
Formation of thiocarbamate and radical reduction



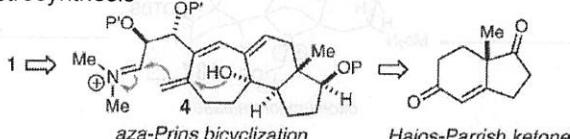
Entry	Conditions	Results
1	KH, CS <sub>2</sub> ; MeI	Decomposed
2	DMAP, PhOC(S)Cl, CH <sub>3</sub> CN, reflux	No reaction
3	KH, CCl <sub>4</sub> , THF, rt; MeOH	14:50%
4	KH, PhNCS, THF, rt	13d: 70% <sup>a</sup>
5	NaH, PhNCS, THF, rt	13d: 30%

<sup>a</sup> 13d was obtained as a mixture of diastereomeric and atrop-isomers.

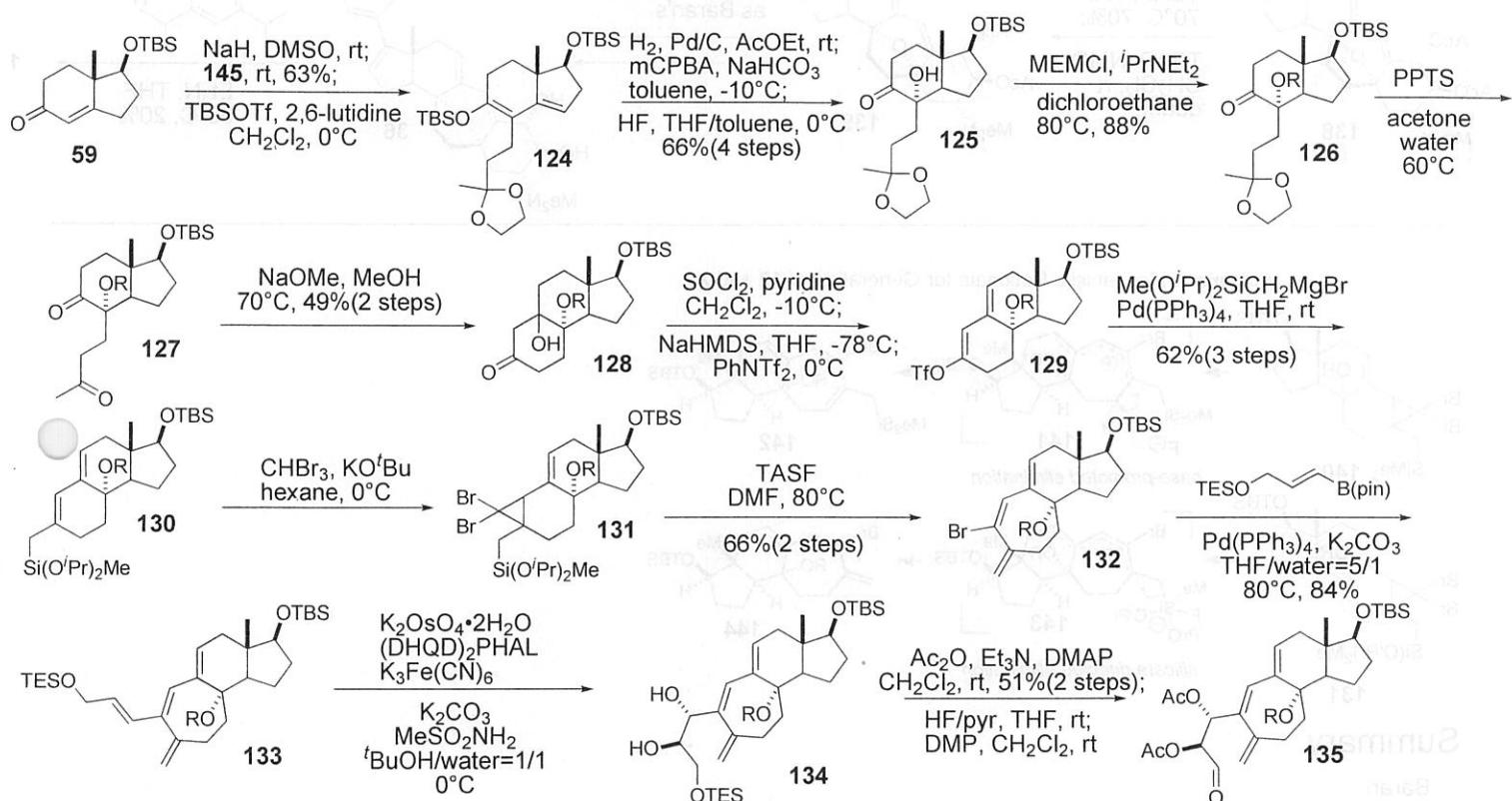


## 4. Total synthesis by shair

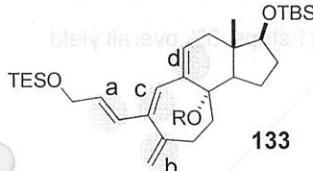
### Retrosynthesis



**Figure 2.** A key step of the cortistatin A synthesis is an aza-Prins/transannular etherification reaction. Enantiomerically enriched Hajos-Parrish ketone is the starting material.



### Regioselectivity of asymmetric dihydroxylation



**Table 27. The Selective Mono-dihydroxylation of Highly Conjugated Systems<sup>117,†</sup>**

Entry	Substrate	Products	Ratio	% ee	% yield
1	1		6 : 1	60	
2	2		4 : 1	92	68
3	3		4 : 1	98	87

<sup>†</sup> The AD reactions were performed under standard conditions<sup>23a</sup> using (DHQD)<sub>2</sub>PHAL and 0.2–1.0 mol % of OsO<sub>4</sub>.

In general, dihydroxylation maintains maximum degree of conjugation.

a, b, d>c

**Table I. Ligand Effects on the Reactivity Hierarchy as a Function of Olefin Substitution Pattern<sup>a</sup>**

OsO <sub>4</sub> alone	OsO <sub>4</sub> + Quinuclidine	OsO <sub>4</sub> + PHAL(DHQD) <sub>2</sub>
><	><	><
14 ± 10% (29)	1200 ± 5% (18)	8800 ± 5% (23)
2.5 ± 10% (5.1)	320 ± 5% (4.8)	4100 ± 5% (11)
1.6 ± 10% (3)	210 ± 5% (3.2)	1400 ± 5% (3.7)
0.58 ± 10% (1.2)	100 ± 5% (1.5)	690 ± 5% (1.8)
0.58 ± 10% (1.2)	73 ± 5% (1.1)	660 ± 5% (1.5)
0.48 ± 10% (1)	66 ± 5% (1)	380 ± 5% (1)

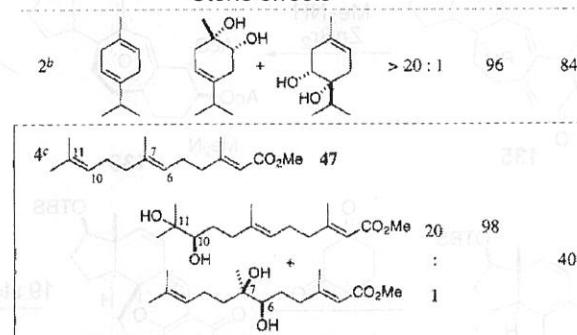
JACS, 1993, 115, 7047

trisubstituted, trans-disubstituted  
V  
cis-disubstituted, 1,1-disubstituted, monosubstituted, tetrasubstituted

a, c, d>b

<sup>a</sup> Rates in M<sup>-1</sup> min<sup>-1</sup>. Relative rates in parentheses. It is important to note that the relative rates only apply within a single column. The absolute rates can, of course, be compared throughout the table.

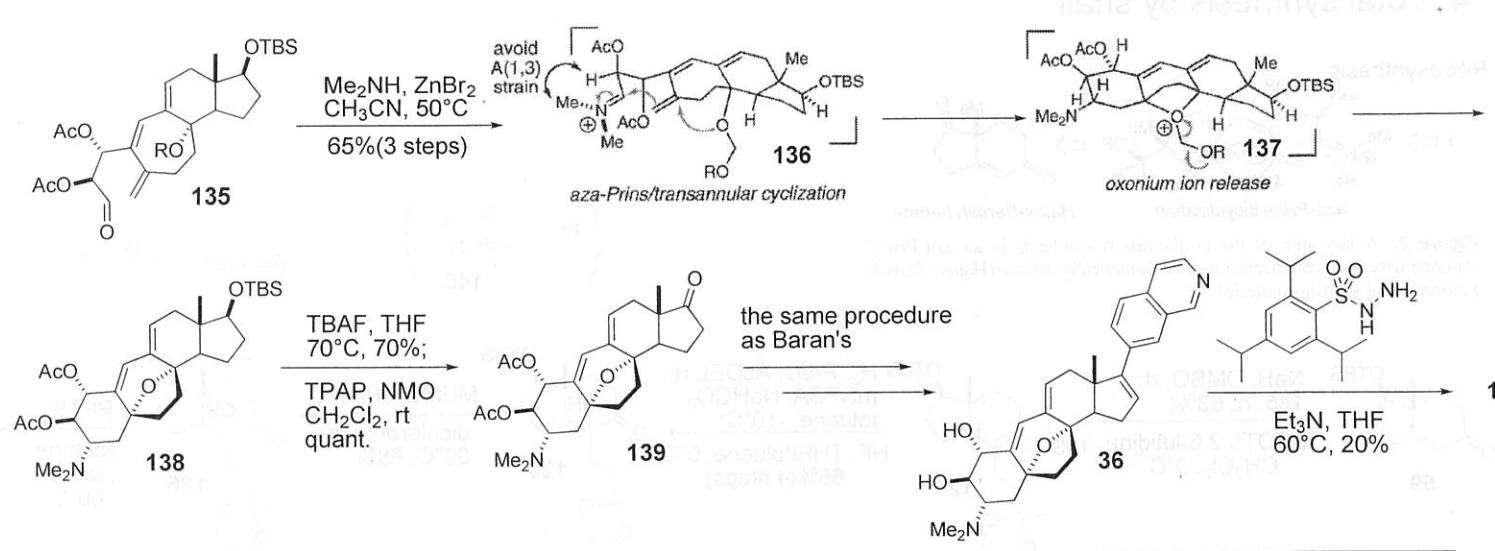
### Steric effects



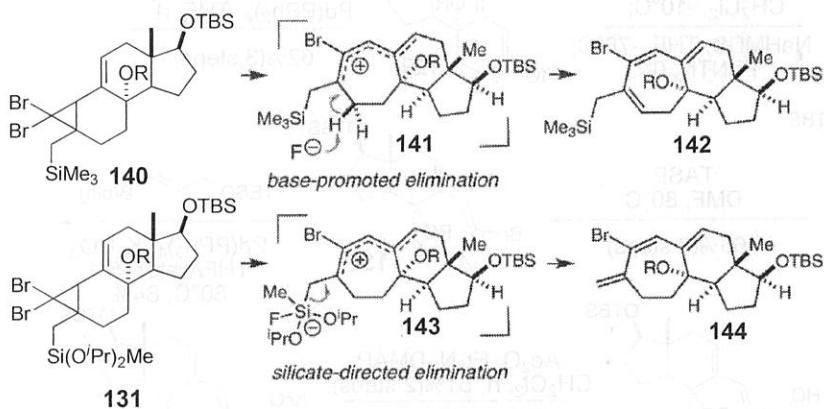
a>d

<sup>†</sup> The AD reactions were performed under standard conditions<sup>23a</sup> using (DHQD)<sub>2</sub>PHAL and 0.2–1.0 mol % of OsO<sub>4</sub>. <sup>a</sup> % ee of the major product. <sup>b</sup> See ref 117. <sup>c</sup> See ref 121.

Chem. Rev. 1994, 94, 2483

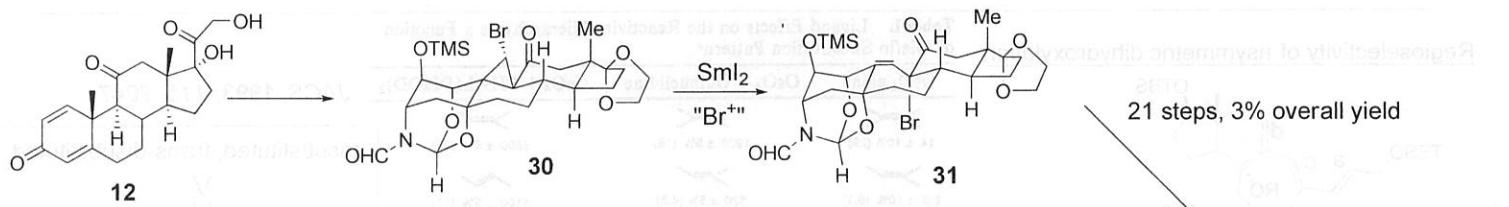


Scheme 3. Mechanistic Rationale for Generation of 12 and 20

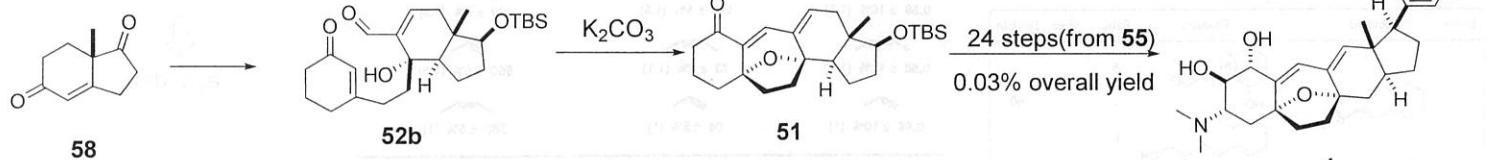


## Summary

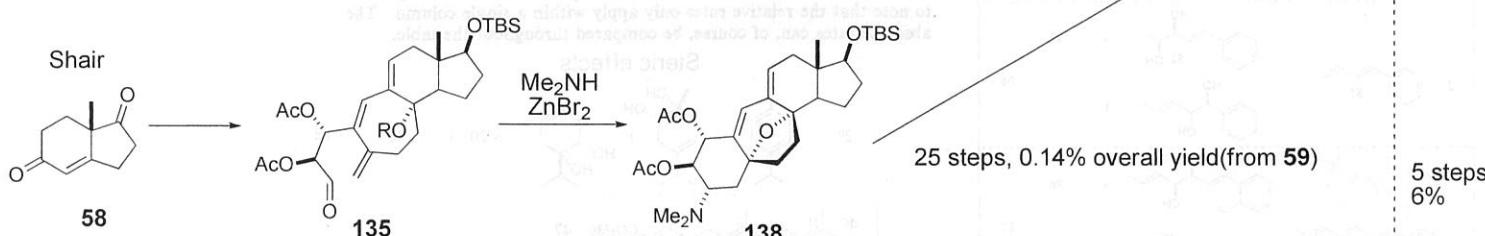
Baran



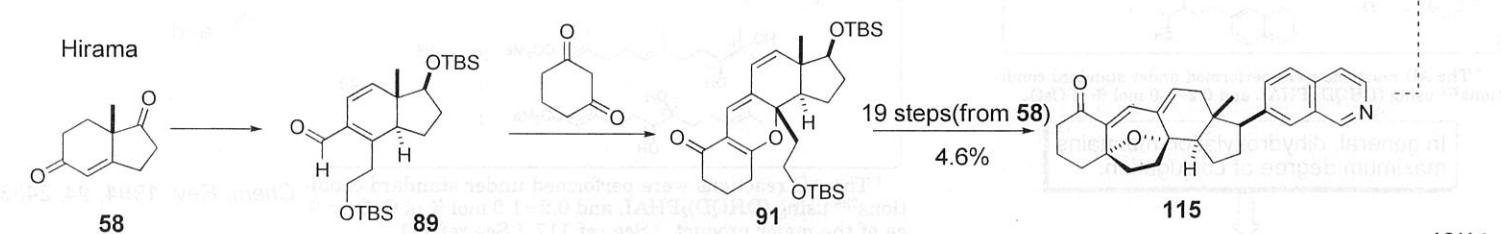
Nicolaou



Shair



Hirama



## 5. Structure-Activity Relationship

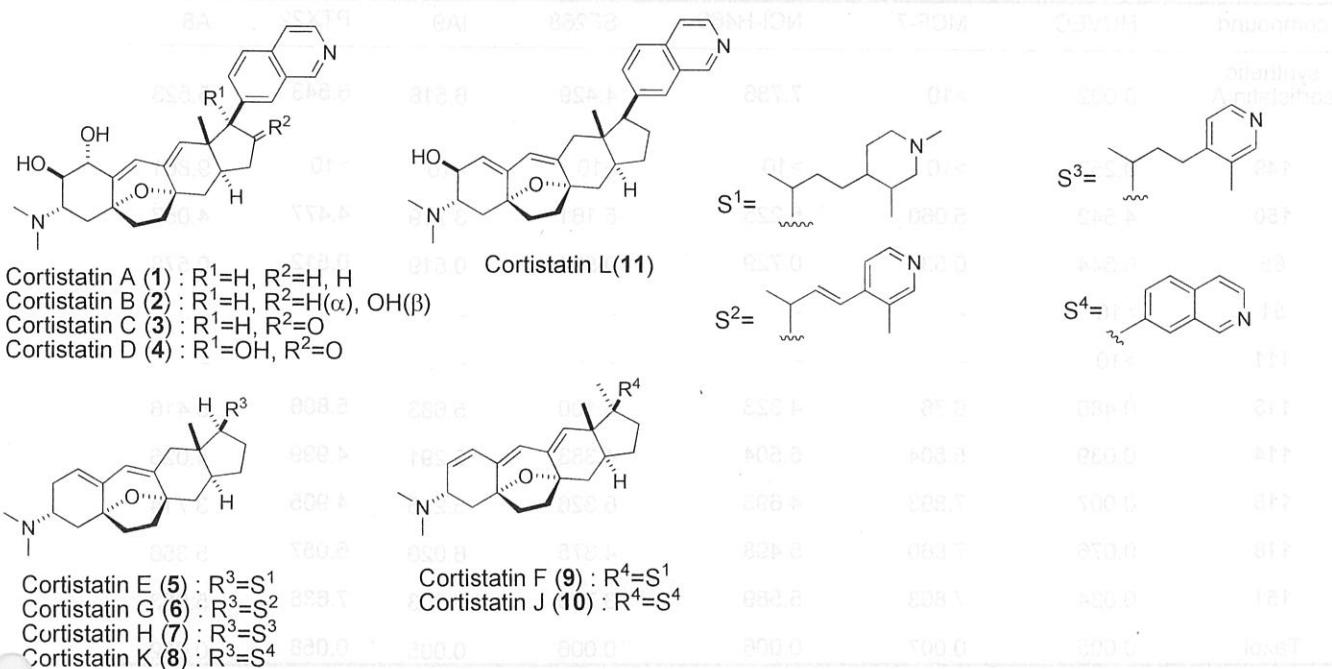


Table 1. Growth inhibition of cortistatins against HUVECs and various type of cell lines

Cell line	A (1)		B (2)		C (3)		D (4)		E (5)														
	IC <sub>50</sub>	SI	IC <sub>50</sub>	SI	IC <sub>50</sub>	SI	IC <sub>50</sub>	SI	IC <sub>50</sub>	SI													
HUVEC	0.0018	1	1.1	1	0.019	1	0.15	1	0.45	1													
KB3-1	7.0	3900	120	110	150	7900	55	460	2.5	6													
Neuro2A	6.0	3300	160	150	180	9500	>300	n.d.	1.9	4													
K562	7.0	3900	200	180	>300	n.d.	>300	n.d.	2.8	6													
NHDF	6.0	3300	>300	n.d.	>300	n.d.	>300	n.d.	1.9	4													
G (6)	IC <sub>50</sub>	SI	H (7)	IC <sub>50</sub>	SI	K (8)	IC <sub>50</sub>	SI	F (9)	IC <sub>50</sub>	SI	J (10)	IC <sub>50</sub>	SI	L (11)	IC <sub>50</sub>	SI						
0.80	1	0.35	1	0.04	1	1.9	1	0.008	1	0.023	1	8.9	11	2.3	7	10.2	250	10.8	6	9.1	1100	14	610
4.0	5	2.2	6	3.0	80	4.0	2	3.3	410	2.8	120	3.8	5	2.7	8	3.9	100	4.0	2	3.3	410	4.3	190
2.9	4	2.7	8	2.5	60	4.1	2	2.4	300	2.4	100												

Kobayashi et al. Bioorg. Med. Chem. 2007, 15, 6758

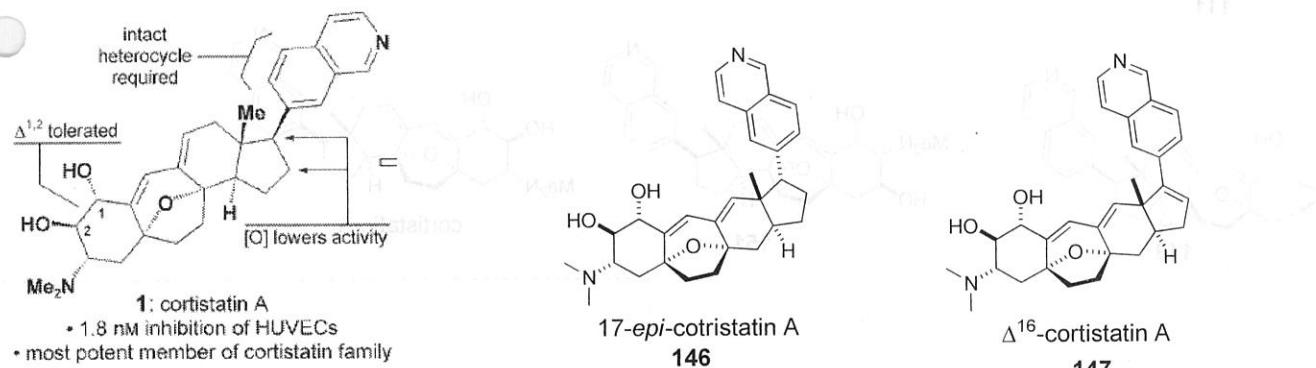


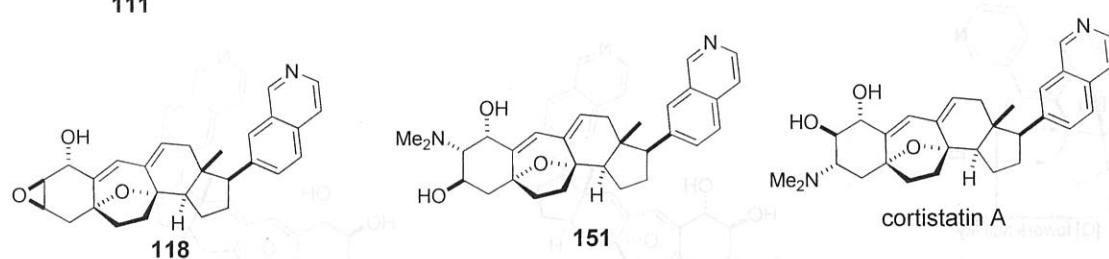
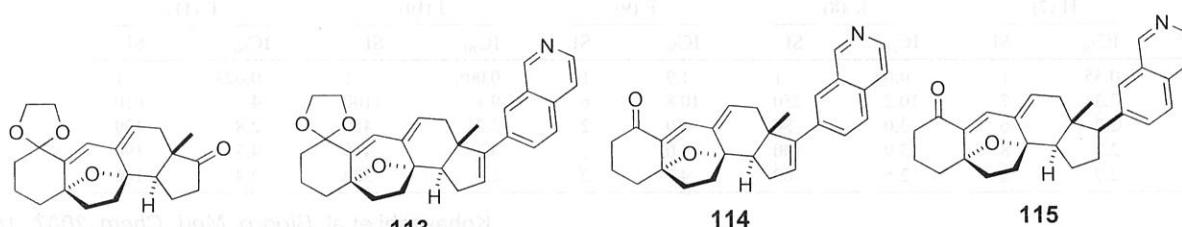
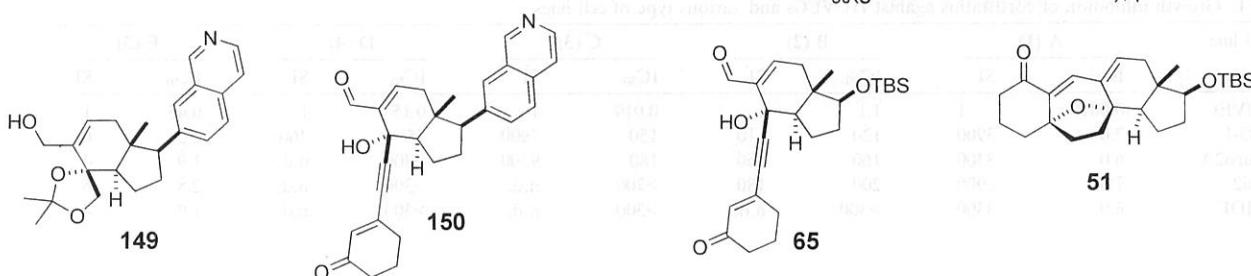
Table 2: Selective growth inhibition of cortistatins against HUVECs.

Substrate	IC <sub>50</sub> [nM]
cortistatin A (1)	2.43 <sup>[a]</sup> , 1.8 <sup>[b]</sup>
$\Delta^{16}$ -cortistatin A (2)	3.88
17-epi-cortistatin A (4)	> 1000
6d-g, 7a-f, 8e, 9a, 9d-e <sup>[c]</sup>	> 1000

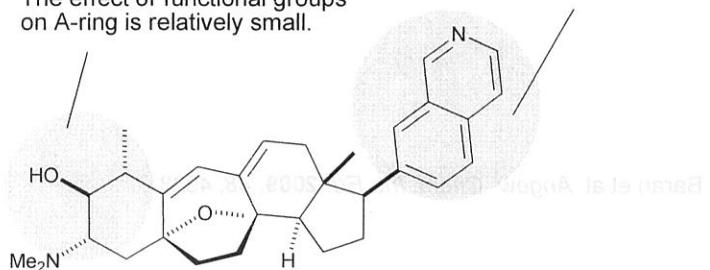
[a] IC<sub>50</sub> of synthetic cortistatin A tested by Pfizer Inc. [b] IC<sub>50</sub> of natural cortistatin A tested by Kobayashi group.<sup>[4a]</sup> [c] The TBS groups were removed prior to testing. The results of 6e and 7e are from Ref. [4f].

Baran et al. Angew. Chem. Int. Ed. 2009, 48, 4328

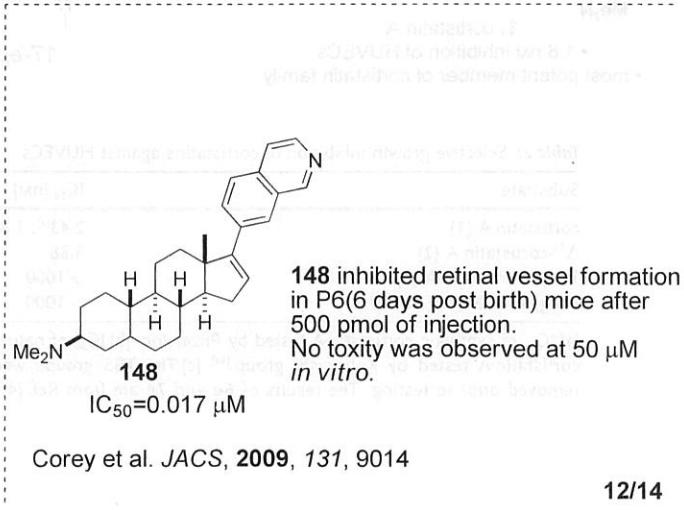
compound	HUVEC	MCF-7	NCI-H460	SF268	IA9	PTX22	A8
synthetic cortistatin A	0.002	>10	7.786	4.429	6.518	6.543	5.523
149	0.253	>10	>10	>10	>10	>10	9.861
150	4.542	5.060	5.225	5.161	3.719	4.477	4.057
65	0.544	0.532	0.729	0.524	0.519	0.512	0.578
51	>10	-	-	-	-	-	-
111	>10	-	-	-	-	-	-
113	0.486	6.76	4.323	5.100	5.683	5.806	5.416
114	0.039	5.504	5.504	4.383	5.291	4.999	4.026
115	0.007	7.893	4.693	6.326	5.236	4.905	3.714
118	0.076	7.860	5.498	4.375	6.020	6.057	5.356
151	0.034	7.803	5.589	3.799	7.303	7.638	5.963
Taxol	0.005	0.007	0.006	0.006	0.005	0.058	0.052

GI<sub>50</sub>(growth inhibition of 50%),  $\mu\text{M}$ 

The effect of functional groups on A-ring is relatively small.



Isoquinoline moiety is essential for the inhibition of cell proliferation. This moiety is also necessary for the selectivity.



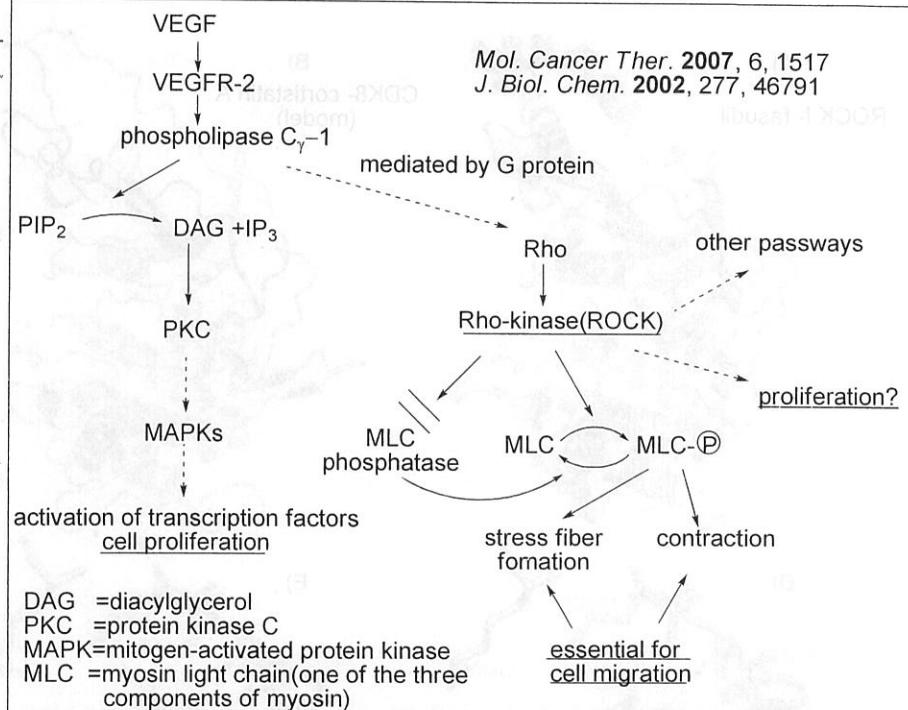
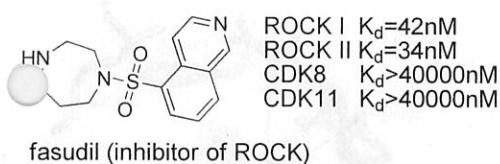
Cortistatin A was tested in high-throughput binding assay against a panel of 402 kinases.

Nicolaou et al. *Angew. Chem., Int. Ed.* 2009, 48, 8952

**Table 1:** Kinase affinity of synthetic cortistatin A.

Kinase	POC (10 $\mu$ M) <sup>[a]</sup>	$K_d$ [nM] <sup>[b]</sup>
ROCK II	0	220 $\pm$ 7
CDK11	0.1	10 $\pm$ 2
CDK8	0.95	17 $\pm$ 2
LTK	2.9	ND
ALK	4.4	ND
PIM2	4.4	ND
PKAC $\alpha$	8.7	3500 $\pm$ 212
PKAC $\beta$	13	ND
MET	18	ND
PRKG2	21	ND
RIOK2	21	ND
ROCK I	21	250 $\pm$ 35
CLK4	26	ND
ROS1	26	ND
CIT	28	ND
JNK1	29	ND

[a] Kinases with POC < 35 are shown. [b] Average of two determinations  $\pm$  SD; ND = not determined. POC = percent of control

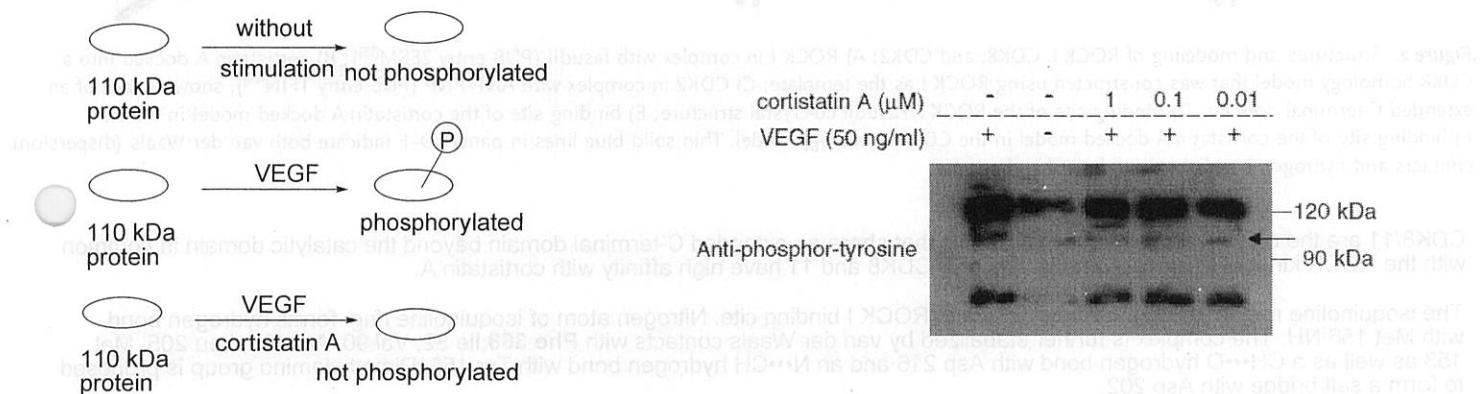


## ROCK is a target protein of cortistatins?

Kobayashi et al. reported in 2007 that cortistatin A inhibited cell proliferation **without inhibition of MAPKs**

→ Another pathway is the target.

They also reported that phosphorylation of **110 kDa protein** was reduced remarkably by the treatment with cortistatin A.



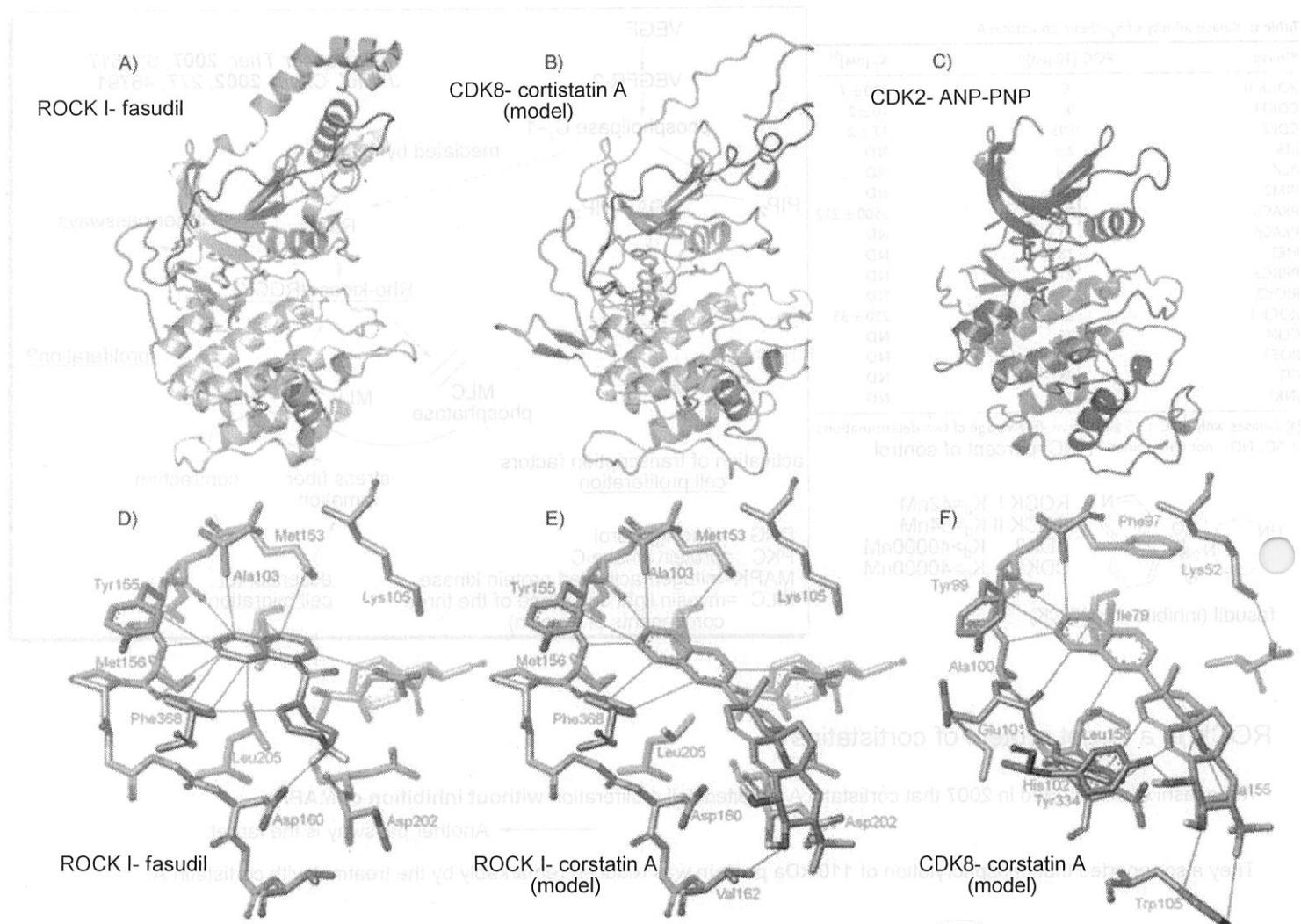
ROCK phosphorylates **110 kDa subunit of MLC phosphatase** to promote cell migration.

→ ROCK seems to be a reasonable target.

( Further investigation is necessary to confirm the conclusion. )

Because the function of CDK family is not known very well, it's difficult to ascertain whether the effects of cortistatin A are consistent with inhibition of CDK8/11.

?

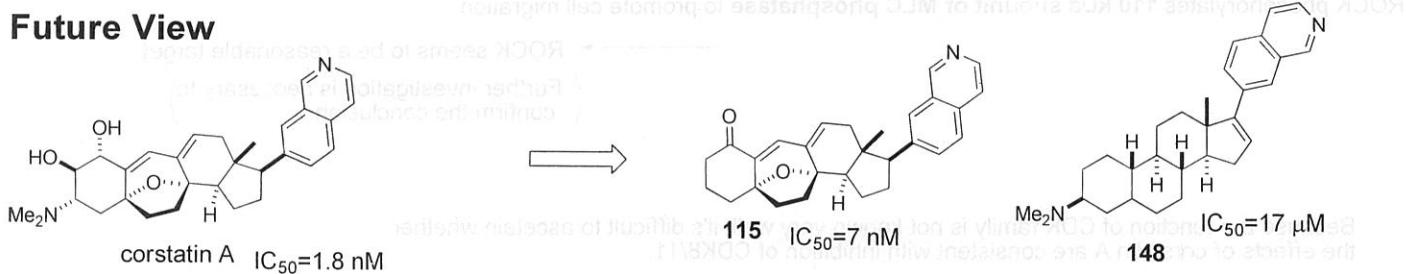


**Figure 2.** Structures and modeling of ROCK I, CDK8, and CDK2: A) ROCK I in complex with fasudil (PDB entry 2ESM<sup>[19]</sup>); B) cortistatin A docked into a CDK8 homology model that was constructed using ROCK I as the template; C) CDK2 in complex with ANP-PNP (PDB entry 1FIN<sup>[20]</sup>), showing lack of an extended C-terminal domain; D) binding site of the ROCK I/Fasudil co-crystal structure; E) binding site of the cortistatin A docked model in ROCK I; F) binding site of the cortistatin A docked model in the CDK8 homology model. Thin solid blue lines in panels D–F indicate both van der Waals (dispersion) contacts and hydrogen bonds between ligand and protein.

CDK8/11 are the only members of the CDK family that share an extended C-terminal domain beyond the catalytic domain in common with the ROCK kinases. This is probably why only CDK8 and 11 have high affinity with cortistatin A.

The isoquinoline ring of cortistatin A interacts with ROCK I binding site. Nitrogen atom of isoquinoline ring forms hydrogen bond with Met 156 NH. The complex is further stabilized by van der Waals contacts with Phe 368, Ile 82, Val 90, Ala 103, Leu 205, Met 153 as well as a CH...O hydrogen bond with Asp 216 and an N...CH hydrogen bond with Tyr 155. Dimethylamino group is proposed to form a salt bridge with Asp 202.

## Future View



Although the synthesis of corstain A was accomplished, only tiny amount of cortistatin A was obtained so far. Further investigation might be difficult for this reason.

These less complex molecules will probably be a target for further investigation and lead compound for a new drug.